





Phytochemical composition and bioactivities of hawthorn (*Crataegus* spp.): review of recent research advances

Petras Rimantas Venskutonis*

Department of Food Science and Technology, Kaunas University of Technology, Radvilénų rd. 19, Kaunas, LT-50254, Lithuania *Corresponding author: Petras Rimantas Venskutonis, Department of Food Science and Technology, Kaunas University of Technology, Radvilénų rd. 19, Kaunas, LT-50254, Lithuania. E-mail: rimas.venskutonis@ktu.lt DOI: 10.31665/JFB.2018.4163

Received: December 04, 2018; Revised received & accepted: December 23, 2018

Abbreviations: ABTS, 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid; AChE, acethylcholinesterase; AMP, adenosine monophosphate; BchE, butyrylcholinesterase; CPT-1, carnitine palmitoyl transferase; DPPH*, stable 2,2-diphenyl-1-picrylhydrazyl radical; DW, dry weight; eNOS, endothelial NO synthase; FRAP, ferric reducing antioxidant power; GAE, gallic acid equivalents; Gpx3, glutathione peroxidase 3; HDLC, high density lipoprotein cholesterol; HFD, high-fat diet; HLF, hawthorn leaf flavonoids; ICAM-1, intercellular adhesion molecule; IL, interleukin; LDLC, low density lipoprotein cholesterols; LPS, lipopolysaccharide; MDA, malondialdehyde; NAFLD, nonalcoholic fatty liver disease; NO, nitric oxide; ORAC, oxygen radical absorbance capacity; PKC-α, protein kinase C α; PPARα, peroxisome proliferator activated receptor α; RAW264.7, murine macrophage cell line; RCT, reverse cholesterol transport; ROS, reactive oxygen species; SOD, superoxide dismutase; SREBP-1c, sterol regulatory element binding protein-1c; TBARS, thiobarbituric acid reactive substances; TC, total cholesterol; TE, trolox equivalents; TFC, total flavonoid content; TG, triacylglycerols; TNF-α, necrosis factor-α; TPC, total phenolic content; VLDLC, very low density lipoprotein cholesterol.

Citation: Venskutonis, P.R. (2018). Phytochemical composition and bioactivities of hawthorn (*Crataegus* spp.): review of recent research advances. J. Food Bioact. 4: 69–87.

Abstract

Hawthorn (*Crataegus* spp.) is one of the most famous plants which has been used as natural medicine and nutraceutical. Its phytochemical composition, bioactive compounds and health benefits have been intensively studied and hawthorn preparations may be recognized as classical natural products for cardiovascular health. Polyphenolic compounds of different hawthorn anatomical parts as well as their extracts have been the focus of a majority of these studies, although various other classes of natural health promoting constituents have also been isolated, identified and characterized. Regardless, numerous published reports have particularly focused on the activity mechanisms which are very important for supporting various health benefits. This review summarizes the most recent studies on hawthorn, mainly published since 2015. Search of different databases indicates that approximately 200 publications, which are relevant to phytochemistry and health benefits of *Crataegus* spp., have appeared since then, most of them have not been included in the existing reviews.

Keywords: Hawthorn (Crataegus spp.); Phytochemical composition; Bioactivities; Health benefits; Toxicity.

1. Introduction

Hawthorns are large genus of small shrubs and trees belonging to Rosaceae family, Amygdaloideae subfamily, Maleae tribe, Malinae subtribe *Crataegus* Tourn. *ex* L. genus. Nowadays it is agreed that the genus is represented by approximately 200 species which are native to temperate regions of the Northern Hemisphere in North America, Europe and Asia (Phipps et al., 2003). The name "hawthorn" is often used in some countries although originally it was applied to the species native to northern Europe, especially to the common hawthorn (*C. monogyna*). Red or black small pome fruits of some *Crataegus* spp. are edible, e.g. fresh fruits of *C. orientalis*, while the fruits of other species become eatable in late autumn. Their flavor has been compared to over-ripe apples and they are mainly used for jellies or homemade wine. Picked in spring young leaves are also edible and are tender enough to be used in salads. In Europe, the fruit, leaves, and flowers were traditionally employed for treating heart problems due to their antispasmodic, cardiotonic, hypotensive, and antiatherosclerotic effects. Nowadays, hawthorn preparations are mainly used as cardioprotective agents (Fong and Bauman, 2002); however, in traditional medicine it remains as a therapeutic agent for many other diseases including cancer, diabetes, cough, flu, asthma, stomach ache, rheumatic pain, nephritis, and hemorrhoids.

Among plant species *C. pinnatifida* (ten varieties of Chinese hawthorns) have long been used in traditional Chinese and European herbal medicines, and are widely consumed in the form of juice, drink, jam and canned fruit, while *C. monogyna* is commonly cultivated in the Mediterranean countries (Calişkan, 2015). Chinese hawthorns comprise 18 species, however currently only *C. pinnatifida* and *C. pinnatifida* var. *major* fruits are included in the Chinese Pharmacopoeia. *C. pinnatifida* and *C. scabrifolia* fruits have also been traditionally used as peptic agents in oriental medicine and recently in a various local sweet foods, mainly soft drinks, jams, juices, tinned foods, and wines (Jurikova et al., 2012).

Several reviews summarizing numerous research findings on hawthorns are given in Table 1. Composition and health effects of phenolic compounds in Crataegus spp. of different origins was reviewed by Yang and Liu (2012), while Edwards et al. (2012) focused their review on the chemistry of the genus. Effects of bioactive natural products of Crataegus spp. on the vascular endothelium were also reviewed (Ahmad et al., 2013). More recently, polyphenolic composition and medical applications of C. monogyna were reviewed (Nabavi et al., 2015) by summarizing the growing evidence on various interesting physiological and pharmacological activities of this species due to the presence of different bioactive natural compounds. Nunes et al. (2017) reviewed the labeling and safety concerns of herbal products containing Crataegus spp. among other ingredients. Rastogi et al. (2016) provided a review on the cardiovascular effects of C. oxyacantha, including ischemic heart disease, congestive heart failure, arrhythmias and hypertension and concluded that, although the mechanisms of action are not very clear, there is enough evidence of their efficacy in various cardiovascular disorders.

C. oxyacantha is a widely used Chinese herb for treating gastrointestinal ailments and heart problems; it is also consumed as a food. In North America, the role of treating heart problems dates back to the 1800's. Currently, the evidence is accumulating from various *in vivo* and *in vitro* studies that hawthorn extracts exert a wide range of cardiovascular and pharmacological properties, including antioxidant activity, positive inotropic, antiinflammatory, anticardiac remodeling, antiplatelet aggregation, vasodilating, and endothelial protective effects, as well as reduction of smooth muscle cell migration and proliferation, protection against ischemia/reperfusion injury, antiarrhythmic, lipidlowering and arterial blood pressure decreasing effects (Wang et al., 2013).

Crataegus spp. were also reviewed together with other botanicals (Chen et al., 2009; Schmitt and Dirsch, 2009; Rastogi et al., 2016; Dennehy, 2001; Bjorklund et al., 2018), Chinese Traditional Medicine plants (Xie et al., 2012; Liu and Huang, 2016; Dong et al., 2017), various herbal products (Ahmad et al., 2013; Nunes et al., 2017), nutraceuticals (Houston, 2005) and natural antioxidants (Zhao, 2005).

2. Bioactive constituents and antioxidant/antiinflammatory activities

The variations in the composition of different Crataegus spp. from various growing regions, as well as of different anatomical parts of hawthorn have been widely studied. Acids, sugars and sugar alcohols, minerals, vitamins, and amino acid composition were among the analyzed constituents, however, overwhelming number of published articles have been focusing on polyphenolic compounds, as the most important hawthorn bioactive phytochemicals which may provide health benefits to humans. Consequently, many hawthornbased preparations are standardized according to the content of their flavonoids and phenolic acids. The interest in phenolic and polyphenolic compounds particularly increased in the era of functional foods (Shahidi, 2004). Besides strong antioxidant activity, phenolic compounds have demonstrated numerous protective effects against chronic diseases, which have recently been reviewed (Shahidi and Yeo, 2018). This section will review only the most recent publications, which report new findings in the hawthorn phytochemistry. In general, the studies were performed for the isolation and purification of flavonoids, other groups of compounds. as well as determination of their variations among different origins and other variables.

Oxidative stress, which reflects an imbalance between the systemic manifestations of reactive oxygen species (ROS), may induce development of various diseases and disorders and, although living cells possess strong endogenous antioxidant system, it is believed that under some conditions ROS-protective function might be insufficient, while exogenous dietary antioxidants could assist in mitigating inflammatory processes and thereby restoring homeostatic state in the organism. In addition, natural antioxidants have attracted an increasing attention as possible alternatives to synthetic additives in foods and cosmetics. Hawthorns are well-known as an excellent source of natural antioxidants as reviewed in the literature (Venskutonis, 2016). The majority of such studies applied simple and easy spectrophotometric in vitro and in situ assays such as DPPH[•] and ABTS^{•+} scavenging, FRAP, Folin-Ciocalteau's reaction (most often considered as TPC), ORAC, β-carotene-linoleic acid co-oxidation, TBARS and others. Although there is an opinion that such in vitro antioxidant capacity values are not appropriate, the experts in this area recently concluded that they, as lowcost and high-throughput tools, cannot be ignored (Granato et al., 2018). Therefore, together with phytochemical information this section will also briefly reviews the most recent findings in antioxidant properties of hawthorn products.

Phytochemical studies of the most widely used C. pinnatifida (Table 2) resulted in separation and purification of 4 new monoterpene glycosides and a new phenolic glycoside in addition to the 10 previously known ones (Li et al., 2015). Furthermore, 15 triterpenoids, including 4 novel acids, in hawthorn berries (Qiao et al., 2015); 2 norditerpenoids with unique carbon skeletons, 4 sesquiterpenoids and 9 nor-sesquiterpenoids from the ethanolic extract of plant leaves (Gao et al., 2017); 8 new phenylpropanoids (crataegusoids A-F) from the fruits (Guo et al., 2018); 8 new lignans (hawthornnins A-H) and 7 known analogues (Huang et al., 2015a); 2 new 8-O-4'-neolignans, huangnin A and B and four known analogs (Huang et al., 2015b); 7 new lignans (crataegusnins A-G) and 5 known compounds from the seeds (Peng et al., 2016) and from crude Crataegus Fructus drug (Kazuma et al., 2016), while 7 known main constituents were separated from the flavonoid fraction of leaves using a combination of high-speed counter-current chromatography (HSCCC) coupled with pre-HPLC (Wen et al., 2017). Crataegusins A and B, new flavanocoumarins, also showed

Plants or their products	Торіс	Summary of content	Reference
Standardized Crataegus extracts	Treatment of heart failure	Positive inotropic, chronotropic, dromotropic effects; negative bathmotropic effects; increases coronary and myocardial perfusion, lowers peripheral resistance, has antiarrhythmic and economizing action with respect to oxygen and energy consumption.	Loew, 1997
Extract from fresh hawthorn berries	Orthostatic hypotension	In combination with D-camphor exerts a significant effect that counteracts an orthostatic fall in blood pressure	Belz and Loew, 2003
Hawthorn extract	Clinical trials to treat chronic heart failure	A significant benefit from hawthorn extract as an adjunctive treatment for chronic heart failure	Pittler et al., 2003
Hawthorn	Cardiovascular disease	May induce anti-ischemia/reperfusion-injury, anti-arrhythmic, hypolipidemic and hypotensive effects, which may in part be due to the presence of antioxidant flavonoid components.	Chang et al., 2005
Hawthorn medicinal extracts (WS 1442, LI 132)	Heart failure	Improvement in patients with mild forms of heart failure, clinical symptoms, pressure-heart rate product, left ventricular ejection fraction; no evidence of a notable reduction in mortality or sudden death	Dahmer and Scott, 2010
Standardized extracts from hawthorn leaves and flowers	Preclinical and clinical studies mild chronic heart failure	Cardiotonic effects, cardio- and vasoprotective properties; may be employed in the prophylactic and therapeutic treatment of endothelial dysfunction, atherosclerosis, coronary heart disease, or prevention of restenosis/reocclusion following peripheral endo-vascular treatment.	Koch and Malek, 2011
C. oxyacantha	Cardiovascular disease prevention	A wide range of cardiovascular pharmacological properties: antioxidant activity, positive inotropic effect, anti-inflammatory effect, anticardiac remodeling effect, antiplatelet aggregation effect, vasodilating effect, endothelial protective effect, reduction of smooth muscle cell migration and proliferation, protective effect against ischemia/reperfusion injury, antiarrhythmic effect, lipid-lowering effect and decrease of arterial blood pressure effect	Wang et al., 2013
C. pinnatifida	Chemical constituents, pharmacology, potential applications	>150 compounds (flavonoids, triterpenoids, steroids, monoterpenoids, sesquiterpenoids, lignans, hydroxycinnamic acids, organic acids and nitrogen- containing) have been isolated and identified; broad pharmacological effects with low toxicity; wide applications in pharmacological therapy	Wu et al., 2014
The genus <i>Crataegus</i>	Chemical, pharmacological, health, uses aspects	Heart (cardiovascular disorders), central nervous system, immune system, eyes, reproductive system, liver, kidney etc; cytotoxic, gastroprotective, anti-inflammatory, anti-HIV and antimicrobial activities. Bioactive phytochemicals: oligomeric procyanidins, flavonoids, triterpenes, polysaccharides, catecholamines; traditional uses; the clinical trials and regulatory status.	Kumar et al., 2012
<i>Crataegus</i> spp. of different origins	Composition and health effects of phenolic compounds	Epicatechin, aglycons and glycosides of B-type oligomeric procyanidins and flavonols, phenolic acids and C-glycosyl flavones as he major groups of phenolics; in vitro and animal studies showing cardioprotective, hypolipidaemic, hypotensive, antioxidant, radical-scavenging and anti-inflammatory potentials	Yang and Liu, 2012

Table 1. Published review articles on Crataegus spp. bioactive compounds, pharmacological and health effects and safety

Plants or their products	Торіс	Summary of content	Reference
C. oxycantha	Treatment of atrial fibrillation	Mechanisms of cardiovascular health benefits; lack of clinical studies evaluating its use in atrial fibrillation	Kanman-thareddy et al., 2015
<i>C. monogyna</i> Jacq.	Polyphenolic composition and medical applications	A critical review about physiological and pharmacological activities due to the presence of different bioactive natural compounds. In addition, scientific evidence suggests that the toxicity of hawthorn is negligible	Nabavi et al., 2015
Hawthorn juice	Various aspects	Nutritional characteristics, bioactives and antioxidant efficacy, phenolics, health effects, novel products/formulations and future trends	Venskutonis, 2016
Crataegus extract WS 1442	Benefit-risk assessment	Positive inotropic and antiarrhythmic properties; protecting from ischemic damage, reperfusion injury, and hypertension-related hypertrophy; improving endothelial functions (NO synthesis, delay of endothelial senescence); favorable safety profile (monotherapy and as add-on therapy); no drug interactions; no specific adverse reactions	Holubarsch et al., 2018
WS 1442 extract from <i>Crataegus</i> spp. leaves and flowers with 18.75% oligomeric procyanidins	Effects on cardiovascular system in vitro and in vivo, inc. large clinical trials	Beneficial cardioprotective values; free radicals scavenger; protect the ischemic heart tissue from neutrophile elastase action successions; vasorelaxant activity, via affecting eNOS synthase, and prevents ischemic heart tissue swelling by influence on calcium signaling pathways, and thus detain hyperpermeability of endothelium.	Zorniak et al., 2017
Crataegus spp.	Phenolic compounds; their bioactivity	Beneficial effects and the mechanisms of action involved are analyzed in a critical and systematic way in order to promote its use in the treatment of various diseases considered in Mexico as public health problems.	Cervantes-Paz et al., 2018
C. meyeri and C. pontica	Antioxidant properties and medicinal uses	The advantages of using natural remedies over their synthetic equivalents, the necessity of thorough investigations of less studied <i>Crataegus</i> spp	Dolatkhani and Jameie, 2015

Table 1. Published review articles on Crataegus spp. bioactive compounds, pharmacological and health effects and safety - (continued)

a significant DPPH' reducing activity compared with epicatechin (Kazuma et al., 2016). Most of the compounds isolated from the 70% ethanolic extract of C. pinnatifida seeds showed moderate DPPH' scavenging activity and significant activities in the FRAP and ABTS assays. Furthermore, 6 compounds exhibited marked NO inhibition, 4 of them had a potent TNF-α inhibitory effect indicating that hawthorn seeds can be regarded as a potential new and cheap source of antioxidants and inflammation inhibitors (Peng et al., 2016). From C. pinnatifida, the isolated lignans demonstrated DPPH*/ABTS*+ scavenging efficacy, while some of them had anti-inflammatory activities, which was assessed by detecting the NO and TNF-α production in the LPS-induced RAW264.7 cells (Huang et al., 2015a). The fractions isolated with different polarity solvents from the crude 70% aqueous acetone extract of wood and bark of C. pinnatifida demonstrated antioxidative and anti-inflammatory activities. Thus, the n-butanol and ethyl acetate fractions of wood and bark, respectively, exhibited the strongest antioxidant effects, while all moisture-free fractions of wood showed high inhibitory effect on NO production (Bae and Kim, 2017).

Seasonal variations (May–October) in the yield of 7 phenolic compounds, the TPC in *C. pinnatifida* leaves, roots, twigs, and fruits were positively correlated with the temperature and length

of daytime, while most remarkable radical scavenging and reducing power was determined for the leaves harvested in September (Luo et al., 2016).

Several articles have reported phytochemical studies of C. azarolus (Table 2). Thus, ethanolic extract of C. azarolus var. euazarolus Maire leaves was fractionated with different solvents and it was found that ethanol extract (70%) showed the highest DPPH. scavenging activity compared to methanol, ethyl acetate and acetone extracts (Abu-Gharbieh and Shehab, 2017). A new ursanetype triterpene acid, named azarolic acid, and 8 known compounds were isolated from the crude ethyl acetate extract of C. azarolus var. aronia (Mahmud et al., 2016). Phytochemical composition of 20 phenolic compounds quantified in C. azarolus and C. monogyna fruits were compared (Mraihi et al., 2015). Methanol and ethanol extracts of C. azarolus exerted substantial antioxidant, anti-inflammatory, and antiproliferative capacities, which were evaluated by measuring the secreted amounts of the proinflammatory mediator prostaglandin E2 (PGE2), and by assaying the mRNA levels of the proinflammatory cytokines (IL- α , IL- β , and Il-6), chemokines (CCL3 and CCL4) and inflammation-sensitive COX2 and iNOS enzymes (Kallassy et al., 2017).

Dibenzo-p-furan derivatives and ursolic acid were isolated from

Table 2. Phytoconstituents of <i>Crataegus</i> spp. (recently reported; newly identified compounds are in itali	Table 2.	Phytoconstituents of	Crataegus spp.	(recently reported;	newly identified	compounds ar	e in italics
---	----------	----------------------	----------------	---------------------	------------------	--------------	--------------

Sample information	Name	Bioactivities, other study objective	Reference
C. pinnatifida, extract of leaves	4 monoterpene glycosides, <i>pinnatifidanosides</i> <i>A-D; phenolic glycoside, pinnatifidanoside E;</i> byzantionoside B, (3S,5R,6R,7E,9R)-3,6-epoxy-7- megastigmen-5, 9-diol-9- <i>O</i> -β-D-glcp, (65,7Z,9R)- roseoside, icariside B6, linalool oxide β-D-glc, shanyenoside A, dihydrocharcone-2'-β-D-glc, eriodectyol, vitexin, 2"- <i>O</i> -rhamnosyl vitexin	Antithrombotic effects	Li et al., 2015
<i>C. pinnatifida</i> , ethanolic extract of leaves	Norhawthornoids A, B; sesquiterpenoids shnyegenin B, shnyeside B, (3S,5R,6R,7E,9S)- megastiman-7-ene-3,5,6,9-tetrol, euodionosides D, (6R,9R)-3-oxo-α-ionol-9-O-β-D-glucopyranoside, (6S,7E,9R)-6,9-dihydroxy-4,7-megastiymadien-3- one-9-O-[β-D-xylopyranosy-β-D-glucopyranoside], linarionosides A, B, C; 3,9-dihydroxy-5- megastigmen-3-O-[β-D-xylopyranosy-β-D- glucopyranoside], pinnatifidanosides C, F, G.	Antithrombotic effects	Gao et al., 2017
C. pinnatifida, hydroethanolic extract of fruits	Crataegusoids: A(–), B, C, D, E (–) and (+), F	Cytotoxicity against cancer cells	Guo et al., 2018
C. pinnatifida, extract of seeds	8 new lignans, <i>hawthornnins</i> A-H and 7 known analogues	Antioxidant and anti-inflammatory	Huang et al., 2015a
C. pinnatifida, extract of seeds	2 new 8-0-4' neolignans, huangnin A and B; 4 known analogs	Tyrosinase inhibition	Huang et al., 2015b
C. pinnatifida, 70% ethanol extract of seeds	7 Crataegusnins A-G; 3 substituted propanetriols (3); leptolepisol D	Antioxidant and anti-inflammatory	Peng et al., 2016
Crataegus spp., 70% ethanol extract from seeds	(7R, 8R, 8S)-, (7'S, 8'R, 8R)-, (7'R, 8'S, 8S)- isolariciresinols, (7'S, 8'R, 8R)-lyoniresinol, (7'S, 8'R, 8R)-isolariciresinol-9'-6-D-glc, lyoniside, nudiposide	Inhibition of amyloid $A_{\beta 1}$ –42 aggregation	Huang et al., 2018b
C. pinnatifida, flavonoid extract of leaves	(-)-Epicatechin, quercetin-3-O-(2,6-di-α-L- rhamnopyranosyl)-β-D-galactopyranoside, 4"-O-glucosyl and 2"-O-rhamnosylvitexins, vitexin, hyperoside and isoquercitrin	Isolation and purification of flavonoids	Wen et al., 2017b
<i>C. pinnatifida</i> var. <i>major,</i> crude Crataegus Fructus drug	Crataegusins A and B (2) (new flavanocoumarins)	DPPH reducing activity	Kazuma et al., 2016
<i>C. azarolus</i> var. <i>eu-azarolus</i> Maire, ethanol extract of leaves and its fractions	EtOH: rutin, salicylic and ellagic acids; chloroform and n-butanol fractions: ursolic, 3-β-O-acetyl ursolic, and ellagic acids, quercetin-3-O-β methyl ether, rutin and apigenin7-O-rutinoside	Anti-hyperglycemic activity	Abu-Gharbieh Shehab, 2017
<i>C. azarolus</i> var. <i>aronia</i> , ethyl acetate extract of leaves	A new ursane-type triterpene acid, azarolic acid, 4 known phenolic compounds; 4 known triterpene acids	Anti-vasoconstriction	Mahmud et al., 2016
C. monogyna, C. azarolus fruit	3 hydroxycinnamic and 1 hydroxybenzoic acid, 6 glucosylated flavonols and 2 flavones, 2 cyanidin glycosides; (-)-epicatechin, a dimer B2, two trimers, C1 and C2.	Phytochemical characterization	Mraihi et al., 2015
C. pycnoloba, total extract	4 dibenzofurans inc. newly discovered compound 6-hydroxy-2,3,4- trimethoxydibenzofuran; ursolic aldehyde.	Melanin synthesis inhibition	Agalou et al., 2018
C. oxyacantha, shade dried plant twigs	2-(3, 4-dimethoxyphenyl)-2-methoxyethanol, 3-hydroxy-1-(4-hydroxy-3-methoxyphenyl propan- 1-one, β-sitosterol-3-O-β-D-glc, lupeol, β-sitosterol, betulin, betulinic and oleanolic acids, chrysin (9);	Inhibition of acetyl and butyryl-cholinesterases	Ali, et al., 2017
Crataegus spp. from Bosnia, leaves with flowers, and berries	In mg/g DW: gallic acid (0.001–0.082), chlorogenic acid (0.19–8.70, rutin (0.03 to 13.49).	Phytochemical characterization	Čulum et al., 2018

Sample information	Name	Bioactivities, other study objective	Reference
<i>C. pubescens</i> , Fruit pulp (from Mexico)	In mg/100 mg DW: (+)-catechin (9.17±0.20), (-)-epicatechin (4.32±0.11), chlorogenic acid (5.60±0.24 mg/100); total proanthocyanidins 84.6±1.4 mg cyanidin; total flavonoids 55.89±1.43 mg quercetin.	Phytochemical characterization	González-Jiménez et al., 2018
<i>C. microphylla</i> Koch. ssp. <i>malyana</i> K. I. Chr. & Janjic, extracts of leaves with flowers	In mg/g DW: gallic acid 0.04, caffeic acid0.60, and hyperoside 2.61; TPC: 2.47 to 13.35 GAE; TFC: 0.01–1.09 QE	Phytochemical characterization	Čopra-Janićijević et al., 2018
Various <i>Crataegus</i> spp., flowers and leaves of 56 samples from Iran	TPC: 7.21–87.73 mg GAE/g DW; TFC: 2.27–17.40 mg/g DW; chlorogenic acid, vitexin-2-O-rhamnoside, vitexin, rutin, hyperoside, quercetin, isoquercetin	Flavonoids profile, antioxidant activity	Alirezalu et al., 2018
C. almaatensis Pojark, leaves, flowers and fruits	22 secondary metabolites (flavonoids and phenolic acids); TPC: 218 mg/g	Copmparison with C. oxyacantha	Bekbolatova et al., 2018
<i>C. pinnatifida,</i> commercial berries	15 triterpenoids, inc. 4 novel hydroxy-olean- 12-en-28-oic (HOA) acids: 3-β,6 β,18 β-triHOA, 3 β,6 β,18 β,23-tetra HOA, 2 α,3 β,6 β,18 β-HOA, 2 α,3 β,6 β,18 β,23-pentaHOA	Antiproliferative and antioxidant activity	Qiao et al., 2015
Fruits, methanol, ethanol, acetone (80%) water extracts	Water (mg/g): vanillic (0.093), gallic (0.279) acids, catechin (3.622), chlorogenic (1.457 0.058) and ferulic (6.909) acid; acetone: epicatechin (2.71), protocatechuic acid (5.827)	Effect of solvents, antioxidant, α-glucosidase inhibitory activity	Miao et al., 2016
Crataegus spp., of leaves and flowers	81 components: benzaldehyde (82.54%) butyraldehyde (38.27%), (E)2-hexenal (21.67%)	Volatile, components, aroma	Ozderin et al., 2016
C. pinnatifida,	GC area %: Methyl acetate (4.40), n-hexane (2.90), 2-methyl-furan (1.80), 3-methyl- butyraldehyde (3.64), hexanal (2.08), furaldehyde (5.77), D-limonene (7.99)	Volatile compounds, aroma	Zhong et al., 2015
Hawthorn, pharmaceutical forms of inflorescence	Essential oil (%): 0.05 to 0.20% v/w; tricosane $(12-17)$, $(11-16)$, $(6-11)$, <i>n</i> -hexadecanoic acid $(1-11)$, nonadecane $(3-7)$, (E,E) - α -farnesene $(1-5)$, caryophyllene oxide $(1-4)$, methyl eugenol (up to 6).	Essential oil, aroma	Kowalski et al., 2018

Table 2. Phytoconstituents of Crataegus spp. (recently reported; newly identified compounds are underlined) - (continued)

Abbreviations are: glcp,glucopyranoside; glc, glucoside; TPC,total phenolic content; TFC,total flavonoid content; GAE,gallic acid equivalents; and QE,quercetin equivalents.

C. pycnoloba extract, including the atom numbering of the newly discovered compound (Agalou et al., 2018); 9 compounds were isolated from *C. oxyacantha*, including 2 new natural products (Ali et al., 2017). Selected phytochemicals and antioxidant potential were studied in the *C. monogyna* ethanolic extracts from bark, leaves and berries: the highest TPC, radical scavenging potency as well as the levels of oleanolic acid, quercetin and lupeol were found in the bark extract, while the highest ursolic acid content was in the berries extract (Rezaei-Golmisheh et al., 2015).

The main phenolics in ethanolic extracts of leaves and berries of *C. orientalis* Pall. *ex* M. Bieb. from F.Y.R. Macedonia were hyperoside, isoquercitrin and chlorogenic acid (Šavikin et al., 2017), while vitexin and hyperoside, commonly found in chemotaxonomic investigations of *Crataegus* spp., were not detected in dry leaves with flowers, and berries of *C. rhipidophylla* Gand., *C. × subsphaericea* Gand., and *C. × macrocarpa* Hegetschw grown in Bosnia (Čulum et al., 2018). The presence of polymeric polyphenols (procyanidin dimers, trimers, and tetramers) in *C. pubescens* fruit from Mexico was reported for the first time (González-Jiménez et al., 2018). The composition and antioxidant activity *in vitro* were reported for different extracts of *C. microphylla* Koch subsp. *malyana* K. I. Chr. and Janjic; interestingly, the Soxhlet extract of leaves with flowers was best in DPPH^{*} scavenging (IC₅₀ = 0.78 mg/mL) while that of berries was stronger as ABTS⁺⁺ scavenger with IC₅₀ of 0.39 mg/mL (Čopra-Janićijević et al., 2018). TPC in methanol extracts of *C. monogyna* from nine different locations in central Spain was in the range of 117.7–204.3 mg GAE/g extract, the amounts of chromatographically quantified flavonoids and phenolic acids was 23.3–143.3 mg/kg, while ORAC and IC₅₀ of DPPH[•] scavenging values were 1.32–2.76 mol TE/mg and 0.82–3.76 g/mL, respectively (Abuashwashi et al., 2016). TPC and DPPH[•] scavenging activity as well as the contents of metals (Zn, Fe, Cu, Mn, Cd, Cr, and Pb) were reported in wild *C. oxyacantha* from Serbia (Veličković et al., 2016).

Chlorogenic acid, hyperoside, and rutin were the most abundant phenolics in the extracts of hawthorn flowers in most genotypes of 56 studied *Crataegus* spp. samples collected from different geographical regions of Iran (Alirezalu et al., 2018). The composition of 22 secondary metabolites (flavonoids and phenolic acids) in leaves, flowers and fruits of endemic in Kazakhstan *C. almaatensis* Pojark was compared with a well-known *C. oxyacantha* flowers. Leaf extracts were the richest sources of metabolites (TPC = 218 mg/g) and the most active DPPH[•] scavengers (IC₅₀ = 48 μ g/mL), while the flowers of the Kazakh species were as rich in polyphenols as the *C. oxyacantha* (Bekbolatova et al., 2018).

Evaluation of 18 hawthorn genotypes selected from repository collection in Malatya province (Turkey) belonging to several Crataegus spp. revealed that the genotype 44MA12 (C. monogyna subsp. azarella) had the highest anthocyanin content (516 mg per 100 g fresh fruit) and the strongest DPPH' scavenging capacity, while the genotype 44MA11 (C. meyeri) had the highest TPC, 3,460 mg GAE/100 g fresh fruit (Ercisli et al., 2015). Thirty-five compounds were isolated from C. dahurica methanol extract for the first time and their structures identified as triterpenoids and polyphenolics as the main components (Wang et al., 2018). Evaluation of antioxidant activities, TPC, TPF of crude methanolic extract and its fractions (ethyl acetate, diethyl ether, and chloroform) obtained from Algerian C. azarolus showed that the aerial parts extracts of this species are a good source of natural antioxidants (Lakache et al., 2016). Anthocyanin composition of different wild and cultivated berry species including Chinese hawthorn were quantified by Veberic et al. (2015). Thirty six compounds were reported in different extracts of hawthorn fruit, 15 of them were tentatively identified in hawthorn fruits for the first time (Miao et al., 2016). TPC, condensed tannin content and DPPH[•]/ABTS[•] scavenging capacities were higher for fruit extracts of C. pinnatifida Bge. var. major N. E. Br. (Shanlihong) than for the other two Chinese hawthorn varieties, namely C. pinnatifida Bge. (Shanzha) and C. pinnatifida Bge. var. pinnatifida (Dou et al., 2015). The advantages of Shanlihong variety, which exhibited elevated levels of TPC and TFC, including free and bond phenolics (procyanidin B-2 epicatechin, chlorogenic acid, hyperoside, and isoquercitrin), were also reported by Wen et al. (2015). Among the three tested varieties, the ORAC, and hydrophilic peroxyl radical scavenging capacity of the free faction were 398.3-555.8 µmol TE/g DW, and 299.1–370.9 µmol vitamin C equivalents/g DW, respectively, while the corresponding cellular antioxidant activity (CAA) values were 678-1,200 µmol of QE/100 g DW in the no PBS wash protocol, and 345.9-532.9 µmol of QE/100 g DW in the PBS wash protocol. Finally, the fruits of C. pinnatifida were subjected to in vitro digestion and it was determined that 37.41 and 31.51 mg GAE/g DW of TPC were released for Shanlihong and Dajinxing, respectively, while procyanidin B2, epicatechin, chlorogenic acid and catechin were the major released flavonoids. ORAC and peroxyl radical scavenging capacity well correlated with the released TPC or flavonoids (Zheng et al., 2018).

Some studies have reported the volatile compounds of hawthorn although the content of essential oil in Crataegus spp. is usually very low. For instance, in 4 tested pharmaceutical forms of hawthorn inflorescence it was from 0.05 to 0.20% (v/w) and composed mainly of long chain hydrocarbons (Kowalski et al., 2018). Zhong et al. (2015) identified 46 volatile compounds in C. pinnatifida with contents higher than 1% in the total GC area by HS-SPME coupled with GC-MS. The contents of furaldehyde, 5-methylfuraldehyde, methyl acetate, 2-methylbutyraldehyde, D-limonene and 2-methylfuraldehyde significantly changed after stir-frying and it might be important in odor changes. Volatile components were also reported in C. pentagyna subsp. pentagyna, C. orientalis subsp. orientalis, C. orientalis subsp. szovitsii, C. tanacetifolia, C. azarolus var. aronia, C. monogyna var. lasiocarpa, C. monogyna var. monogyna leaf and flower samples collected from different provinces of Western Anatolia (Ozderin et al., 2016).

The changes of phytochemicals during processing and storage were also studied. "Xinglongzirou" hawthorn wine bioactives and antioxidant capacity were determined during production and storage. Only six anthocyanins were detected after fermentation. Microwaving and heating pretreatments significantly increased the total amount of anthocyanin and antioxidant capacity values in the stored hawthorn wine while the TPC decreased (Liu et al., 2018). Heat and microwave pretreatments had also a significant impact on anthocyanins in hawthorn drink; more of them remained after heat treatment than after microwaving (0.745 mg/100 mL); these were 52.4% higher than those in the control group after storage for 7 days (Liu et al., 2016). Chemometric methods (spectrophotometry and HPLC) were successfully applied to differentiate raw and processed Crataegi Fructus (Fei et al., 2018). Significant differences in physicochemical characteristics associated with fruit quality and free amino acids were found during maturation of hawthorn fruits; for instance, the content of moisture, total soluble sugars, soluble pectin, reduced ascorbic acid, total ascorbic acid, fructose, and sucrose increased while crude protein content decreased significantly (Li et al., 2015).

Some studies were evaluating extraction methods. In general, selection of extraction solvents and procedures may have different goals, e.g. obtaining high total yields or recovery target bioactive fractions/constituents. Ethanol and water, as the most friendly and effective polar solvents, have been most widely used for food, natural pharmaceuticals and cosmetics. For instance, different ethanol concentrations (0-100%) were applied for C. pinnatifida and the yields were 19.03, 33.16, 27.79, 21.88, and 21.71% at 100, 70, 50, 30 and 0% (water) concentration, however the highest TPC and DPPH[•]/ABTS^{•+} scavenging capacity was obtained in case of 50% (Kang, 2015). The technical and economic advantages of pressurized hot water (90 °C) extraction of C. monogyna compared to traditional percolation with ethanol and water (70:30, v/v) was shown by successful incorporation of natural batch variability into the physico-chemical process modelling concept (Sixt and Strube, 2018). Matrix solid-phase dispersion process was also suggested as a good alternative to the classic methods for extracting polyphenols from C. oxyacantha fruits and leaves, although phenolic profile was solvent-dependent (Benabderrahmane et al., 2018). In addition, this study claimed that epicatechin gallate, caftaric acid and orientin were not previously reported in this species. Ultrasonic assisted extraction (UAE) using an alkyl polyglycoside surfactant and further separation and purification with commercial macroporous resins was shown as an effective techniques for recovery of vitexin and its 2"-O-rhamnoside from the leaves (Han et al., 2016a). The extract obtained by UAE from C. pinnatifida fruits demonstrated higher TPC, TFC and DPPH[•] scavenging capacity (Park et al., 2017b). Aqueous solutions of citric acid and methanol were compared for C. monogyna fruit; it was found that the former was favorable in terms of total yield, TPC and ascorbic acid, while DPPH and ABTS + scavenging capacity was not dependent on the solvent (Pliszka et al., 2016). Hawthorn seeds, as a byproduct of manufacturing hawthorn juice and jam, were evaluated as a potential new and cheap source of antioxidants and inflammation inhibitor (Huang et al., 2015).

A protocol for preparative isolation of oligomeric and polymeric procyanidins from an acetone-water extract of Crataegi folium *cum fore* was developed, yielding procyanidin reference clusters with defined degree of polymerization (DP) from 2 to 10 (Hellenbrand et al., 2015). In addition, monitoring of procyanidin distribution during seasonal growth of fresh *C. monogyna* plants showed that their contents were between 20 and 55 mg/g DW of oligomeric procyanidins during the growing season in the different plant organs with strong accumulation in the flowers and fruits (55 mg/g DW). Regarding other components, the viscoelastic behavior and texture profile of gels, composed of the high methoxypectin in

Venskutonis

hawthorn, were compared with commercial citrus gels (Linares-Garcia et al., 2015); the homogenous polyphenolic-polysaccharide conjugates (MW > 760×10^3 and 970×10^3 g/mol) containing some flavonoid units and rich in galacturonic acid with low esterification degree were isolated from flowers and fruits of *C. monogyna* (Pawlaczyk-Graja, 2018).

3. Cardioprotective benefits

Cardiovascular health benefits of various *Crataegus* drugs and other preparations have been most widely studied. In addition, the potential role of hawthorn in cardiovascular diseases was specifically reviewed by Chang et al. (2005). Hawthorn extracts have been used for this purpose in many countries, particularly against mild forms of chronic heart failure. For instance, search word combination *Crataegus*+cardiovascular in Clavirate Analytics Web of Science (WoS) since 1980 gave 286 records, and even during last 4 years (since 2015) the number of publications on this topic remains quite high, 61 (accessed on December 7, 2018). Consequently, the interest in hawthorn as a source of natural cardioprotective medicine, nutraceutical and functional food is evident.

Hawthorn extract WS 1442 containing a range of pharmacologically active substances including oligomeric proanthocyanidins and flavonoids is, possibly, the most famous Crataegus fruit-based drug with 89 and 14 records in WoS since 1980 and 2015, respectively. Moreover, two most recent articles reviewed WS 1442 in terms of experimental and clinical experiences (Zorniak et al., 2017) and benefit-risk assessment (Holubarsch et al., 2018). Therefore, only those studies, which were not reviewed in these publications, will be covered briefly. Thus, Fuchs et al. (2017) reported that WS 1442 protective effects against mild forms of chronic heart failure dysfunction may be due to the promotion of endothelial barrier integrity and inhibition of endothelial hyperpermeability, which are exerted by activating barrier enhancing (cortactin activation) and blocking barrier disruptive (calcium signaling) pathways, respectively. Bioactivity-guided fractionation of WS 1442 using successive elution with water, 95% ethanol, methanol, and 70% acetone, revealed that only the ethanolic fraction interfered with calcium signaling and only the methanolic fraction led to an activation of cortactin. Moreover, the role of phenolic compounds was excluded from the calcium active substance, whereas cortactin activation was attributed to oligomeric procyanidins with a distinct degree of polymerization (Fuchs et al., 2017). The study with human umbilical vein endothelial cells (HUVECs) exposed to TNF- α , with or without simvastatin (positive control) and WS 1442, suggested that the factors upregulating miR-155 expression, which decreases eNOS expression and endothelial function impairment, may be mitigated by WS 1442 (Wang et al., 2018). The results demonstrated that both TNF- α and WS 1442 increased miR-155 expression and decreased eNOS expression, however, unlike TNF-a, WS 1442 increased phosphorylated eNOS expression and NO concentrations and mitigated the overall negative effect of miR-155. Protecting effects of total flavones of hawthorn combined with resveratrol on the endothelial cells injury after artery bypass graft surgery were demonstrated in the study with rabbits, which determined that the level of circulating endothelial cells, density and the expressions of albumen and mRNA of ICAM-1 were significantly decreased (Zhu et al., 2018). It was also reported that WS (R) 1442 improved vascular function in diet-induced obese mice (Xia et al., 2016). The activity of WS 1442 in stimulating cardiomyocyte differentiation from murine and human embryonic stem cells (ESC) after forming mesoderm was validated in a mESC-based spontaneous differentiation assay, while bioassay-guided fractionation suggested that this activity is exerted by specific classes of compounds (Halver et al., 2015). Aerobic exercise and administration of *C. oxyacantha* extract reduced ICAM-1 and E-selectin in serum levels of 80 stable angina pectoris patients and such combined treatment was suggested as an effective complementary strategy for lowering the risk of atherosclerosis and heart problems (Jalaly et al., 2015).

The cardiac effects of hydroalcoholic extract of C. pentagyna leaf as well as isoquercetin and vitexin were studied using cardiomyocytes differentiated from healthy human embryonic stem cells, namely long QT syndrome type 2 and catecholaminergic polymorphic ventricular tachycardia type 1 (CPVT1) patient-specific induced pluripotent. It was concluded that tested preparations may be introduced as a novel nutraceutical with antiarrhythmic potential for CPVT1 patients (Pahlavan et al., 2018). The effects of polyphenolic extracts of C. pentagyna and C. microphylla fruits on hypoxia, which is a state of oxygen deficiency occurring in heart diseases, ischemia, bleeding and heart attack, were studied using male Swiss albino mice model. It was found that the extracts exhibited significant anti-hypoxic activity and prolonged animal survival time (Ebrahimzadeh et al., 2018). The activity of these ex-tracts was attributed to a high TPC, 1,136.0 and 721.1 GAE mg/g in C. pentagyna and C. microphylla, respectively. Polyphenolicpolysaccharide conjugates from flowers and fruits of C. monogyna prolonged the plasma coagulation process in in vitro tests, even at a concentration of 31.25 μ g/mL; however, only the product from flowers was highly selective in its action. It was mainly the indirect inhibitor of factor Xa, mediated by antithrombin, where such mechanism of activity is typical for highly sulfated glycosaminoglycans (Pawlaczyk-Graja, 2018).

Flavonoid-rich preparations of C. pinnatifida leaves (HLF) have been used to treat cardiovascular diseases; however, their ability to attenuate atherosclerosis development and possible mechanism are not evident. To fill this gap, Dong et al. (2017) by the large scope in vivo studies using apoE knockout mice showed that administration of HLF resulted in the following effects: (1) reduction of the mean atherosclerotic lesion area in en face aortas; (2) decrease in total cholesterol (TC) and very low density lipoprotein cholesterol plus low density lipoprotein cholesterol (VLDLc+LDLc) levels; (3) increase in PPAR α mRNA and RCT; (4) decrease in SREBP-1c; (5) induction of CPT-1 mRNA, SOD1 and SOD2, Gpx3 mRNA, LDL receptor mRNA and protein levels; and (6) inhibition of the foam cell formation. HLF also protected rats against diabetic cardiomyopathy and the mechanism may be involved in reducing oxidative stress and inflammation via inactivation of the PKC-α signaling pathway (Min et al., 2017).

The new norditerpenoids (Table 2) isolated from the leaves of C. pinnatifida exhibited exceptionally potent antithrombotic activities in vitro and in vivo, pinnatifidanoside F being the most potent one (Gao et al., 2017). It inhibited ADP induced platelet aggregation, which is mediated through the response to the specific receptor of P2Y(12), and prolonged the time to form thrombocytes induced by FeCl₂ in the caudal vessels of zebrafish. In vivo study with diabetic rats demonstrated that resistance training and C. oxyacantha extract can synergistically decrease ischemia-reperfusion injury by oxidative stress reduction: injury indices (plasma lactate dehydrogenase, creatine kinase myocardial band isoenzyme and infarction size) were significantly lower after sedentary diabetic group treatment with extract, whereas glutathione peroxidase and myeloperoxidase levels after reperfusion increased and decreased, respectively, in response to training and C. oxyacantha (Ranjbar et al., 2018). On the other hand, the study with 116 patients who underwent cardiac surgery showed that those who recently consumed hawthorn extract had a significantly higher rate of postoperative bleeding and overall mortality rate, which indicates that hawthorn extract consumption increases the potential for bleeding and the amount of chest tube output after cardiac surgery (Rababa'h et al., 2016).

C. oxvacantha alcoholic extract produced an antiarrhythmic effect that was induced by digoxin in Wistar rats; however, in the clinical use of this extract, the hypotensive effect should be considered (Alp et al., 2015). Another in vivo study reported that administration of methanolic C. songarica extract at the dose of 300 mg/kg body weight to albino rats with CCl₄-iduced toxicity significantly decreased serum creatinine, urea, and cholesterol, malondialdehyde (MDA) in kidney and heart tissues, along with recovery in antioxidant enzyme levels (Ganie et al., 2016). It was also reported that standardized Crataegus extract prevented DOCA-salt-induced hypertension and alteration of cardiac, vascular and renal structure and function in rats (Schini-Kerth et al., 2015). One of the main hawthorn bioactives vitexin inhibited cytochrome P450 enzyme (CYP3A1) in a concentration-dependent and time-dependent manner while CYP2C11 enzyme activity was induced after short period treatment but inhibited after long period treatment. It was concluded that vitexin can either inhibit or induce the activities of these enzymes and possible herb-drug interactions should be considered when vitexin is co-administered with some CYP2C11 or CYP3A1 substrates in clinic (Wang et al., 2015).

Some studies reported antithrombotic and anti-vasoconstriction effects exerted by *Crataegus* bioactives. Thus, among 15 from *C. pinnatifida* leaves isolated compounds (Table 2) (65,7Z,9R)-roseoside, eriodectyol and 2"-O-rhamnosyl vitexin exerted potent antithrombotic activity *in vitro*, which was in agreement with the *in vivo* results obtained in transgenic zebrafish system (Li et al., 2015). *C. monogyna* and *C. davisii* significantly inhibited the carrageenan-induced mouse tail thrombosis and may potentially be used as therapeutic agents or complementary treatments against thrombosis (Arslan et al., 2015). Euscaphic acid isolated from the crude ethyl acetate extract of *C. azarolus* var. *aronia* showed high anti-vasoconstriction effects on aortic rings, supporting the use of this medicinal plant in cardiovascular disease (Mahmud et al., 2016).

4. Hypolipidemic and hypoglycemic activity

4.1. Hypolipidemic effects

Lipid-lowering effects of hawthorn have been widely studied *in vivo* in many cases using HFD animals. Plant preparations demonstrated the ability to lower tricylglycerol (TG), total cholesterol (TC), LDLC and VLDLC, and to increase HDLC in plasma and these effects have been explained by several possible mechanisms (Yang and Liu, 2012). Recently published articles extended existing knowledge on hypolipidemic benefits of *Crataegus* preparations, particularly HLF, some of them carried out in *in vivo* studies with NAFLD animals and reported beneficial effects on liver health, which highly depends on HFD.

C. pinnatifida extract decreased the levels of serum TC, LDLC, hepatic TG and MDA, increased mRNA expression of nuclear erythroid 2-related factor (Nrf2), heme oxygenase-1 (HO-1), Gpx and reversed the suppression of protein levels in ovariectomized rats (Yoo et al., 2016). Li et al. (2015) reported that *C. pinnatifida* HLF ameliorated hepatic steatosis by enhancing the adiponectin/AMPK pathway in the liver of HFD-induced NAFLD rats. Several beneficial events were determined: (1) lowering body and liver

weight and liver/body weight ratio; (2) improving serum parameters and liver dysfunction; (3) decreasing hepatic lipid accumulation; (4) increasing circulating adiponectin levels and up-regulating the expression of adiponectin receptors (AdipoR2) in the liver; and (5) activating AMP-activated protein kinase (AMPK) and altering AMPK-mediated SREBP-1c, PPAR- α and their downstream targets (Li et al., 2015). The inhibitory activities of *C. pinnatifida* extract isolated with 50% ethanol were more effective against formation of advanced glycation end products (AGEs) and α -glucosidase, while water extract better inhibited aldose reductase (Kang, 2015).

Investigation of the effects of aqueous extract of *C. aronia* (syn. *azarolus* L.) on sperm parameter and testicular structure in control and NAFLD-induced rats showed the improvements of the following indicators: (1) lower hepatic TG and cholesterol contents; (2) higher levels of testosterone, luteinizing hormone and follicle-stimulating hormone; (3) increased epididymis weight, sperm count and motility; and (4) increased testicular levels of glutathione, higher protein levels of Nrf2, γ -glutamylcysteine synthetase and SOD (Dallak, 2018).

The potential of novel multi-targeted herbal formula containing Crataegus Fructus, sylimarin, Schisandrae Fructus and *Momordica charantia*, as a therapeutic agent for diet-induced metabolic syndrome with special emphasis on NAFLD, dyslipidemia and type 2 diabetes, was studied using male C57Bl/6 mice. Crataegus Fructus aqueous extract inhibited differentiation of 3T3-L1 preadipocytes and cholesterol uptake into Caco-2 cells, while the whole herbal formula exhibited mitigation of diet-induced increase of various indicators *in vivo*, namely (1) a trend to reduce body weight and fat pad mass, (2) significant reduction in liver weight, liver lipid, and plasma lipid, (3) reduction of liver inflammation, and (4) improvement of adiponectin level (Elaine et al., 2018).

The results of a number of studies support the hypothesis that hawthorn possesses both therapeutic and protective effect for NAFLD. C. aronia extract and simvastatin significantly reduced lipids and thiobarbituric acid reactive substances (TBARS), which were increased by oxidative stress in HFD rat group, and treated damage to hepatic cells, while only the extract induced GSH. C. aronia and simvastatin treatments differentially reversed hepatic injuries (Al Humayed et al., 2017). Another in vivo study demonstrated that C. aronia significantly reduced liver index (3.85% versus 6.22% in HFD rat group), increased HDLC/LDLC and improved oxidative stress biomarker and enzymes indicating liver damage (Al Humaved, 2017). Ethanol extract of C. oxvacantha administered at 10 and 20 mg/kg to HFD rats reduced the level of some liver biomarkers, especially lactate dehydrogenase (LDH), increased GSH and FRAP and decreased lipid peroxidation, which may be beneficial in restoring the hepatocyte damage (Saeedi et al., 2018).

Polyphenols-enriched extracts from hawthorn fruit peels (HPP) and flesh (HFP), containing chlorogenic acid, epicatechin, rutin and hyperoside as the main polyphenolic constituents (HPP > HFP), reduced alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) activities, as well as the ratio of pro-apoptotic protein with B-cell lymphoma-2 (Bax/Bcl-2) in mice with liver injury, which was induced by a high-fructose diet. HPP was more effective than HFP in mitigating liver inflammation and oxidative stress by inhibiting inflammatory cytokine (TNF- α , IL-1 and IL-6) release, elevating antioxidant enzyme activities and PPAR α expression, reducing Nrf-2 and antioxidant response element (ARE) expression in mice. HPPtreated mice also had lower levels of TC, TG, LDLC, VLDLC and apoprotein B (Apo-B), and higher levels of HDLC and Apo-A1 than HFP-treated mice due to reduced expression of fatty acid syn-

thetase (FAS) (Han et al., 2016b).

Hypolipidemic and antioxidant effects of aqueous and ethanolic extracts of C. pinnatifida fruits were compared in hyperlipidemia rats and obvious differences between extracts were observed, probably due to the existing differences in the content of phenols (Shao et al., 2016). Polyphenolic extracts of C. azarolus and C. monogyna leaves, fruit peel, and pulp, possessing antioxidant activity, protected against thermal-cholesterol degradation while pre-treatment with the extracts preserved liposomes and rat liver-homogenate from oxidative Cu2+ and Fe2+ induced damage (Belkhir et al., 2016). Synergic effects and potential mechanisms of action of Astragalus polysaccharides combined with Crataegus flavonoids were studied with diabetic mice. The mixture significantly reduced the fasting blood glucose, food and water intake and restored increased serum insulin levels and islet cell function; it also increased protein expression levels of pancreatic and duodenal homeobox-1 and phosphorylated AMPK in the pancreatic and liver tissue samples, respectively. The mixture significantly increased the mRNA expression levels of neurogenin 3 (v-maf musculoaponeurotic fibrosarcoma oncogene), protein A and insulin, and simultaneously decreased the expressions of IL6, TNF- α and chemokine (C-C motif) ligand 2 in the pancreatic islet cells of diabetic mice (Cui et al., 2016).

One of the most important *Crataegus* phytochemical, hyperoside, induced apoptosis in human hepatic stellate LX-2 cells and decreased the levels of α -smooth muscle actin (α -SMA), type I collagen, and intracellular ROS. Remarkably, hyperoside also inhibited the DNA-binding activity of the transcription factor NF- κ B and altered expression levels of NF- κ B-regulated genes related to apoptosis, including pro-apoptotic genes Bcl-Xs, DR4, Fas, and FasL and anti-apoptotic genes A20, c-IAP1, Bcl-X-L and RIP1. These results suggest that hyperoside may have potential as a therapeutic agent for the treatment of liver fibrosis (Wang et al., 2016).

4.2. Hypoglycemic effects

Enzyme inhibitory activity *in vitro* is a simple and widely used method for the preliminary evaluation of hypoglycemic effects. Comparison of hawthorn extracts prepared from plant fruit using 80% ethanol, 80% methanol, 80% acetone and pure water revealed that acetone extract had the highest α -glucosidase inhibitory activity while the highest DPPH[•] scavenging capacity and FRAP was exhibited by the water extract. Polyphenols, triterpenoids, protocatechuic acid and epicatechin contributed to the α -glucosidase inhibitory activity, while flavonoids, polyphenols, vanillic acid, gallic acid, catechin and chlorogenic acid contributed to the antioxidant activity (Miao et al., 2016).

Antihyperglycemic (Male albino mice) and antihyperlipidemic (Sprague Dawley rats) activities of the ethanolic extract of C. azarolus var. eu-azarolus leaves and the isolated 3β-O acetyl ursolic acid were proved by significantly reduced mice blood glucose level in a time-and dose-dependent manner, while ethanolic extract significantly reduced LDLC, VLDLC, TC and TG and increased HDLC. In addition, ethanolic extract and 3β-O acetyl ursolic acid reduced the activity of pancreatic lipase in vitro (Abu-Gharbieh and Shehab, 2017). C. aronia significantly improved the oral glucose tolerance test, lowered plasma glucose, serum lipid levels and the hepatic glycogen content. In addition, it significantly lowered the levels of hepatic lipid peroxidation, TNF- α and IL-6, while the level of reduced glutathione (GSII) was enhanced and SOD activity was increased (Mostafa et al., 2018). Regarding the mechanism involved, it enhanced hepatic mRNA expression of the insulin receptor A isoform (IR-A) and glucose 6-phosphatase (G6Pase), while the expression of glucose transporter-2 (GLUT-2) and glycerol kinase (GK) mRNA was lowered. Consequently, *C. aronia* ameliorated type 2 diabetes mellitus by inhibiting hepatic glucose output. Out of six tested plants, ethanolic extract of *C. monogyna* leaves exhibited the best α -glucosidase inhibition activity with IC₅₀ = 10.39 µg/mL, and was stronger inhibitor of α -amylase and α -glucosidase than its traditionally made with hot water tea (Sayin et al., 2018).

Complex herbal formulas with Crataegus have also been tested. The potential anti-obesity effects (inhibition of lipogenesis and adipogenesis) of HT048 (a combination of C. pinnatifida leaf and Citrus unshiu peel extracts) was investigated in vitro and in vivo using 3T3-L1 adipocytes and male Sprague Dawley rats, respectively (Lee et al., 2015). HT048 suppressed, dose-dependently, adipocyte differentiation and stimulated glycerol release and decreased the expressions of PPAR and C/EBP mRNA in adipocytes while in rats it significantly reduced the body and fat, as well as serum lipid levels, decreased expression of the hepatic lipogenesis-related genes and increased the expression of the oxidation-related genes (Lee et al., 2015). The effects were supported one year later when it was demonstrated that HT048 decreased body and total white adipose tissue weight and serum insulin levels in HFD-fed obese rats, whereas at the molecular level the supplement downregulated genes involved in lipogenesis, gluconeogenesis, and adipogenesis, and up-regulated β-oxidation genes. In addition, no interactions were observed between HT048 supplementation and orlistat drug, which was also used in the study (Lee et al., 2016). Orlistat and C. monogyna ethanolic extracts significantly lowered the hypercholesterolemia-increased serum level of hepatic enzymes and lipid peroxidation level in the hypercholesterolemia-induced oxidative stress in rats, protected from hepatic thiol depletion, and improved lipid profile and hepatic damages (Rezaei-Golmisheh et al., 2015). The results of Cheng et al. (2015) study with mice demonstrated that supplementation with a formula composed of Rhizoma dioscorea, Lycium barbarum, Prunella vulgaris and hawthorn may be a potent alternative as an anti-diabetic health-promoting diet. Finally, hawthorn pectin pentasaccharide (HPPS) was shown to be more effective than pectin (HP) and pectin hydrolyzates (HPH) in decreasing the body weight gain, liver weight, and plasma and hepatic TC of hamsters fed by high-cholesterol diets. However, the HP group had higher cholesterol excretion capacities than the HPH and HPPS groups by inhibiting cholesterol absorption in the diet (Zhu et al., 2015a). Thus, HPPS could be a promising antiatherogenic dietary ingredient for the development of functional food to improve cholesterol metabolism.

5. Anticancer effects

The majority of studies evaluating anticarcinogenic properties of plant origin products are based on cell cytotoxicity and apoptosis assays, while *in vivo* studies, which provide more relevant physiologically information, are rather scarce. In addition, the mechanism of activity is very important issue in evaluating anticancer properties. Several recent studies, which were focusing on anticancer effects of *Crataegus* extracts, their fractions and purified compounds, are reviewed in this section.

Hawthorn's health benefits are mainly associated with the high contents of polyphenolics. *C. pinnatifida* var. *major* fruits, containing high amounts of flavonoids and triterpenoids, showed potent antiproliferative effect against human cancer cells lines (liver, Hep G2; breast, MCF-7 and MDA-MB-231); particularly strong activity was determined for triterpenoids-enriched fraction and its main

ingredient ursolic acid, which induced restriction point G1 arrest, downregulated proliferating cell nuclear antigen (PCNA), cyclindependent kinase (CDK4), and a protein Cyclin D1 (in humans encoded by the CCND1 gene) and upregulated cyclin-dependent kinase inhibitor 1p21(Waf1/Cip1) in MDA-MB-231 cells (Wen et al., 2017). In addition, the supplements induced MDA-MB-231 apoptosis *via* mitochondrial death pathway induced by caspase (9 and 3) activation.

The studies performed by Mustapha et al. (2015, 2016a, b, c) demonstrated various effects of C. azarolus ethyl acetate extract as well as isolated from it hyperoside and vitexin-2"-O-rhamnoside. The extract and vitexin rhamnoside exhibited anti-proliferative activity against B16F10 melanoma cells and the ability to reduce melanin content by inhibiting the tyrosinase activities (Mustapha et al., 2015); it also demonstrated significant cellular antioxidant capacity against the ROS in B16F10 and primary human keratinocyte cells (Mustapha et al., 2016b). In addition, total oligomeric flavonoids extract (150 mg/kg body weight, 21 days) significantly inhibited tumor growth volume and weight in Balb/c mice inoculated with B16F10 cells and therefore was suggested as a new candidate for skin care products (Mustapha et al., 2016b). The extract and hyperoside also exhibited an immunomodulatory effect by modulating macrophage lysosomal enzyme activity and NO release in mice and anti-inflammatory activity, which was concomitant with the cellular antioxidant effect against macrophages and solenocytes (Mustapha et al., 2016a). Extract-induced growth inhibitory effect in human colorectal cancer cell lines HCT-116 and HT-29 was associated with DNA fragmentation, sub-G1 peak, loss of mitochondrial potential, and poly (ADP-ribose) polymerase (PARP) cleavage. The extract also induced the cleavage of caspase-8 and had no effect on steady-state levels of total Bcl-2 protein, whereas Bax levels decreased significantly in a dose-dependent manner in both tested cell lines (Mustapha et al., 2016b). It may also be mentioned that in searching of relatively mild and safe tyrosinase inhibitors, one of six isolated from C. pinnatifida seeds 8-0-4' neolignans, demonstrated promising tyrosinase-inhibiting and good antioxidant activities (Huang et al., 2015b)

Two other studies also investigated the effects of hyperoside. It was suggested as a candidate of concomitant treatment for leukemia due to the ability to potentiate As_2O_3 -dependent apoptosis of HL-60 human acute myeloid leukemia cells by upregulating LC-II and inducing autophagy effects (Zhang et al., 2015). In addition, hyperoside inhibited Bcl-2-associated agonist of cell death BAD from phosphorylating, reactivated caspase-9, and increased cell cycle regulating protein p27 levels. In another study, hyperoside dose-dependently (0.5, 1, 2 mmol/L) increased the expression of LC3-II and autophagosome numbers in human non-small cell lung cancer cell line A549 cells; however, such effects in human bronchial epithelial cell line BEAS-2B cells were not observed. Moreover, hyperoside inhibited the phosphorylation of Akt, mTOR, p70S6K and 4E-BP1, but increased the phosphorylation of ERK1/2 in A549 cells (Fu et al., 2016).

The extract from hawthorn fruit peel (EPHF), rich in phenolic compounds, exhibited dose-dependent cytotoxicity on MCF-7 and SKOV-3 human tumor cell lines with the IC₅₀ of 2.76 and 80.11 μ g/mL, respectively (Wu et al., 2017). Polyphenolic extracts of *C. azarolus* and *C. monogyna* leaves, fruit peel, and pulp (0.24–4.8 mg/mL) were cytotoxic to cancer Caco-2 cells, at the same time being nontoxic for differentiated Caco-2 cells (Belkhir et al., 2016). Ethanolic extract of *C. microphylla* flowers and leaf buds demonstrated cytotoxic and antiproliferative activity against HeLa cell (IC₅₀ = 0.871 mg/mL) and least cytotoxicity on normal human peripheral mononuclear cells (Bura et al., 2016). Methanol, ethanol and ethyl acetate extracts of *C. songarica* K. Koch exhib-

ited potent in vitro anticancer activity on MCF-7, HeLa, HepG2, SF-295, SW480 and IMR-32 cancer cell lines with IC₅₀ values of 28.57-85.106 µg/mL, while methanol extract demonstrated protective activity for albino rats' kidney and heart tissue against CCl₄induced toxicity in vivo (Ganie et al., 2016). C. monogyna extract at 100–75 µg/mL concentrations exhibited better antiproliferative activity against C6 cell lines than 5-flurouracil drug (5-FU) and cisplatin, and also demonstrated considerable antiproliferative activity against HeLa cells (Yaglioglu et al., 2016). Ethanolic C. meyeri flower extracts, which contained flavonoids and procyanidins and had high in vitro antioxidant capacity in DPPH[•], β-carotenelinoleic acid and TPC assays, also demonstrated antiproliferative activity against PC3 and PC14 cells and decreased the levels of serum alanine aminotransferase and aspartate aminotransferase in the blood of partially hepatectomized rats (Ozay et. al., 2015). Thus, the extracts were suggested as protective agents against partial hepatectomy-induced liver injury in rats and inhibitors of the proliferation of human non-small lung cancer cells.

Recently isolated from the fruit of *C. pinnatifida* phenylpropanoids crataegusoids (Table 2) were cytotoxic against human hepatocellular carcinoma cells HepG2 and Hep3B, while crataegusoids C and D with two methoxy groups at C-3' most remarkably induced apoptosis in HepG2 cells indicating on structure-activity relationships (Guo et al., 2018). Finally, the hydroxy-olean-12en-28-oic acid (HOA) triterpenoids from hawthorn berry, namely 3β , 6β , 18β ,23-tetraHOA, 2β , 3β , 6β , 18β ,tetraHOA and 2 α , 3β , 6β , 18β ,23-pentaHOA were potent inhibitors of HepG2 and MCF-7 cells, with the EC₅₀ values lower than 5 μ M (Qiao et al., 2015).

6. Neuroprotective effects

Neuroprotective effects of sesquineolignans (Table 2) isolated from the hydroethanolic (70%) extract of *C. pinnatifida* var. *major* seeds towards H_2O_2 -induced damage in human neuroblastoma SH-SY5Y cells were recently reported by Huang et al. (2018a, b). All of these exhibited significant neuroprotective activity, compared with trolox, while 6 compounds demonstrated the survival rate of 90.74% at the 50 µM concentration, by inhibiting cellular apoptosis determined by Hoechst 33258 staining and annexin V/ PI analysis (Huang et al., 2018a). Sesquineolignans also inhibited β -amyloid aggregation; (7'S, 8'R, 8R)-isolariciresinol-9'- β -Dglucopyranoside and lyoniside were stronger inhibitors of A β_{1-42} aggregation than curcumin (Huang et al., 2018b). Possible mechanism of interactions was investigated by molecular docking.

The potential of hawthorn as natural antidepressant was also reported. The levels of typical markers of depression in animal models, monoamine oxidase B and reduced spine numbers along neuronal dendrites, were improved by chlorogenic acid or the extract of C. pinnatifida both in vivo using mice model with induced depression-like phenotypes by daily injection of stress hormone, and in vitro using cultured astrocyte type I clone C8-D1A cells (Lim et al., 2018). A protective effect of vitexin, which is one of the most important flavonoid in hawthorn leaves, against neurotoxicity has also been reported: however, the mechanisms of action remain elusive. Systemic vitexin treatment significantly reduced neurological deficit, cerebral infarct volume and neuronal damage when compared with the ischemia/reperfusion (I/R) injury mice. Vitexin markedly upregulated extracellular signal-regulated kinases (p-ERK1/2), downregulated c-Jun N-terminal kinases (p-INK) and p38 phosphorylation (p-p38), increased B-cell lymphoma 2 (Bcl-2) expression and suppressed the overexpression of Bax in the I/R mice, thus, protecting brain against cerebral I/R injury

regulated by mitogen-activated protein kinase (MAPK) and apoptosis signaling pathways (Wang et al., 2015a). Vitexin presented no cytotoxicity in RAW 264.7 cells and effectively reduced leukocyte migration *in vivo*, TNF- α , IL-1 β , PGE2 and NO releases and increased in IL-10 release in the LPS-challenged mice; it was also able to regulate transcriptional factors for pro-inflammatory mediators, by reducing the expression of p-p38, p-ERK1/2 and p-JNK (Rosa et al., 2016). Chen et al. (2016) also attempted to obtain evidence on mechanisms using Sprague Dawley rats and human PC12 pheochromocytoma neurosecretory cells with isoflurane-induced neurotoxicity and cytotoxicity, respectively, and suggested that vitexin mediates by targeting the transient receptor potential cation channel subfamily V member 1 gene (TRPV1) and N-methyl D-aspartate receptor subtype 2B protein (NR2B) signaling pathways. The study of Hu et al. (2018) demonstrated that vitexin from C. pinnatifida protected dopaminergic neurons against methyl-4-phenylpyridine/1-methyl-4-phenyl-1,2,3,-6-tetrahydropyridine (MPP+/MPTP)-induced neurotoxicity through the activation of phosphatidylinositol-4,5-bisphosphate 3-kinase/ protein kinase B (PI3K/Akt) signaling pathway, and these findings may facilitate the clinical application of vitexin in the Parkinson's disease therapy.

The combination of Radix Puerariae and hawthorn fruit tested with diabetic rats showed the following effects: (1) reduced random blood glucose, TC, TG and improved glucose tolerance; (2) reversed the loss in body weights; (3) reduced depressive-like behavior as measured by open field, the elevated plus-maze, locomotor activity, and forced swimming tests; and (5) upregulated brain-derived neurotrophic factor and activated extracellular signal-regulated protein kinase (Luo et al., 2016). Vitexin exerted behaviorally-specific antinociceptive effect against postoperative pain using a mouse model with surgical incision on the right hindpaw, which was mediated through opioid and GABA(A) receptors (Zhu et al., 2019). It was observed that C. pinnatifida containing Naodesheng formula could reverse most of the cerebral ischemia reperfusion induced imbalanced metabolites in brain tissue, plasma and urine, which indicates on its protective effect on ischemic stroke rats by mechanisms involving multiple metabolic pathways, including energy metabolism, amino acid metabolism, oxidative stress and inflammatory injury (Luo et al., 2016). Most recently potential efficacy of Naodesheng for treating Alzheimer's disease was evaluated by combined machine learning, molecular docking, and pharmacophore mapping, and as a result constituent-target network, constituent-target-target network and target-biological pathway network was built for this formula as the virtual screening and network pharmacology method for the first time (Pang et al., 2018). C. oxyacantha extract ameliorated some seizure parameters in penicillin-induced epilepsy in gerbils; however, further and more advanced physiologic and neurochemical studies are required to determine the mechanisms involved (Cakir et al., 2016).

The compounds from *C. oxyacantha* effectively inhibited AChE, from $IC_{50} = 5.22 \ \mu M$ (β -sitosterol-3-*O*- β -D-glucopyranoside) to 44.47 μ M, and BChE activity with $IC_{50} = 0.55-15.36 \ \mu$ M. In addition, docking procedures with Genetic Optimization for Ligand Docking suit v5.4.1 were applied to study the inhibition mechanism, while software admetSAR predicted that both of the isolated compounds can cross blood brain barrier (BBB+) (Ali et al., 2017). AChE catalyzes the breakdown of acetylcholine and of some other choline esters that function as neurotransmitters, while BChE in plasma can be used as a liver function test in both hypercholinesterasemia and hypocholinesterasemia. Potential antioxidant, hypoglycemic, and neuroprotective effects of acidified methanol or ethanol extracts of *C. microphylla* stem barks and leaves were supported by evaluating their AChE, tyrosinase and α -glucosidase inhibitory, DPPH[•] scavenging capacity and protective activity against hydroxyl radicals-induced DNA strand scission (Renda et al., 2018). Macroporous resin Diaion HP-20 adsorption chromatography was applied to obtain rich in phenolics extract from hawthorn fruit peel (EPHF), which contained ideain, epicatechin and chlorogenic acid as the main constituents (179.4, 40.9, and 10.0 mg/g, respectively) and exhibited strong antioxidant (ORAC = 11.65 μ M TE/mg, IC₅₀ in DPPH[•] scavenging = 6.72 μ g/mL) and AChE inhibitory (IC₅₀ = 11.72 μ g/mL) activities (Wu et al., 2017). Methanolic extract of *C. microphylla* was shown as a good source of antioxidant, antimicrobial, antidiabetic, anticholinesterase, and anticancer agents in various *in vitro* assays (Aksoy-Sagirli at al., 2017).

7. Skin protective and other health effects

The extract from the Greek hawthorn C. pycnoloba was tested as a potent inhibitor of melanin synthesis during early zebrafish embryo development. The activity based subfractionation enabled to identify 3 melanogenesis inhibiting dibenzofurans (Table 2), acting not via inhibition of tyrosinase or interfering with neural crest differentiation or migration, but via binding to the aryl hydrocarbon receptor by activating its signaling pathway and causing the induction of the target genes expression (Agalou et al., 2018). Ursolic aldehyde, isolated from the active fragments of C. pycnoloba, had no melanin synthesis inhibition activity. The assays with HaCaT human keratinocytes, normal human dermal fibroblasts (HDFs), and mice showed that hawthorn polyphenol extract (HPE), containing chlorogenic acid, procyanidin B2, and epicatechin (13.5, 19.2, and 18.8% of the TPC, respectively), can prevent UVB radiation-induced skin photo-aging by promoting human cell proliferation in vitro and regulating matrix metallopeptidase expression and type I procollagen production *in vivo* (Liu et al., 2018). Ethanol extract from C. pinnatifida fruits (TPC = 61.31 mg/g, TFC = 25.42 mg/g) at 50 µg/mL reduced melanin contents by 8.5% while at 1,000 µg/mL reduced intracellular tyrosinase activity by 46.83%, thus suggesting that the extract could be used as a whitening agent in cosmetics (Park et al., 2017a). The mixture of P. ginseng and C. pinnatifida improved procollagen type I expression, diminished matrix metalloproteinase-1 secretion and improved various other human skin values associated with aging, wrinkle formation and moisture (Hwang et al., 2017). Creams with 5-15% of C. pontica leaf extract have a healing effect on dermal toxicity caused by T-2 toxin as it was shown by the study with Iranian rabbits (Kalantari et al., 2016). The penetration of active substances into the full undamaged human skin from the semisolid preparations containing C. oxyacantha extract was evaluated ex vivo. However, it was observed that, although the extract exhibited antimicrobial and antiradical activity, its compounds stay on the surface of the undamaged human skin, thus only traces of hyperoside and isoquercitrin were found in the epidermis (Stelmakiene et al., 2016).

The effects of *C. orientalis* M Bieber extract on serum oxidative status and alveolar bone loss in experimental periodontilis was studied *in vivo* and showed inhibitory effect on periodontal inflammation and alveolar bone loss by regulating total antioxidant (TAS)/oxidant (TOS) status and oxidative stress index levels in periodontal disease in rats when administered systemically (Hatipoglu et al., 2015). *C. azarolus* extract was active against herpes simplex virus-1 (HSV-1), and antiviral activity correlated with TPC (R = 0.773, p < 0.001) and free radical scavenging capacity (R = -0.684, p < 0.01) (Moradi et al., 2018). Hawthorn hyperoside (10, 50, 100 µmol/L) dose-dependently inhibited LPS-induced proliferation and migration of human rheumatoid fibroblast-like synoviocytes (RA FLSs) *in vitro*. Furthermore, hyperoside decreased LPS-stimulated production of TNF- α , IL-6, IL-1 and MMP-9 in the cells, inhibited LPS-induced phosphorylation of p65 and IkB α , and suppressed nuclear translocation of p65 and DNA biding of NF- κ B in the cells. Three-week administration of hyperoside significantly decreased the clinical scores, and alleviated synovial hyperplasia, inflammatory cell infiltration and cartilage damage on mouse with collagen-induced arthritis. Hyperoside inhibited LPS-induced proliferation, migration and inflammatory responses in RA FLSs *in vitro* by suppressing activation of the NF- κ B signaling pathway, which contributes to the therapeutic effects observed in mice with collagen-induced arthritis (Jin et al., 2016).

In the study with male Sprague-Dawley rats hydroalcoholic extract of C. azarolus ssp. aronia fruit significantly lowered systolic blood pressure and phenylephrine maximal response and increased acetylcholine maximal response, serum SOD, and serum glutathione reductase in the renal artery-clipped group receiving vehicle. It was concluded that antihypertensive effects of extract may be partly due to antioxidant and NO releasing effects (Haydari et al., 2017). Ethyl acetate fraction of C. dahurica methanol extract demonstrated the greatest antioxidant activity while n-butanol fraction significantly accelerated the gastrointestinal transit in mice. Thus, this plant was suggested to serve as a good source of antioxidants and digestion-improving agents (Wang et al., 2018). Ethanolic extracts of C. orientalis leaves and berries inhibited cyclooxygenase-1 (COX-1) and 12-lipoxygenase (12-LOX), while its leaf extract showed a concentration dependent inhibition of COX-1 pathway products, 12-HHT and TXB2 (Šavikin et al., 2017).

Rhodiolae Kirliowii Radix et Rhizoma and C. pinnatifida Fructus may provide sources of potential antiviral compounds that was evidenced by hemagglutination inhibition activity on B human polyomaviruses BK KPyV and JCPyV VLPs and reducing their expression in infected cells (Chen et al., 2017). C. oxycantha buds fluid extract and the hydro-ethanolic macerate were active against thirty-due clinical strains of U. urealvticum, with MIC ranges of 15.6-250 and 15.6-62.5 µg/mL, respectively. Meanwhile, among the major purified flavonoids luteolin 3,7-diglucoside and apigenin-7-O-glucoside were the most active compounds with MICs 0.48-1.95 and 0.48-3.9 µg/mL, respectively (Bisignano et al., 2016). Ethanolic extract of C. azarolus var. eu-azarolus Maire leaves and its hexane, chloroform, and n-butanol fractions as well as ursolic acid, 3β-O-acetyl ursolic acid and quercetin 3-O-methyl ether showed variable antimicrobial activities against E. coli, P. aeruginosa, S. aureus, and C. albicans (Abu-Gharbieh and Shehab, 2017).

8. Toxicity of hawthorn preparations

Safety of *Crataegus* preparations has been proven by many studies and safe history of use. Some recent studies with *C. oxyacantha* extend the knowledge on this issue. Evaluation of *C. oxyacantha* fruit extract showed that it did not produce marked genotoxic effects at concentrations of 2.5 or 5 µg/mL in leukocytes and human liver hepatocellular carcinoma HepG2 cultured cells; however, at concentrations of 10 µg/mL or higher significant DNA damage and clastogenic/aneugenic responses were observed. The extract induced mutagenic effects in TA98 strain of *S. typhimurium* (Ames test) with metabolic activation at all tested concentrations (2.5 to 500 µg/mL). Consequently, under certain experimental conditions, the extract exerts genotoxic and clastogenic/aneugenic effects in human cells, and mutagenicity in bacterial cells (De Quadros et al., 2017). Previous *in vitro* findings were also confirmed by the study of Yonekubo et al. (2018) with C. oxyacantha extract, showing that in comet assay it did not markedly induce DNA damage in leukocytes and bone marrow cells; however, in the micronucleus test the extract produced a significant rise in micronucleated polychromatic erythrocytes (PCE) in a non-dose dependent manner. The PCE/ normochromatic erythrocytes (NCE) ratio indicated no significant cytotoxicity under applied experimental conditions, however, C. oxyacantha fruits extract exhibited weak clastogenic and/or aneugenic effects in bone marrow cells of male mice (Yonekubo et al., 2018). It suggests that prolonged or high dose use of such extracts needs to be undertaken with caution. Most recently, the electrocardiographic effects of hawthorn in healthy adult volunteers was tested and it was concluded that a single dose of oral C. oxyacantha had no effect on electrocardiographic parameters in healthy volunteers (Trexler et al., 2018).

9. Processing, quality control and other uses

Raw plant material and natural products should be carefully prepared, considering various aspects, e.g. protection of sensitive active ingredients, convenient forms for dosing and administration, standardization in terms of concentration of active ingredients and others. Some recent studies have focused on such issues.

Drying is an important process for many natural ingredients for medicines and food supplements. Aral and Bese (2016) investigated convection-drying of Crataegus spp. fruit at 50, 60 70 °C with air velocities of 0.5, 0.9 and 1.3 m/s and applied different mathematical models for the experimental data. They observed that while the shrinkage decreased, the rehydration ratio increased with increasing air temperature and air velocity. Microwave-drying, oven-drying at 50 and 70 °C, sun-drying and shade-drying were compared for the fresh fruits of C. azarolus and C. orientalis. The results showed that antioxidant capacity increased, and at the same time the TPC decreased with the temperature increase in oven drying, whereas in other drying methods (microwave, sun and shade-drying) the TPC increased; meanwhile, vitamin C content decreased in all samples. The samples dried in a microwave appeared to have the highest antioxidant capacity and it was concluded that it was the best method for preserving bioactive phytochemicals (Saadatian et al., 2016). Freeze-drying was reported to be better than hot air drying for hawthorn (C. pinnatifida) in terms of antioxidant, α-glucosidase inhibitory potential, the TPC/TFC values and color (Kwon, 2016).

Immersing of hawthorn fruits in a glycine betaine solution for 15 min at 20 °C effectively prevented fruit chilling injury and improved nutritional characteristics at low-temperature storage (Razayi et al., 2018). It significantly delayed fruit pitting development during storage at 1 °C for 20 days, increased accumulation of glycine betaine, proline, phenols, flavonoids, anthocyanins, and ascorbic acid, which was concurrent with higher SOD, catalase (CAT), ascorbate peroxidase (APX), and DPPH[•] scavenging capacity.

Emulsions prepared from the extracted from hawthorn pectin and candelilla wax were used as an edible film for treating *Pleurotus ostreatus* mushroom. This process reduced weight loss, improved firmness, and lightness, compared to untreated mushroom slices during 20 days of storage at 4 °C (Lozano-Grande et al., 2016). The functional Ag-Fe₃O₄ nanocomposites, which were prepared *via* one-pot hydrothermal method using aqueous extract of *C. pinnatifida* leaves as reducing and capping agent, exhibited

sustainable antibacterial activity against S. aureus and E. coli. The nanocomposites can be easily separated from the medium by a magnet and continues to exhibit recyclable antibacterial activity (Li and Yang, 2016). Tonic wine was produced by maceration of hawthorn fruits in red wine; as a result its TPC and antioxidant capacity remarkably increased (Muresan et al., 2016). Effects of 2,4-dichlorophenoxyacetic acid (2,4-D) combined to 6-benzylaminopurine (BAP) on callus induction, TPC and ascorbic acid production, as well as antioxidant activities in C. azarolus L. var. aronia leaf tissue cultures were assessed and it was found that the use of high level of 2,4-D over BAP could enhance the quality even more than the quantity of bioactive compounds in hawthorn leaf callus (Chaabani et al., 2015). The microcapsules containing 751 mg/g of C. monogyna bark procyanidins, mainly (-)-epicatechin, dimer B2, and trimer C1, were produced by spray drying using inulin and maltodextrin as encapsulants (Wyspiańska et al., 2017). The addition of C. monogyna phenolic-rich extracts significantly improved the oxidative stability of cooked pork patties as measured by TBARS and hexanal production, and the degree of consumer satisfaction regarding product's odor perception (Akcan et al, 2017). Ethanolic extract of C. monogyna berry was more effective than synthetic antioxidant BHA in reducing lipid oxidation and protein degradation, as well as for maintaining firmness and consistency of minced pork during 6 days of refrigeration at 4 °C (Papuc et al., 2018).

For quality control purposes, HPLC analysis was established for chlorogenic acid and hyperoside, which are among the major compounds in Crataegi fructus; the average contents (w/w %) of these compounds in the 31 batches of products from Korea and China were 0.0438 and 0.0416%, and 0.0399 and 0.0325%, respectively (Wen et al., 2016). An accurate and advanced method has been established for the simultaneous determination of chlorogenic acid, vitexin and its 4"-O-glucoside, 2"-O-rhamnoside, orientoside, rutin and hyperoside in C. pinnatifida leaves. Under optimized conditions the yields of these phytochemicals were 0.46, 0.30, 0.38, 4.37, 0.033, 0.036, and 1.19 mg/g, respectively, while DPPH' and ABTS'+ scavenging and reducing power of extract were 0.69 mg/mL (IC₅₀), 0.86 mM TE, and 0.24 mg/mL (IC₅₀), respectively (Luo et al., 2015). A simple, sensitive liquid chromatography-tandem mass spectrometry (LC-MS/MS) method, which may serve for evaluating the pharmacokinetics, was developed and fully validated for the simultaneous determination of rutin, vitexin and its glycosides, hyperoside, shanyenoside in rat plasma after intravenous administration of hawthorn leaves flavonoids using lysionotin as an internal standard (Zhu et al., 2015).]

10. Conclusion

This review clearly shows that hawthorns remain an interesting and important topic both from the scientific investigation and practical application points of view. Recent studies of *Crataegus* spp. were focused on various aspects, including phytochemical characterization of different plant anatomical parts and drug forms, variations of bioactive compounds and antioxidant activities between plant species and cultivars from different origins, health benefits and some analytical, quality control and processing issues. Identification of new bioactive phytochemicals, particularly those belonging to non-phenolic structures, as well as intensive attempts for explaining the mechanisms in the treatment of various diseases and disorders with hawthorn preparations, extracts, separated fractions and purified compounds may be recognized as important required studies that would significantly complement the existing knowledge on *Crataegus* spp. in particular and natural products in general. In addition, the findings of recent years provide new ideas and suggest the trends for further studies of *Crataegus* spp.

References

- Abuashwashi, M.A., Palomino, O.M., and Gómez-Serranillos, M.P. (2016). Geographic origin influences the phenolic composition and antioxidant potential of wild *Crataegus monogyna* from Spain. Pharm. Biol. 54(11): 2708–2713. doi:10.1080/13880209.2016.1179769.
- Abu-Gharbieh, E., and Shehab, N.G. (2017). Therapeutic potentials of Crataegus azarolus var. eu-azarolus Maire leaves and its isolated compounds. BMC Complement. Altern. Med. 17(1): 218. doi:10.1186/ s12906-017-1729-9.
- Agalou, A., Thrapsianiotis, M., Angelis, A., Papakyriakou, A., Skaltsounis, A.L., Aligiannis, N., and Beis, D. (2018). Identification of novel melanin synthesis inhibitors from *Crataegus pycnoloba* using an *in vivo* Zebrafish phenotypic assay. Front. Pharmacol. 9: 265. doi:10.3389/ fphar.2018.00265.
- Akcan, T., Estévez, M., Rico, S., Ventanas, S., and Morcuende, D. (2017). Hawberry (*Crataegus monogyna* Jaqc.) extracts inhibit lipid oxidation and improve consumer liking of ready-to-eat (RTE) pork patties. J. Food Sci. Tech. Mys. 54(5): 1248–1255. doi:10.1007/s13197-017-2578-8.
- Aksoy-Sagirli, P., Yilmaz-Ozden, T., Ozsoy, N., Celik, B.O., Kultur, S., and Melikoglu, G. (2017). *In vitro* biological effects of *Crataegus microphylla* C. Koch. Indian J. Tradit. Know. 16(2): 189–196.
- Al Humayed, S. (2017). Protective and therapeutic effects of *Crataegus aronia* in non-alcoholic fatty liver disease. Arch. Physiol. Biochem. 123(1): 23–30. doi:10.1080/13813455.2016.1205097.
- Al Humayed, S., Eid, R.A., Shatoor, A.S., Haidara, M.A., Zaki, M.S.A., and Al-Ani, B. (2017). Differential therapeutic effects of *Crataegus aronia* and simvastatin on the hepatocyte ultrastructure in hepatic steatosis. Int. J. Morphol. 35(2): 578–583. doi:10.4067/S0717-95022017000200032.
- Ali, M., Muhammad, S., Shah, M.R., Khan, A., Rashid, U., Farooq, U., Ullah, F., Sadiq, A., Ayaz, M., Ali, M., Ahmad, M., and Latif, A. (2017). Neurologically potent molecules from *Crataegus oxyacantha*; Isolation, anticholinesterase inhibition, and molecular docking. Front. Pharmacol. 8: 327. doi:10.3389/fphar.2017.00327.
- Çalişkan, O. (2015). Chapter 55 Mediterranean hawthorn fruit (*Crataegus*) species and potential usage. The Mediterranean Diet. An Evidence-Based Approach. In: Preedy, V.R., and Watson, R.R. (Ed.). Academic Press, London, pp. 621–628.
- Alirezalu, A., Salehi, P., Ahmadi, N., Sonboli, A., Aceto, S., Maleki, H.H., and Ayyari, M. (2018). Flavonoids profile and antioxidant activity in flowers and leaves of hawthorn species (*Crataegus* spp.) from different regions of Iran. Int. J. Food Prop. 21(1): 452–470. doi:10.1080/1094 2912.2018.1446146.
- Alp, H., Soner, B.C., Baysal, T., and Sahin, A.S. (2015). Protective effects of hawthorn (*Crataegus oxyacantha*) extract against digoxin-induced arrhythmias in rats. Anatolian J. Cardiol. 15(12): 970–975. doi:10.5152/akd.2014.5869.
- Aral, S., and Bese, A.V. (2016). Convective drying of hawthorn fruit (*Crataegus* spp.): Effect of experimental parameters on drying kinetics, color, shrinkage, and rehydration capacity. Food Chem. 210: 577–584. doi:10.1016/j.foodchem.2016.04.128.
- Arslan, R., Bektas, N., Bor, Z., and Sener, E. (2015). Evaluation of the antithrombotic effects of *Crataegus monogyna* and *Crataegus davisii* in the carrageenan-induced tail thrombosis model. Pharm. Biol. 53(2): 275–279. doi:10.3109/13880209.2014.914957.
- Šavikin, K.P., Krstić-Milošević, D.B., Menković, N.R., Beara, I.N., Mrkonjić, Z.O., and Pljevljakušić, D.S. (2017). *Crataegus orientalis* leaves and berries: Phenolic profiles, antioxidant and anti-inflammatory activity. Nat. Prod. Commun. 12(2): 159–162.
- Bae, Y., and Kim, Y.K. (2017). Evaluation of biological activity on hawthorn tree (*Crataegus pinnatifida*) extracts. J. Korean Wood Sci. Technol. 45(3): 317–326.
- Bekbolatova, E., Kukula-Koch, W., Baj, T., Stasiak, N., Ibadullayeva, G.,

Koch, W., Glowniak, K., Tulemissov, S., Sakipova, Z., and Boylan, F. (2018). Phenolic composition and antioxidant potential of different organs of Kazakh *Crataegus almaatensis* Pojark: A comparison with the European *Crataegus oxyacantha* L. flowers. Open Chem. 16(1): 415–426. doi:10.1515/chem-2018-0048.

- Belkhir, M., Dhaouadi, K., Rosa, A., Atzeri, A., Nieddu, M., Tuberoso, C.I.G., Rescigno, A., Amri, M., and Fattouch, S. (2016). Protective effects of azarole polyphenolic extracts against oxidative damage using *in vitro* biomolecular and cellular models. Ind. Crops Prod. 86: 239–250. doi:10.1016/j.indcrop.2016.04.003.
- Belz, G.G., and Loew, D. (2003). Dose-response related efficacy in orthostatic hypotension of a fixed combination of D-camphor and an extract from fresh *Crataegus* berries and the contribution of the single components. Phytomedicine 10: 61–67. doi:10.1078/1433-187X-00303.
- Benabderrahmane, W., Lores, M., Lamas, J.P., and Benayache, S. (2018). Matrix solid-phase dispersion as a tool for phytochemical and bioactivities characterisation: *Crataegus oxyacantha* L. A case study. Nat. Prod. Res. 32(10): 1220–1223. doi:10.1080/14786419.2017.1326040.
- Bisignano, C., Furneri, P.M., and Mandalari, G. (2016). In vitro efficacy of Crataegus oxycantha L. (hawthorn) and its major components against ATCC and clinical strains of Ureaplasma urealyticum. Adv. Microbiol. 6(12): 909–916. doi:10.4236/aim.2016.612085.
- Bjorklund, G., Dadar, M., Martins, N., Chirumbolo, S., Goh, B.H., Smetanina, K., and Lysiuk, R. (2018). Brief challenges on medicinal plants: An eye-opening look at ageing-related disorders. Basic Clin. Pharmacol. Toxicol. 122(6): 539–558. doi:10.1111/bcpt.12972.
- Bura, F.T., Firuzja, R.A., and Nemati, F. (2016). Cytotoxic effect of the flower and leaf bud extract of *Crataegus Microphylla* C. Koch on hela cell line. IIOAB J. 7: 214–218.
- Cakir, S., Orallar, H., Cetinkaya, A., Kayacan, Y., Onal, A.C., Yildirim, A., Beyazcicek, E., Benek, S., Ozkan, M., Ince, O.B., and Okur, N. (2016). Ameliorating effect of hawthorn (*Crataegus oxyacantha*) and physical exercise on acute penicillin induced seizures in gerbils. Afr. J. Tradit. Complement. Altern. Med., Nigeria 13(2): 223–228. doi:10.4314/ aitcam.v13i2.26.
- Cervantes-Paz, B., Ornelas-Paz, J.D., Gardea-Béjar, A.A., Yahia, E.M., Rios-Velasco, C., Zamudio-Flores, P.B., Ruiz-Cruz, S., and Ibarra-Junquera, V. (2018). Phenolic compounds of hawthorn (*Crataegus* spp.): Their biological activity associated to the protection of human health. Rev. Fitotec. Mex. 41(3): 339–349.
- Chaabani, G., Tabart, J., Kevers, C., Dommes, J., Khan, M.I., Zaoui, S., Chebchoub, L., Lachaal, M., and Karray-Bouraoui, N. (2015). Effects of 2,4-dichlorophenoxyacetic acid combined to 6-Benzylaminopurine on callus induction, total phenolic and ascorbic acid production, and antioxidant activities in leaf tissue cultures of *Crataegus azarolus* L. var. aronia. Acta Physiol. Plant. 37(2): 16. doi:10.1007/s11738-014-1769-4.
- Chang, W.T., Dao, J., and Shao, Z.H. (2005). Hawthorn: Potential roles in cardiovascular disease. Am. J. Chin. Med. 33(1): 1–10. doi:10.1142/ S0192415X05002606.
- Chen, L.L., Zhang, B., Shan, S.Q., and Zhao, X. (2016). Neuroprotective effects of vitexin against isoflurane-induced neurotoxicity by targeting the TRPV1 and NR2B signaling pathways. Mol. Med. Rep. 14(6): 5607–5613. doi:10.3892/mmr.2016.5948.
- Chen, Z.Y., Peng, C., Jiao, R., Wong, Y.M., Yang, N., and Huang, Y. (2009). Anti-hypertensive nutraceuticals and functional foods. J. Agric. Food Chem. 57(11): 4485–4499. doi:10.1021/jf900803r.
- Chen, S.Y., Teng, R.H., Wang, M.L., Chen, P.L., Lin, M.C., Shen, C.H., Chao, C.N., Chiang, M.K., Fang, C.Y., and Chang, D.C. (2017). *Rhodiolae Kirliowii Radix et Rhizoma* and *Crataegus pinnatifida Fructus* extracts effectively inhibit BK virus and JC virus infection of host cells. Evid. Based Complement. Alternat. Med. 2017: 5620867.
- Cheng, Q., Zhang, X.F., Wang, O., Liu, J., Cai, S.B., Wang, R.J., Zhou, F., and Ji, B.P. (2015). Anti-diabetic effects of the ethanol extract of a functional formula diet in mice fed with a fructose/fat-rich combination diet. J. Sci. Food Agric. 95(2): 401–408. doi:10.1002/jsfa.6737.
- Cui, K., Zhang, S.B., Jiang, X., and Xie, W.D. (2016). Novel synergic antidiabetic effects of *Astragalus* polysaccharides combined with *Crataegus* flavonoids *via* improvement of islet function and liver metabolism. Mol. Med. Rep. 13(6): 4737–4744. doi:10.3892/mmr.2016.5140.
- Dahmer, S., and Scott, E. (2010). Health effects of hawthorn. Am. Fam.

Physician. 81(4): 465-468.

- Dallak, M. (2018). Crataegus aronia enhances sperm parameters and preserves testicular architecture in both control and non-alcoholic fatty liver disease-induced rats. Pharm. Biol. 56(1): 535–547. doi:10.1080 /13880209.2018.1523934.
- Dennehy, C. (2001). Botanicals in cardiovascular health. Clin. Obstet. Gynecol. 44(4): 814–823. doi:10.1097/00003081-200112000-00019.
- De Quadros, A.P.O., Mazzeo, D.E.C., Marin-Morales, M.A., Perazzo, F.F., Rosa, P.C.P., and Maistro, E.L. (2017). Fruit extract of the medicinal plant *Crataegus oxyacantha* exerts genotoxic and mutagenic effects in cultured cells. J. Toxicol. Environ. Health A. 80(3): 161–170. doi:10 .1080/15287394.2016.1272517.
- Dong, Y., Liao, J.Q., Yao, K.W., Jiang, W.R., and Wang, J. (2017). Application of traditional Chinese Medicine in treatment of atrial fibrillation. Evid. Based Complement. Alternat. Med. 2017: 1381732.
- Dong, P.Z., Pan, L.L., Zhang, X.T., Zhang, W.W., Wang, X., Jiang, M.X., Chen, Y.L., Duan, Y.J., Wu, H.H., Xu, Y.T., Zhang, P., and Zhu, Y. (2017). Hawthorn (*Crataegus pinnatifida* Bunge) leave flavonoids attenuate atherosclerosis development in apoE knock-out mice. J. Ethnopharmacol. 198: 479–488. doi:10.1016/j.jep.2017.01.040.
- Dou, D.Q., Leng, P.S., Li, Y.H., Zeng, Y., and Sun, Y.X. (2015). Comparative study of antioxidant compounds and antiradical properties of the fruit extracts from three varieties of *Crataegus pinnatifida*. J. Food Sci. Technol. 52(1): 430–436. doi:10.1007/s13197-013-0954-6.
- Ebrahimzadeh, M.A., Khalili, M., Jafari, N., Zareh, G., Farzin, D., and Amin, G. (2018). Antihypoxic activities of *Crataegus pentaegyn* and *Crataegus microphylla* fruits-an *in vivo* assay. Braz. J. Pharm. Sci. 54(2): .
- Edwards, J.E., Paula, P.N., Brown, N., Talent, N., Dickinson, T.A., and García-Mateos and Shipley, P.R. (2012). A review of the chemistry of the genus *Crataegus*. Phytochem. 79: 5–26. doi:10.1016/j.phyto-chem.2012.04.006.
- Elaine, W., Wang, Y.P., Ken, C., Wai, L.H., Man, K.C., Man, L.K., Chung, L.P., Choly, Y., and San, L.C.B. (2018). An *in vitro* and *in vivo* study of a 4-herb formula on the management of diet-induced metabolic syndrome. Phytomedicine 42: 112–125. doi:10.1016/j.phymed.2018.03.028.
- Ercisli, S., Yanar, M., Sengul, M., Yildiz, H., Topdas, E.F., Taskin, T., Zengin, Y., and Yilmaz, K.U. (2015). Physico-chemical and biological activity of hawthorn (*Crataegus* spp. L.) fruits in Turkey. Acta Sci. Pol. Hortorum Cultus 14(1): 83–93.
- Fei, C.H., Dai, H., Wu, X.Y., Li, L., Lu, T.L., Li, W.D., Cai, B.C., Yin, W., and Yin, F.Z. (2018). Quality evaluation of raw and processed Crataegi Fructus by color measurement and fingerprint analysis. J. Sep. Sci. 41(2): 582–589. doi:10.1002/jssc.201700575.
- Fong, H., and Bauman, J. (2002). Alternative medicines for cardiovascular diseases: Hawthorn. J. Cardiovasc. Nurs. 16: 1–8.
- Fu, T., Wang, L., Jin, X.N., Sui, H.J., Liu, Z., and Jin, Y. (2016). Hyperoside induces both autophagy and apoptosis in non-small cell lung cancer cells in vitro. Acta Pharmacol. Sin. 37(4): 505–518. doi:10.1038/ aps.2015.148.
- Fuchs, S., Bischoff, I., Willer, E.A., Bräutigam, J., Bubik, M.F., Erdelmeier, C.A.J., Koch, E., Faleschini, M.T., De Mieri, M., Bauhart, M., Zahler, S., Hensel, A., Hamburger, M., Potterat, O., and Fürst, R. (2017). The dual edema-preventing molecular mechanism of the *Crataegus* extract WS 1442 can be assigned to distinct phytochemical fractions. Planta Med. 83(8): 701–709.
- Ganie, S.A., Dar, T.A., Zargar, S., Bhat, A.H., Dar, K.B., Masood, A., and Zargar, M.A. (2016). *Crataegus songarica* methanolic extract accelerates enzymatic status in kidney and heart tissue damage in albino rats and its in vitro cytotoxic activity. Pharm. Biol. 54(7): 1246–1254. doi:10.3 109/13880209.2015.1066398.
- Gao, P.Y., Li, L.Z., Liu, K.C., Sun, C., Sun, X., Wu, Y.N., and Song, S.J. (2017). Natural terpenoid glycosides with *in vitro/vivo* antithrombotic profiles from the leaves of *Crataegus pinnatifida*. RSC Adv. 7(76): 48466– 48474. doi:10.1039/C7RA10768D.
- González-Jiménez, F.E., Salazar-Montoya, J.A., Calva-Calva, G., and Ramos-Ramírez, E.G. (2018). Phytochemical characterization, *in vitro* antioxidant activity, and quantitative analysis by micellar electrokinetic chromatography of hawthorn (*Crataegus pubescens*) fruit. J. Food Qual. 2018: 2154893.
- Granato, D., Shahidi, F., Wrolstad, R., Kilmartin, P., Melton, L.D., Hidalgo, F.J., Miyashita, K., van Camp, J., Alasalvar, C., Ismail, A.B., Elmore, S.,

Birch, G.G., Charalampopoulos, D., Astley, S.B., Pegg, R., Zhou, P., and Finglas, P. (2018). Antioxidant activity, total phenolics and flavonoids contents: Should we ban in vitro screening methods? Food Chem. 264: 471–475. doi:10.1016/j.foodchem.2018.04.012.

- Guo, R., Lin, B., Shang, X.Y., Zhou, L., Yao, G.D., Huang, X.X., and Song, S.J. (2018). Phenylpropanoids from the fruit of *Crataegus pinnatifida* exhibit cytotoxicity on hepatic carcinoma cells through apoptosis induction. Fitoterapia 127: 301–307. doi:10.1016/j.fitote.2018.03.003.
- Halver, J., Garcia, C.C., Willems, E., and Schade, D. (2015). Crataegus ssp promotes late-stage cardiac differentiation and regeneration. Planta Med. 81(16): 1509–1509. Meeting Abstract: PW-61.
- Han, F., Guo, Y.P., Gu, H.Y., Li, F.L., Hu, B.Z., and Yang, L. (2016a). Application of alkyl polyglycoside surfactant in ultrasonic-assisted extraction followed by macroporous resin enrichment for the separation of vitexin-2"-O-rhamnoside and vitexin from *Crataegus pinnatifida* leaves. J. Chromatogr. B 1012: 69–78.
- Han, X., Li, W.F., Huang, D., and Yang, X.B. (2016b). Polyphenols from hawthorn peels and fleshes differently mitigate dyslipidemia, inflammation and oxidative stress in association with modulation of liver injury in high fructose diet-fed mice. Chem. Biol. Interact. 257: 132–140. doi:10.1016/j.cbi.2016.08.002.
- Hatipoglu, M., Saglam, M., Koseoglu, S., Koksal, E., Keles, A., and Esen, H.H. (2015). The effectiveness of *Crataegus orientalis* M Bieber. (Hawthorn) extract administration in preventing alveolar bone loss in rats with experimental periodontitis. PLoS ONE 10(6): e0128134. doi:10.1371/journal.pone.0128134.
- Haydari, M.R., Panjeshahin, M.R., Mashghoolozekr, E., and Nekooeian, A.A. (2017). Antihypertensive effects of hydroalcoholic extract of *Crataegus Azarolus* subspecies *Aronia* fruit in rats with renovascular hypertension: An experimental mechanistic study. Iran. J. Med. Sci. 42(3): 266–274.
- He, M., Min, J.W., Kong, W.L., He, X.H., Li, J.X., and Peng, B.W. (2016). A review on the pharmacological effects of vitexin and isovitexin. Fitoterapia 115: 74–85. doi:10.1016/j.fitote.2016.09.011.
- Hellenbrand, N., Sendker, J., Lechtenberg, M., Petereit, F., and Hensel, A. (2015). Isolation and quantification of oligomeric and polymeric procyanidins in leaves and flowers of hawthorn (*Crataegus* spp.). Fitoterapia. 104: 14–22. doi:10.1016/j.fitote.2015.04.010.
- Holubarsch, C.J.F., Colucci, W.S., and Eha, J. (2018). Benefit-risk assessment of *Crataegus* extract WS 1442: An evidence-based review. Am. J. Cardiovasc. Drugs 18(1): 25–36. doi:10.1007/s40256-017-0249-9.
- Houston, M.C. (2005). Nutraceuticals, vitamins, antioxidants, and minerals in the prevention and treatment of hypertension. Prog. Cardiovasc. Dis. 47(6): 396–449. doi:10.1016/j.pcad.2005.01.004.
- Hu, M., Li, F.M., and Wang, W.D. (2018). Vitexin protects dopaminergic neurons in MPTP-induced Parkinson's disease through PI3K/Akt signaling pathway. Drug Des. Dev. Ther. 12: 565–573. doi:10.2147/DDDT. S156920.
- Huang, X.X., Bai, M., Zhou, L., Lou, L.L., Liu, Q.B., Zhang, Y., Li, L.Z., and Song, S.J. (2015a). Food byproducts as a new and cheap source of bioactive compounds: Lignans with antioxidant and anti-inflammatory properties from *Crataegus pinnatifida* seeds. J. Agric. Food Chem. 63(32): 7252–7260. doi:10.1021/acs.jafc.5b02835.
- Huang, X.X., Liu, Q.B., Zhou, L., Liu, S., Cheng, Z.Y., Sun, Q., Li, L.Z., and Song, S.J. (2015b). The antioxidant and tyrosinase-inhibiting activities of 8-O-4' neolignans from *Crataegus pinnatifida* Seeds. Rec. Nat. Prod. 9(3): 305–311.
- Huang, X.X., Ren, Q., Song, X.Y., Zhou, L., Yao, G.D., Wang, X.B., and Song, S.J. (2018a). Seven new sesquineolignans isolated from the seeds of hawthorn and their neuroprotective activities. Fitoterapia 125: 6–12. doi:10.1016/j.fitote.2017.12.010.
- Huang, X.X., Xu, Y., Bai, M., Zhou, L., Song, S.J., and Wang, X.B. (2018b). Lignans from the seeds of Chinese hawthorn (*Crataegus pinnatifida* var. *major* NEBr.) against β-amyloid aggregation. Nat. Prod. Res. 32(14): 1706–1713. doi:10.1080/14786419.2017.1399378.
- Hwang, E., Park, S.Y., Yin, C.S., Kim, H.T., Kim, Y.M., and Yi, T.H. (2017). Antiaging effects of the mixture of Panax ginseng and *Crataegus pinnatifida* in human dermal fibroblasts and healthy human skin. J. Ginseng Res. 41(1): 69–77. doi:10.1016/j.jgr.2016.01.001.
- Jalaly, L., Sharifi, G., Faramarzi, M., Nematollahi, A., Rafieian-Kopaei, M., Amiri, M., and Moattar, F. (2015). Comparison of the effects of *Cra*-

taegus oxyacantha extract, aerobic exercise and their combination on the serum levels of ICAM-1 and E-Selectin in patients with stable angina pectoris. Daru J. Pharm. Sci. 23: 54. doi:10.1186/s40199-015-0137-2.

- Jin, X.N., Yan, E.Z., Wang, H.M., Sui, H.J., Liu, Z., Gao, W., and Jin, Y. (2016). Hyperoside exerts anti-inflammatory and antiarthritic effects in LPSstimulated human fibroblast-like synoviocytes in vitro and in mice with collagen-induced arthritis. Acta Pharmacol. Sin. 37(5): 674–686. doi:10.1038/aps.2016.7.
- Jung, D., and Shim, J. (2017). Crataegus pinnatifida extract increases lifespan of Drosophila melanogaster. Mol. Biol. Cell 28: Meeting Abstract: P3476.
- Jurikova, T., Sochor, J., Rop, O., Mlcek, J., Balla, S., Szekeres, L., Adam, V., and Kizek, R. (2012). Polyphenolic profile and biological activity of Chinese hawthorn (*Crataegus pinnatifida* BUNGE) fruits. Molecules 17: 14490–14509. doi:10.3390/molecules171214490.
- Kalantari, H., Hemmati, A.A., Goudarzi, M., Forouzandeh, H., Kalantar, M., Aghel, N., Aslani, M.K., and Ehsan, T.S. (2016). Healing Effect of Hawthorn (Crataegus pontica C. Koch) Leaf Extract in Dermal Toxicity Induced by T-2 Toxin in Rabbit. Jundishapur J. Nat. Pharm. Prod. 11(3).
- Kallassy, H., Fayyad-Kazan, M., Makki, R., El-Makhour, Y., Hamade, E., Rammal, H., Leger, D.Y., Sol, V., Fayyad-Kazan, H., Liagre, B., and Badran, B. (2017). Chemical composition, anti-oxidant, anti-inflammatory, and antiproliferative activities of the plant Lebanese *Crataegus azarolus* L. Med. Sci. Monit. Basic Res. 23: 270–284. doi:10.12659/MSMBR.905066.
- Kang, I. (2015). Anti-diabetic and anti-oxidative activities of extracts from *Crataegus pinnatifida*. J. East Asian Soc. Diet. Life 25(2): 270–277. doi:10.17495/easdl.2015.4.25.2.270.
- Kanmanthareddy, A., Reddy, M., Ponnaganti, G., Sanjani, H.P., Koripalli, S., Adabala, N., Buddam, A., Janga, P., Lakkireddy, T., Bommana, S., Vallakati, A., Atkins, D., and Lakkireddy, D. (2015). Alternative medicine in atrial fibrillation treatment-Yoga, acupuncture, biofeedback and more. J. Thorac. Dis. 7(2): 185–192.
- Kazuma, K., Isobe, Y., Asahina, H., Nehira, T., Satake, M., and Konno, K. (2016). Crataegusins A and B, new flavanocoumarins from the dried fruits of *Crataegus pinnatifida* var. major (Rosaceae). Nat. Prod. Commun. 11(7): 965–969.
- Koch, E., and Malek, F.A. (2011). Standardized extracts from hawthorn leaves and flowers in the treatment of cardiovascular disorders - Preclinical and clinical studies. Planta Med. 77(11): 1123–1128. doi:10. 1055/s-0030-1270849.
- Kowalski, R., Kowalska, G., Kałwa, K., and Sujka, M. (2018). Essential oil composition of hawthorn *Crataegus monogyna* inflorescence. Chem. Nat. Comp. 54(5): 995–997. doi:10.1007/s10600-018-2533-6.
- Kumar, D., Arya, V., Bhat, Z.A., Khan, N.A., and Prasad, D.N. (2012). The genus *Crataegus*: Chemical and pharmacological perspectives. Rev. Bras. Farmacogn. 22(5): 1187–1200. doi:10.1590/S0102-695X2012005000094.
- Kwon, Y.R. (2016). Antioxidant abilities and physiological properties of dried Haw extracts prepared using different drying methods. Korean J. Food Preserv. 23(2): 246–251. doi:10.11002/kjfp.2016.23.2.246.
- Lakache, Z., Tigrine-Kordjani, N., Tigrine, C., Aliboudhar, H., and Kameli, A. (2016). Phytochemical screening and antioxidant properties of methanolic extract and different fractions of *Crataegus azarolus* leaves and flowers from Algeria. Intern. Food Res. J. 23(4): 1576–1583.
- Lee, Y.H., Kim, Y.S., Song, M., Lee, M., Park, J., and Kim, H. (2015). A herbal formula HT048, *Citrus unshiu* and *Crataegus pinnatifida*, prevents obesity by inhibiting adipogenesis and lipogenesis in 3T3-L1 preadipocytes and HFD-induced obese rats. Molecules 20(6): 9656–9670. doi:10.3390/molecules20069656.
- Lee, Y.H., Jin, B., Lee, S.H., Song, M., Bae, H., Min, B.J., Park, J., Lee, D., and Kim, H. (2016). Herbal formula HT048 attenuates diet-induced obesity by improving hepatic lipid metabolism and insulin resistance in obese rats. Molecules. 21(11): 1424. doi:10.3390/molecules21111424.
- Li, W.Q., Hu, Q.P., and Xu, J.G. (2015). Changes in physicochemical characteristics and free amino acids of hawthorn (*Crataegus pinnatifida*) fruits during maturation. Food Chem. 175: 50–56. doi:10.1016/j. foodchem.2014.11.125.
- Li, L.Z., Gao, P.Y., Song, S.J., Yuan, Y.Q., Liu, C.T., Huang, X.X., and Liu, Q.B.

(2015). Monoterpenes and flavones from the leaves of *Crataegus pinnatifida* with anticoagulant activities. J. Funct. Foods. 12: 237–245. doi:10.1016/j.jff.2014.11.012.

- Li, Z.P., Xu, J.Y., Zheng, P.Y., Xing, L.J., Shen, H.Y., Yang, L.L., Zhang, L., and Ji, G. (2015). Hawthorn leaf flavonoids alleviate nonalcoholic fatty liver disease by enhancing the adiponectin/AMPK pathway. Int. J. Clin. Exp. Med. 8(10): 17295–17307.
- Li, W.H., and Yang, N. (2016). Green and facile synthesis of Ag-Fe₃O₄ nanocomposites using the aqueous extract of *Crataegus pinnatifida* leaves and their antibacterial performance. Mater. Lett. 162: 157– 160. doi:10.1016/j.matlet.2015.09.064.
- Lim, D.W., Han, T., Jung, J., Song, Y., Um, M.Y., Yoon, M., Kim, Y.T., Cho, S., Kim, I.H., Han, D., Lee, C., and Lee, J. (2018). Chlorogenic acid from hawthorn berry (*Crataegus pinnatifida* Fruit) prevents stress hormone-induced depressive behavior, through monoamine oxidase Breactive oxygen species signaling in hippocampal astrocytes of mice. Mol. Nutr. Food Res. 62(15): 1800029. doi:10.1002/mnfr.201800029.
- Linares-Garcia, J.A., Ramos-Ramirez, E.G., and Salazar-Montoya, J.A. (2015). Viscoelastic properties and textural characterisation of high methoxyl pectin of hawthorn (*Crataegus pubescens*) in a gelling system. Int. J. Food Sci. Technol. 50(6): 1484–1493. doi:10.1111/ ijfs.12792.
- Liu, S.W., Chang, X.D., Liu, X.F., and Shen, Z.W. (2016). Effects of pretreatments on anthocyanin composition, phenolics contents and antioxidant capacities during fermentation of hawthorn (*Crataegus pinnatifida*) drink. Food Chem. 212: 87–95. doi:10.1016/j.foodchem.2016.05.146.
- Liu, C.Q., and Huang, Y. (2016). Chinese herbal medicine on cardiovascular diseases and the mechanisms of action. Front. Pharmacol. 7: 469.
- Liu, S.W., You, L., Zhao, Y.X., and Chang, X.D. (2018). Hawthorn polyphenol extract inhibits UVB-induced skin photoaging by regulating MMP expression and Type I procollagen production in mice. J. Agric. Food Chem. 66(32): 8537–8546. doi:10.1021/acs.jafc.8b02785.
- Liu, S.W., Zhang, X., You, L., Guo, Z.Y., and Chang, X.D. (2018). Changes in anthocyanin profile, color, and antioxidant capacity of hawthorn wine (*Crataegus pinnatifida*) during storage by pretreatments. LWT Food Sci. Technol. 95: 179–186. doi:10.1016/j.lwt.2018.04.093.
- Loew, D. (1997). Phytotherapy in heart failure. Phytomedicine 4(3): 267– 271. doi:10.1016/S0944-7113(97)80080-3.
- Lozano-Grande, M.A., Valle-Guadarrama, S., Aguirre-Mandujano, E., Lobato-Calleros, C.S.O., and Huelitl-Palacios, F. (2016). Films based on hawthorn (*Crataegus* spp.) fruit pectin and candelilla wax emulsions: Characterization and application on *Pleurotus ostreatus*. Agrociencia-Mexico. 50(7): 849–866.
- Luo, M., Hu, J.Y., Song, Z.Y., Jiao, J., Mu, F.S., Ruan, X., Gai, Q.Y., Qiao, Q., Zu, Y.G., and Fu, Y.J. (2015). Optimization of ultrasound-assisted extraction (UAE) of phenolic compounds from Crataegus pinnatifida leaves and evaluation of antioxidant activities of extracts. RSC Adv. 5(83): 67532–67540. doi:10.1039/C5RA07445B.
- Luo, C., Ke, Y.T., Yuan, Y.Y., Zhao, M., Wang, F.Y., Zhang, Y.S., and Bu, S.Z. (2016). A novel herbal treatment reduces depressive-like behaviors and increases brain-derived neurotrophic factor levels in the brain of type 2 diabetic rats. Neuropsychiatr. Dis. Treat. 12: 3051–3059. doi:10.2147/NDT.S117337.
- Luo, L., Zhen, L.F., Xu, Y.T., Yang, Y.X., Feng, S.X., Wang, S.M., and Liang, S.W. (2016). H-1 NMR-based metabonomics revealed protective effect of Naodesheng bioactive extract on ischemic stroke rats. J. Ethnopharmacol. 186: 257–269. doi:10.1016/j.jep.2016.03.059.
- Luo, M., Yang, X., Hu, J.Y., Jiao, J., Mu, F.S., Song, Z.Y., Gai, Q.Y., Qiao, Q., Ruan, X., and Fu, Y.J. (2016). Antioxidant properties of phenolic compounds in renewable parts of *Crataegus pinnatifida* inferred from seasonal variations. J. Food Sci. 81(5): C1102–C1109. doi:10.1111/1750-3841.13291.
- Mahmud, S.A., Al-Habib, O.A.M., Bugoni, S., Clericuzio, M., and Vidari, G. (2016). A new ursane-type triterpenoid and other constituents from the leaves of *Crataegus azarolus* var. *aronia*. Nat. Prod. Commun. 11(11): 1637–1639.
- Miao, J., Li, X., Fan, Y.Y., Zhao, C.C., Mao, X.H., Chen, X.T., Huang, H.H., and Gao, W.Y. (2016). Effect of different solvents on the chemical composition, antioxidant activity and α -glucosidase inhibitory activity of hawthorn extracts. Int. J. Food Sci. Technol. 51(5): 1244–1251.

doi:10.1111/ijfs.13076.

- Min, Q., Bai, Y.T., Zhang, Y.C., Yu, W., Zhang, M.L., Liu, D.Y., Diao, T.T., and Lv, W. (2017). Hawthorn leaf flavonoids protect against diabetesinduced cardiomyopathy in rats via PKC-α signaling pathway. Evid. Based Complement. Alternat. Med. 2017: 2071952.
- Moradi, M.T., Karimi, A., Alidadi, S., and Hashemi, L. (2018). In vitro antiherpes simplex virus activity, antioxidant potential and total phenolic compounds of selected Iranian medicinal plant extracts. Indian J. Tradit. Know. 17(2): 255–262.
- Mostafa, D.G., Khaleel, E.E., and Abdel-Aleem, G.A. (2018). Inhibition of the hepatic glucose output is responsible for the hypoglycemic effect of *Crataegus aronia* against type 2 diabetes mellitus in rats. Arch. Biol. Sci. 70(2): 277–287. doi:10.2298/ABS170510044M.
- Mraihi, F., Hidalgo, M., de Pascual-Teresa, S., Trabelsi-Ayadi, M., and Cherif, J.K. (2015). Wild grown red and yellow hawthorn fruits from Tunisia as source of antioxidants. Arab. J. Chem. 8(4): 570–578. doi:10.1016/j.arabjc.2014.11.045.
- Muresan, A.E., Muste, S., Petrut, G., Vlaic, R.A., Man, S.M., and Muresan,
 V. (2016). New uses of hawthorn fruits in tonic wines technology.
 Bull. Univ. Agric. Sci. Vet. Med. Cluj Napoca 73(2) 117-122: 12327.
- Mustapha, N., Bzeouich, I.M., Ghedira, K., Hennebelle, T., and Chekir-Ghedira, L. (2015). Compounds isolated from the aerial part of *Crataegus azarolus* inhibit growth of B16F10 melanoma cells and exert a potent inhibition of the melanin synthesis. Biomed. Pharmacother. 69: 139–144. doi:10.1016/j.biopha.2014.11.010.
- Mustapha, N., Mokdad-Bzeouich, I., Sassi, A., Abed, B., Ghedira, K., Hennebelle, T., and Chekir-Ghedira, L. (2016a). Immunomodulatory potencies of isolated compounds from *Crataegus azarolus* through their antioxidant activities. Tumor Biol. 37(6): 7967–7980. doi:10.1007/ s13277-015-4517-5.
- Mustapha, N., Mokdad-Bzeouich, I., Maatouk, M., Ghedira, K., Hennebelle, T., and and Chekir-Ghedira, L. (2016b). Antitumoral, antioxidant, and antimelanogenesis potencies of hawthorn, a potential natural agent in the treatment of melanoma. Melanoma Res. 26(3): 211–222. doi:10.1097/CMR.0000000000240.
- Mustapha, N., Pinon, A., Limami, Y., Simon, A., Ghedira, K., Hennebelle, T., and Chekir-Ghedira, L. (2016c). *Crataegus azarolus* leaves induce antiproliferative activity, cell cycle arrest, and apoptosis in human HT-29 and HCT-116 colorectal cancer cells. J. Cell. Biochem. 117(5): 1262–1272. doi:10.1002/jcb.25416.
- Nabavi, S.F., Habtemariam, S., Ahmed, T., Sureda, A., Daglia, M., Sobarzo-Sanchez, E., and Nabavi, S.M. (2015). Polyphenolic composition of *Crataegus monogyna* Jacq.: From chemistry to medical applications. Nutrients 7(9): 7708–7728. doi:10.3390/nu7095361.
- Nunes, M.A., Rodrigues, F., Alves, R.C., and Oliveira, M.B.P.P. (2017). Herbal products containing *Hibiscus sabdariffa* L., *Crataegus* spp., and *Panax* spp.: Labeling and safety concerns. Food Res. Int. 100: 529–540. doi:10.1016/j.foodres.2017.07.031.
- Čopra-Janićijević, A., Čulum, D., Vidic, D., Tahirović, A., Klepo, L., and Bašić, N. (2018). Chemical composition and antioxidant activity of the endemic *Crataegus microphylla* Koch subsp *malyana* K. I. Chr. & Janjić from Bosnia. Ind. Crops Prod. 113: 75–79. doi:10.1016/j.indcrop.2018.01.016.
- Ozay, C., Mammadov, R., Tasdelen, G., Karagur, E.R., and Akca, H. (2015). Potential antioxidant, antiproliferative and hepatoprotective effects of *Crataegus Meyeri*. J. Food Biochem. 39(5): 548–553. doi:10.1111/ jfbc.12161.
- Ozderin, S., Fakir, H., and Donmez, I.E. (2016). Chemical properties of hawthorn (*Crataegus* L. spp.) taxa naturally distributed in Western Anatolia part of turkey. Sumarski List. 140(7-8): 369–376. doi:10.31298/ sl.140.7-8.5.
- Pahlavan, S., Tousi, M.S., Ayyari, M., Alirezalu, A., Ansari, H., Saric, T., and Baharvand, H. (2018). Effects of hawthorn (*Crataegus pentagyna*) leaf extract on electrophysiologic properties of cardiomyocytes derived from human cardiac arrhythmia-specific induced pluripotent stem cells. FASEB J. 32(3): 1440–1451. doi:10.1096/fj.201700494RR.
- Pang, X.C., Kang, D., Fang, J.S., Zhao, Y., Xu, L.J., Lian, W.W., Liu, A.L., and Du, G.H. (2018). Network pharmacology-based analysis of Chinese herbal Naodesheng formula for application to Alzheimer's disease. CJNM. 16(1): 53–62.
- Papuc, C., Predescu, C.N., Tudoreanu, L., Nicorescu, V., and Gâjâilă, I.

(2018). Comparative study of the influence of hawthorn (*Crataegus monogyna*) berry ethanolic extract and butylated hydroxylanisole (BHA) on lipid peroxidation, myoglobin oxidation, consistency and firmness of minced pork during refrigeration. J. Sci. Food Agric. 98(4): 1346–1361. doi:10.1002/jsfa.8599.

- Park, S.-J., Kwon, S.-P., and Rha, Y.-A. (2017a). Antioxidative activities and whitening effects of ethanol extract from *Crataegus pinnatifida* Bunge Fruit. J. Korean Soc. Food Sci. Nutr. 46(10): 1158–1163.
- Park, S.-J., Kwon, S.-P., and Rha, Y.-A. (2017b). Enhancement of antioxidant activities of *Crataegus pinnatifida* Bunge fruit by ultrasonification extraction processes. J. Korean Soc. Food Sci. Nutr. 46(7): 891–895.
- Pawlaczyk-Graja, I. (2018). Polyphenolic-polysaccharide conjugates from flowers and fruits of single-seeded hawthorn (*Crataegus monogyna* Jacq.): Chemical profiles and mechanisms of anticoagulant activity. Int. J. Biol. Macromol. 116: 869–879. doi:10.1016/j.ijbiomac.2018.05.101.
- Peng, Y., Lou, L.L., Liu, S.F., Zhou, L., Huang, X.X., and Song, S.J. (2016). Antioxidant and anti-inflammatory neolignans from the seeds of hawthorn. Bioorg. Med. Chem. Lett. 26(22): 5501–5506. doi:10.1016/j. bmcl.2016.10.012.
- Phipps, J.B., O'Kennon, R.J., and Lance, R.W. (2003). Hawthorns and Medlars. Royal Horticultural Society, Cambridge U.K.
- Pittler, M.H., Schmidt, K., and Ernst, E. (2003). Hawthorn extract for treating chronic heart failure: Meta-analysis of randomized trials. Am. J. Med. 114(8): 665–674.(03)00131-1. doi:10.1016/S0002-9343(03)00131-1.
- Pliszka, B., Huszcza-Ciolkowska, G., and Wierzbicka, E. (2016). Effects of solvents and extraction methods on the content and antiradical activity of polyphenols from fruits Actinidia arguta, Crataegus monogyna, Gaultheria procumbens and Schisandra chinensis. Acta Sci. Pol. Technol. Aliment. 15(1): 57–63. doi:10.17306/J.AFS.2016.1.6.
- Qiao, A.M., Wang, Y.H., Xiang, L.M., Zhang, Z.X., and He, X.J. (2015). Novel triterpenoids isolated from hawthorn berries functioned as antioxidant and antiproliferative activities. J. Funct. Foods 13: 308–313. doi:10.1016/j.jff.2014.12.047.
- Rababa'h, A.M., Altarabsheh, S.E., Haddad, O., Deo, S.V., Obeidat, Y., and Al-Azzam, S. (2016). Hawthorn herb increases the risk of bleeding after cardiac surgery: An evidence-based approach. Heart Surg. Forum 19(4): E175–E179. doi:10.1532/hsf.1570.
- Ranjbar, K., Zarrinkalam, E., Salehi, I., Komaki, A., and Fayazi, B. (2018). Cardioprotective effect of resistance training and *Crataegus oxyacantha* extract on ischemia reperfusion-induced oxidative stress in diabetic rats. Biomed. Pharmacother. 100: 455–460. doi:10.1016/j. biopha.2018.02.021.
- Rastogi, S., Pandey, M.M., and Rawat, A.K.S. (2016). Traditional herbs: A remedy for cardiovascular disorders. Phytomedicine 23(11): 1082– 1089. doi:10.1016/j.phymed.2015.10.012.
- Razayi, F., Mahmoudi, R., Rabiei, V., Aghdam, M.S., and Soleimani, A. (2018). Glycine betaine treatment attenuates chilling injury and maintains nutritional quality of hawthorn fruit during storage at low temperature. Sci. Hortic. 233: 188–194. doi:10.1016/j.scienta.2018.01.053.
- Renda, G., Ozel, A., Barut, B., Korkmaz, B., and Yayli, N. (2018). In vitro protection by Crataegus microphylla extracts against oxidative damage and enzyme inhibition effects. Turk. J. Pharm. Sci. 15(1): 77–84.
- Rezaei-Golmisheh, A., Malekinejad, H., Asri-Rezaei, S., Farshid, A.A., and Akbari, P. (2015). Hawthorn ethanolic extracts with triterpenoids and flavonoids exert hepatoprotective effects and suppress the hypercholesterolemia-induced oxidative stress in rats. Iranian J. Basic Med. Sci. 18(7): 691–699.
- Rosa, S.I.G., Rios-Santos, F., Balogun, S.O., and Martins, D.T.D. (2016). Vitexin reduces neutrophil migration to inflammatory focus by down-regulating pro-inflammatory mediators via inhibition of p38, ERK1/2 and JNK pathway. Phytomedicine 23(1): 9–17. doi:10.1016/j. phymed.2015.11.003.
- Saadatian, M., Najda, A., and Jasour, M.S. (2016). Drying process affects bioactive compounds in hawthorn species. Acta Sci. Pol. Hortoru. 15(4): 3–16.
- Saeedi, G., Jeivad, F., Goharbari, M., Gheshlaghi, G.H., and Sabzevari, O. (2018). Ethanol extract of *Crataegus oxyacantha* L. ameliorate dietary non-alcoholic fatty liver disease in rat. Drug Res. 68(10): 553–

559. doi:10.1055/a-0579-7532.

- Sarris, J. (2007). Herbal medicines in the treatment of psychiatric disorders: A systematic review. Phytother. Res. 21(8): 703–716. doi:10.1002/ ptr.2187.
- Sayin, F.K., Altincam, M.H., and Kara, H.H. (2018). In vitro anti-diabetic activity of selected herbal teas extracted with different methods. Z. Arznei- Gewurzpfla. 23(2): 75–78.
- Schini-Kerth, B., Ribeiro, T., Auger, C., Jabeen, Q., Da Silva, G.C., Medeiros, I.A., Boehm, N., and Monassier, L. (2015). Intake of a standardized crataegus extract prevents DOCA-salt-induced hypertension, and alteration of cardiac, vascular and renal structure and function in rats: role of oxidative stress. Eur. J. Heart Fail. 17: 368–369.
- Schmitt, C.A., and Dirsch, V.M. (2009). Modulation of endothelial nitric oxide by plant-derived products. Nitric Oxide. 21(2): 77–91. doi:10.1016/j.niox.2009.05.006.
- Shahidi, F. (2004). Functional foods: Their role in health promotion and disease prevention. J. Food Sci. 69(5): R146–R149.
- Shahidi, F., and Yeo, J. (2018). Bioactivities of phenolics by focusing on suppression of chronic diseases: A review. Int. J. Mol. Sci. 19(6): 1573. doi:10.3390/ijms19061573.
- Shao, F., Gu, L.F., Chen, H.J., Liu, R.H., Huang, H.L., and Ren, G. (2016). Comparation of hypolipidemic and antioxidant effects of aqueous and ethanol extracts of *Crataegus pinnatifida* fruit in high-fat emulsion-induced hyperlipidemia rats. Pharmacogn. Mag. 12(45): 64–69. doi:10.4103/0973-1296.176049.
- Sixt, M., and Strube, J. (2018). Systematic design and evaluation of an extraction process for traditionally used herbal medicine on the example of hawthorn (*Crataegus monogyna* JACQ. Processes 6(7): 73. doi:10.3390/pr6070073.
- Stelmakienė, A., Ramanauskienė, K., Petrikaitė, V., Jakštas, V., and Briedis, V. (2016). Application of dry hawthorn (*Crataegus Oxyacantha* L.) extract in natural topical formulations. Acta Pol. Pharm. 73(4): 955–965.
- Trexler, S.E., Nguyen, E., Gromek, S.M., Balunas, M.J., and Baker, W.L. (2018). Electrocardiographic effects of hawthorn (*Crataegus oxyacantha*) in healthy volunteers: A randomized controlled trial. Phytother. Rese. 32(8): 1642–1646. doi:10.1002/ptr.6094.
- Čulum, D., Čopra-Janićijević, A., Vidic, D., Klepo, L., Tahirović, A., Bašić, N., and Maksimović, M. (2018). HPLC-ED Analysis of phenolic compounds in three Bosnian *Crataegus* species. Foods 7(5): 66. doi:10.3390/foods7050066.
- Veberic, R., Slatnar, A., Bizjak, J., Stampar, F., and Mikulic-Petkovsek, M. (2015). Anthocyanin composition of different wild and cultivated berry species. LWT Food Sci. Technol. 60(1): 509–517. doi:10.1016/j. lwt.2014.08.033.
- Veličković, J.M., Ilić, S., Mitić, S.S., Mitić, M.N., and Kostić, D.A. (2016). Comparative analysis of phenolic and mineral composition of hawthorn and blackthorn from southeast Serbia. Oxid. Commun. 39(3): 2280–2290.
- Venskutonis, P.R. (2016). Hawthorn Juice. Handbook of Functional Beverages and Human Health In: Shahidi, F., and Alasalvar, C. (Ed.). CRC Press, 311–320.
- Wang, Y.N., Zhen, Y.L., Wu, X., Jiang, Q., Li, X.L., Chen, Z.W., Zhang, G.L., and Dong, L.Y. (2015a). Vitexin protects brain against ischemia/ reperfusion injury via modulating mitogen-activated protein kinase and apoptosis signaling in mice. Phytomedicine 22(3): 379–384. doi:10.1016/j.phymed.2015.01.009.
- Wang, X.S., Hu, X.C., Chen, G.L., Yuan, X., Yang, R.N., Liang, S., Ren, J., Sun, J.C., Kong, G.Q., Gao, S.G., and Feng, X.S. (2015b). Effects of Vitexin on the Pharmacokinetics and mRNA Expression of CYP Isozymes in Rats. Phytother. Res. 29(3): 366–372. doi:10.1002/ptr.5260.
- Wang, X.R., Zhang, C.L., Peng, Y.J., Zhang, H.M., Wang, Z.G., Gao, Y., Liu, Y., and Zhang, H.L. (2018a). Chemical constituents, antioxidant and gastrointestinal transit accelerating activities of dried fruit of *Crataegus dahurica*. Food Chem. 246: 41–47. doi:10.1016/j.foodchem.2017.11.011.
- Wang, X.W., Liang, Y., Shi, J., Zhu, H.J., and Bleske, B.E. (2018b). Crataegus special extract WS 1442 effects on eNOS and microRNA 155. Planta Med. 84(15): 1094–1100. doi:10.1055/a-0601-7083.
- Wang, L.W., Yue, Z.W., Guo, M.Z., Fang, L.Y., Bai, L., Li, X.Y., Tao, Y.Q., Wang, S.Y., Liu, Q., Zhi, D.X., and Zhao, H. (2016). Dietary flavonoid hypero-

side induces apoptosis of activated human LX-2 hepatic stellate cell by suppressing canonical NF- κ B signaling. Biomed Res. Int. 2016: 1068528.

- Wen, L.R., Guo, X.B., Liu, R.H., You, L.J., Abbasi, A.M., and Fu, X. (2015). Phenolic contents and cellular antioxidant activity of Chinese hawthorn "Crataegus pinnatifida". Food Chem. 186: 54–62. doi:10.1016/j. foodchem.2015.03.017.
- Wen, L.R., Guo, R.X., You, L.J., Abbasi, A.M., Li, T., Fu, X., and Liu, R.H. (2017). Major triterpenoids in Chinese hawthorn "Crataegus pinnatifida". and their effects on cell proliferation and apoptosis induction in MDA-MB-231 cancer cells. Food Chem. Toxicol. 100: 149–160.
- Wen, L., Lin, Y.L., Lv, R.M., Yan, H.J., Yu, J.Q., Zhao, H.Q., Wang, X., and Wang, D.J. (2017b). An efficient method for the preparative isolation and purification of flavonoids from leaves of *Crataegus pinnatifida* by HSCCC and pre-HPLC. Molecules. 22(5): 767. doi:10.3390/molecules22050767.
- Weon, J.B., Jung, Y.S., and Ma, C.J. (2016). Quality analysis of chlorogenic acid and hyperoside in Crataegi fructus. Pharmacogn. Mag. 12(46): 98–103. doi:10.4103/0973-1296.177904.
- Wu, J.Q., Peng, W., Qin, R.X., and Zhou, H. (2014). Crataegus pinnatifida: Chemical constituents, pharmacology, and potential applications. Molecules. 19(2): 1685–1712. doi:10.3390/molecules19021685.
- Wu, P.P., Li, F.J., Zhang, J.Y., Yang, B., Ji, Z.J., and Chen, W.D. (2017). Phytochemical compositions of extract from peel of hawthorn fruit, and its antioxidant capacity, cell growth inhibition, and acetylcholinesterase inhibitory activity. BMC Complement. Altern. Med. 17: 151. doi:10.1186/s12906-017-1662-y.
- Wyspiańska, D., Kucharska, A.Z., Sokół-Łętowska, A., and Kolniak-Ostek, J. (2017). Physico-chemical, antioxidant, and anti-inflammatory properties and stability of hawthorn (*Crataegus monogyna* Jacq.) procyanidins microcapsules with inulin and maltodextrin. J. Sci. Food Agric. 97(2): 669–678. doi:10.1002/jsfa.7787.
- Xia, N., Schramm, E., Koch, E., Burkart, M., Reifenberg, G., Forstermann, U., and Li, H. (2016). *Crataegus* extract WS (R) 1442 improves vascular function in diet-induced obese mice. Naunyn Schmiedebergs Arch. Pharmacol. 389(1): S38–S39.
- Xie, W.D., Zhao, Y.A., and Du, L.J. (2012). Emerging approaches of traditional Chinese medicine formulas for the treatment of hyperlipidemia. J. Ethnopharmacol. 140(2): 345–367. doi:10.1016/j.jep.2012.01.027.

- Yaglioglu, A.S., Eser, F., Tekin, S., and Onal, A. (2016). Antiproliferative activities of several plant extracts from Turkey on rat brain tumor and human cervix carcinoma cell lines. Front. Life Sci. 9(1): 69–74. doi:10 .1080/21553769.2015.1089949.
- Yang, B.R., and Liu, P.Z. (2012). Composition and health effects of phenolic compounds in hawthorn (*Crataegus* spp.) of different origins. J. Sci. Food Agric. 92(8): 1578–1590. doi:10.1002/jsfa.5671.
- Yonekubo, B.T., Alves, H.D.C., Marques, E.D., Perazzo, F.F., Rosa, P.C.P., Gaivao, I.O.D., and Maistro, E.L. (2018). The genotoxic effects of fruit extract of *Crataegus oxyacantha* (hawthorn) in mice. J. Toxicol. Environ. Health A 81(19): 974–982. doi:10.1080/15287394.2018.1503 982.
- Yoo, J.H., Liu, Y., and Kim, H.S. (2016). Hawthorn fruit extract elevates expression of Nrf2/HO-1 and improves lipid profiles in ovariectomized rats. Nutrients 8(5): 283. doi:10.3390/nu8050283.
- Zhang, F., Zhu, F.B., Li, J.J., Zhang, P.P., and Zhu, J.F. (2015). Hyperoside enhances the suppressive effects of arsenic trioxide on acute myeloid leukemia cells. Int. J. Clin. Exp. Med. 8(9): 15290–15295.
- Zhao, B.L. (2005). Natural antioxidants for neurodegenerative diseases. Mol. Neurobiol. 31(1-3): 283-293.:31 1-3: 283.
- Zheng, G.Q., Deng, J., Wen, L.R., You, L.J., Zhao, Z.G., and Zhou, L. (2018). Release of phenolic compounds and antioxidant capacity of Chinese hawthorn "Crataegus pinnatifida". during in vitro digestion. J. Funct. Foods. 40: 76–85.
- Zhong, L., Wang, Y.W., Peng, W., Liu, Y.J., Wan, J., Yang, S.L., Li, L., Wu, C.J., and Zhou, X. (2015). Headspace Solid-Phase Microextraction coupled with Gas Chromatography-Mass Spectrometric analysis of volatile components of raw and stir-fried fruit of C-Pinnatifida (FCP). Trop. J. Pharm. Res. 14(5): 891–898. doi:10.4314/tjpr.v14i5.20.
- Zhu, Q., Mao, L.N., Liu, C.P., Sun, Y.H., Jiang, B., Zhang, W., and Li, J.X. (2016). Antinociceptive effects of vitexin in a mouse model of postoperative pain. Sci. Rep. 6: 19266. doi:10.1038/srep19266.
- Zhu, Y., Feng, B., He, S.M., Su, Z.Q., and Zheng, G.J. (2018). Resveratrol combined with total flavones of hawthorn alleviate the endothelial cells injury after coronary bypass graft surgery. Phytomedicine 40: 20–26. doi:10.1016/j.phymed.2017.12.037.
- Zorniak, M., Szydlo, B., and Krzeminski, T.F. (2017). Crataegus special extract WS 1442: Up-to-date review of experimental and clinical experiences. J. Physiol. Pharmacol. 68(4): 521–526.