

Encapsulation strategies for dietary chlorogenic acid: advances in delivery systems and functional applications

Xin Guo^{a,#}, Qian-Lan Wu^{a,#}, Tian-Le Mao^a, Jing Li^a, Kiran Thakur^{a,b}, Ya-Fang Shang^{a,b},
Shao-Hua Yang^a, Yi-Long Ma^{a,b*} and Zhao-Jun Wei^{a,b*}

^aSchool of Food and Biological Engineering, Hefei University of Technology, Hefei 230009, China

^bSchool of Biological Science and Engineering, North Minzu University, Yinchuan 750021, China

[#]These authors contributed equally to this work.

*Corresponding author: Yi-Long Ma and Zhao-Jun Wei, School of Food and Biological Engineering, Hefei University of Technology, Hefei 230009, China., E-mail: yilong.ma@hfut.edu.cn (YLM) and zjwei@hfut.edu.cn (ZJW)

DOI: 10.26599/JFB.20xx.000xx

Received: August 19, 2025; Revised received & accepted: September 05, 2025

Citation: Guo, X., Wu, Q.-L., Mao, T.-L., Li, J., Thakur, K., Shang, Y.-F., Yang, S.-H., Ma, Y.-L., and Wei, Z.-J. (2025). Encapsulation strategies for dietary chlorogenic acid: advances in delivery systems and functional applications. J. Food Bioact. 000: 000–000.

Abstract

Chlorogenic acid (CGA) is a naturally occurring dietary polyphenol that has attracted significant attention due to its anti-oxidant, anti-inflammatory, anti-microbial, neuroprotective, and anti-cancer properties. These bioactivities highlight CGA's potential applications across food, nutraceutical, pharmaceutical, and cosmetic industries. However, its application remains currently limited by low stability, poor physiological stability, and insufficient bioavailability. To address these challenges, diverse encapsulation systems including nanoparticles, micelles, gels, liposomes, and metal-organic frameworks have been developed to protect CGA and from degradation and improve its bioactive efficacy. This review systematically summarizes the state of art encapsulation system for CGA, highlighting their design principles, release characteristics, and therapeutic applications. We aim to compare the strengths and limitations of each system, and provide a comprehensive reference to guide future research and promote the industrial translation of CGA-based formulations.

Keywords: Chlorogenic acid; Encapsulation; Nanocarriers; Bioavailability; Controlled release; Application.

1. Introduction

Chlorogenic acid (CGA) is one of the most abundant dietary phenolic acids, widely distributed in coffee, fruits, and vegetables (Gupta et al., 2022; Lu et al., 2020). Structurally characterized by its multiple hydroxyl groups, CGA exhibits strong antioxidant capacity through free radical scavenging, metal chelation, and inhibition of lipid peroxidation, which further contributes to its diverse biological activities, including anti-inflammatory, antimicrobial, neuroprotective, and anticancer effects (Kumar and Goel, 2019). These antioxidant properties are fundamental to CGA's therapeutic mechanisms, as oxidative stress underlies many pathological conditions. Owing to these properties, CGA has emerged as a promising candidate for therapeutic and functional food applications in

the pharmaceutical, nutraceutical, and cosmetic industries.

Despite its broad bioactivity, the application of CGA is severely restricted by its poor physicochemical stability and low bioavailability. CGA undergoes rapid degradation through oxidation, isomerization, and enzymatic hydrolysis during storage and under gastrointestinal (GI) conditions, resulting in diminished efficacy (Kiokias et al., 2020). Furthermore, its low aqueous solubility, limited intestinal absorption, and extensive metabolism significantly reduce systemic bioavailability. These limitations necessitate the development of advanced stabilization and delivery strategies to fully harness CGA's therapeutic potential.

Encapsulation technologies have recently emerged as a promising solution, providing structural protection, controlled release, and targeted delivery of bioactive molecules (Zhang et al., 2021b). Nanostructured carriers such as liposomes, polymeric nanoparti-

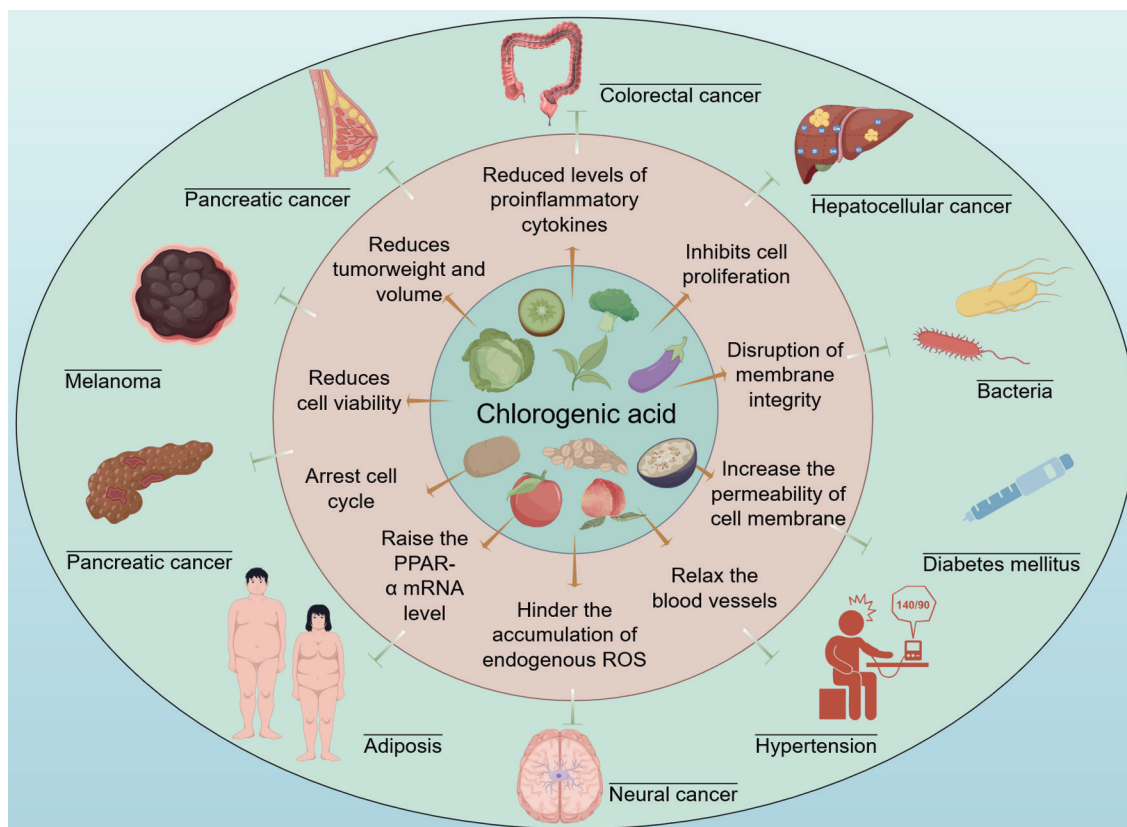


Figure 1. The bioactivity and potential application of CGA by Figdraw (ID: AUAYOc77fd).

cles, micelles, emulsions, and metal–organic frameworks can be engineered to enhance solubility, improve permeability, and extend circulation time (Ge et al., 2022; Bae and Park, 2020; Miao et al., 2025). By tuning the internal architecture and surface functionality, these systems can achieve controlled release kinetics and enhanced mucosal penetration, thereby overcoming physiological barriers such as the gastrointestinal tract (Laffleur and Bauer, 2021; Wang et al., 2022). These features collectively render encapsulation systems particularly promising candidates for improving the application of unstable bioactive compounds like CGA.

While numerous studies have reported the encapsulation strategies for CGA, a comprehensive comparative analysis of their mechanisms, efficacy, and translational potential is still lacking. This review therefore aims to summarize the current advancements in CGA encapsulation strategies, highlight their biological and technological advantages, and discuss future research directions to support the development of scalable, clinically relevant formulations.

2. Biological functions of CGA

CGA, one of the most abundant dietary polyphenols, has attracted substantial interest due to its broad spectrum of pharmacological activities, including anti-inflammatory, antimicrobial, and anti-cancer effects. Beyond its well-established antioxidant properties, CGA exerts additional pharmacological functions, including cardioprotective, antidiabetic, anti-obesity, and neuroprotective properties, highlighting its potential as a multifunctional therapeutic agent (Figure 1).

2.1. Anti-inflammatory activity

CGA modulates inflammatory responses primarily by suppressing the expression of pro-inflammatory cytokines. In Caco-2 cells stimulated with tumor necrosis factor- α (TNF- α) and hydrogen peroxide (H_2O_2), CGA treatment significantly reduced the expression of interleukin-8 (IL-8), a key inflammatory mediator (Shin et al., 2015). Feng et al. (2023) demonstrated that CGA treatment in a murine mastitis model downregulated serum IL-1 β , IL-6, cyclooxygenase-2 (COX-2), and soluble adhesion molecules (SSA). Notably, CGA showed superior suppression of IL-1 β and SSA compared with conventional antibiotics, underscoring its potential as a safer alternative in inflammatory disease management. Mechanistically, CGA's anti-inflammatory effects are linked to inhibition of NF- κ B and MAPK signaling pathways, which regulate cytokine expression and immune cell activation.

2.2. Antimicrobial activity

Beyond its anti-inflammatory properties, CGA exhibits broad-spectrum antimicrobial activity against both bacterial and fungal pathogens. Its mode of action involves disrupting the formation and structural integrity of microbial biofilms, thereby enhancing the susceptibility of pathogens to environmental stress. CGA has been shown inhibitory activity against a wide range of microorganisms, including *Candida albicans* (*C. albicans*) (Sung and Lee, 2010), *Staphylococcus aureus* (*S. aureus*) (Sun et al., 2021), *Yersinia enterocolitica* (*Y. enterocolitica*) (Chen et al., 2022), *Fusarium*

fujikuroi (*F. fujikuroi*) (Kai et al., 2021), and *Bacillus subtilis* (*B. subtilis*) (Wu et al., 2020b). Importantly, CGA showed minimal inhibitory effects on beneficial probiotic strains, supporting its potential in food preservation and biomedical formulations as well as anti-microbial coatings (Bondam et al., 2022).

2.3. Anticancer activity

Extensive studies support the antineoplastic potential of CGA across multiple cancer types, including colorectal, breast, hepatic, melanoma, and pancreatic cancers. In colorectal cancer models, CGA inhibited the proliferation and migration of HCT116, HCT15, and CT26 cell lines *in vitro*, and significantly reduced tumor volume and weight in corresponding xenografted mice (Li et al. 2021b). Similar effects have been observed in breast cancer model (via p53 and caspase-3 activation), hepatocellular carcinoma, melanoma, and pancreatic cancer. Changizi et al. (2021) demonstrated that CGA suppressed tumor progression and mitigated cancer-induced weight loss in mice bearing 4T1 tumors. Similar inhibitory effects have also been observed in hepatocellular carcinoma (Yan et al., 2017), melanoma (Li et al., 2021a), and pancreatic cancer models (Yang et al., 2021), further validating CGA's robust antitumor efficacy. Mechanistic studies suggest that CGA induces apoptosis through modulation of p53, Bax/Bcl-2, and caspase pathways, while also interfering with oncogenic signaling such as c-Myc–TFR1 axis in pancreatic carcinoma. These findings highlight CGA as a promising candidate for integrative cancer therapy.

2.4. Metabolic and neurological benefits

In addition to its well-documented anti-inflammatory, antibacterial, and anticancer properties, CGA demonstrates protective effects in metabolic and neurological disorders. It has been reported to lower blood pressure in hypertensive models by modulating endothelial nitric oxide production (Tom et al., 2016), reduce adiposity and improve lipid metabolism through upregulation of PPAR- α (Wan et al., 2013), regulate glucose metabolism in diabetes mellitus by enhancing insulin sensitivity and modulating hepatic glucose metabolism (Flanagan et al., 2014), and protect against neurodegenerative conditions (excitotoxicity and oxidative stress) by attenuating glutamate-induced apoptosis (Rebai et al., 2017). Collectively, these studies confirm CGA's role as a multifunctional bioactive compound, with therapeutic potential extending from chronic inflammation and infectious diseases to cancer and metabolic syndromes. However, its clinical application remains hindered by limited stability and systemic bioavailability, emphasizing the need for advanced delivery strategies.

3. CGA stability and bioavailability

Despite its diverse biological functions and considerable therapeutic potential, the clinical and industrial applications of CGA remain limited due to its poor physicochemical stability and low bioavailability (Liu et al., 2022; Abrankó and Clifford, 2017). These limitations are attributed to several factors: (1) its poor aqueous solubility due to strong intermolecular hydrogen bonding and hydrophobic interactions, which promote molecular aggregation in solution and reduce its dispersion, absorption, and utilization efficiency (Liu et al., 2022); (2) its low intestinal permeability due to

multiple hydroxyl groups, unsaturated double bonds, and rigid ring configurations, reduce its ability to cross epithelial membranes via passive diffusion (Lv et al., 2025); (3) its chemical instability under physiological and storage conditions. Its ester bond is prone to enzymatic hydrolysis during digestion, generating caffeic and quinic acids. Additionally, CGA can undergo oxidation, or isomerization, and polymerization, leading to substantial degradation even after absorption (Kumar and Goel, 2019; Abrankó and Clifford, 2017); (4) its extensive metabolism through hydrolysis, conjugation (e.g., sulfation, glucuronidation, methylation), oxidation, and transformation by intestinal microbiota, further decrease its systemic availability (Clifford et al., 2020). Even when absorbed, CGA demonstrates short half-life and rapid renal clearance, which limit its therapeutic persistence and systemic exposure. Collectively, these barriers significantly compromise the therapeutic effectiveness of CGA. Therefore, the development of effective stabilization strategies, encapsulation systems such as nanoparticles, micelles, hydrogels, liposomes, and metal–organic frameworks offer promising solutions by (i) improving solubility, (ii) shielding CGA from chemical and enzymatic degradation, (iii) enabling controlled release, and (iv) enhancing absorption across biological barriers (Laffleur and Keckeis, 2020).

4. CGA encapsulation strategies and applications

Encapsulation systems are nanostructured platforms composed of organic or inorganic hybrid materials designed to improve stability, solubility, and bioavailability of bioactive compounds. These systems not only protect sensitive molecules such as CGA from premature degradation, but also enhance their absorption and therapeutic efficacy. Furthermore, encapsulation systems can be engineered to achieve sustained release and site-specific delivery of active ingredients by incorporating specific functional components, including ligands that target cell surface receptors or other relevant proteins. In recent years, multiple nanotechnology-based systems including nanoparticles, micelles, hydrogels, liposomes, and metal–organic frameworks have been engineered for CGA delivery (Figure 2).

4.1. Nanoparticles

Nanoparticles are versatile nanostructured carriers composed of organic, inorganic, or hybrid materials, typically formed through techniques such as self-assembly or layer-by-layer deposition. These systems are capable of efficiently encapsulating bioactive molecules, thereby significantly enhancing the stability, solubility, and bioavailability of these compounds (Ma and Moulton, 2011; Awad et al., 2023). To date, a variety of nanoparticles have been developed for CGA encapsulation, utilizing diverse materials such as proteins, polysaccharides, graphene, selenium, and other nano-materials (Table 1).

Encapsulation of CGA significantly enhances its stability, release profile, and bioavailability compared to free CGA. For example, in polyvinyl alcohol (PVA)/ γ -polyglutamic acid (γ PGA) electrospun nanofiber mats, free CGA released only ~36% after 72 h, whereas encapsulated CGA in mats containing 5% and 10% γ -PGA released ~65% and ~82%, respectively, demonstrating both reduced burst release and higher sustained release (Sandoval-Herrera et al., 2021). Similarly, pea protein nanoparticles (PPCNPs) achieved ~61% encapsulation efficiency and increased *in vitro* ABTS (2,2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid))

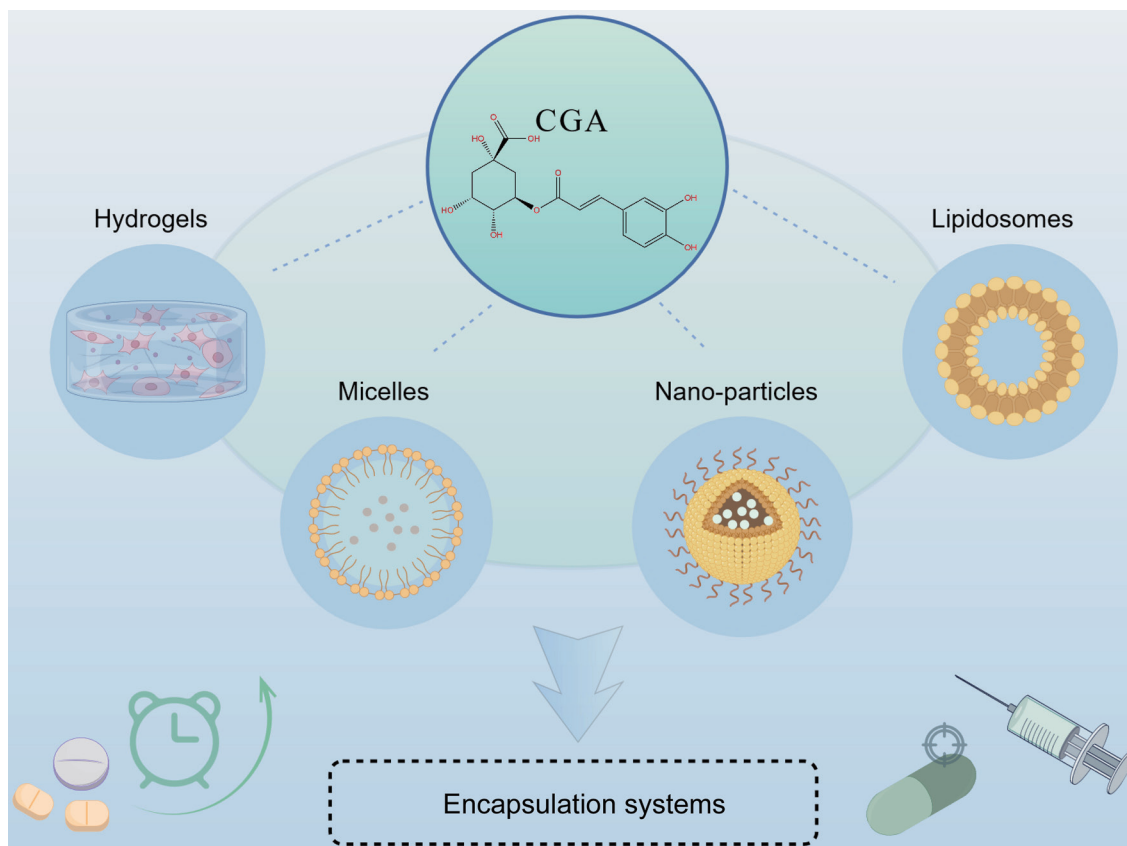


Figure 2. Encapsulation strategies of CGA by Figdraw (ID: AYRWY85a6f).

radical scavenging capacity, resulting in approximately 7.75% higher bioaccessibility versus free CGA. Additionally, [Chen et al. \(2017\)](#) demonstrated that oral administration of CGA-self-micro-emulsifying drug delivery system (CGA-SMEDDS) led to a 2.5-fold increase in bioavailability (249.4%) relative to conventional CGA suspensions, further supporting the bioavailability enhancement effect of nanocarrier systems. [Jiang et al. \(2024\)](#) reported that cucurbit[7]uril (Q[7]) @2CGA nanocomposites achieved a more stable release profile, with approximately 70% CGA released steadily over 5 days, offering enhanced structural protection compared to free CGA. These quantitative data support the claim the nanocarrier-based encapsulation yields superior protection, release control, and systemic exposure of CGA relative to its unencapsulated form.

In addition to conventional nanocarrier systems, co-assembly and macromolecular encapsulation strategies have also demonstrated significant potential in enhancing CGA functionality. [Fu et al. \(2024\)](#) developed supramolecular nanoparticles through the self-assembly of berberine (BBR) and CGA, which exhibited strong antibacterial activity against *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* (MRSA). The co-assembled nanoparticles effectively disrupted bacterial membranes, downregulated resistance genes, and enhanced CGA utilization, leading to improved anti-inflammatory efficacy and accelerated wound healing in a murine skin infection model.

Similarly, [Zhang et al. \(2023\)](#) utilized the reversible assembly properties of apoferritin (Apo) to load CGA, either alone or with sodium alginate (SA). The resulting Apo/SA system improved CGA's stability under heat and UV exposure and enabled con-

trolled release in both gastric and intestinal environments, thereby enhancing oral bioavailability. In addition, [Ma et al. \(2024\)](#) formulated chitosan-based nanoparticles incorporating rhamnolipids (RL) and CGA via ionic crosslinking. The composite films (F/CRC) displayed excellent antioxidant activity, ultraviolet A (UVA) shielding, and biocompatibility, while also allowing for sustained CGA release, making them suitable for active packaging applications. These findings collectively demonstrate that both small molecule co-assembly and macromolecular carriers (e.g., proteins, polysaccharides) can significantly improve CGA's stability, bioavailability, and therapeutic efficacy in diverse applications.

Inorganic nanomaterials have also shown great promise as delivery platforms for CGA due to their structural stability and functional versatility. [Barahuie et al. \(2017\)](#) developed a graphene oxide-based nanocomposite (CAGO) via hydrogen bonding and π - π stacking, which significantly enhanced CGA stability. Compared to free CGA, the CAGO system retained over 70% of its content after 72 h, while free CGA degraded by more than 50%, offering a more sustained release profile and reducing therapeutic fluctuations. While the > 103 h *in vitro* release suggests depot potential, clinical translation requires pharmacokinetic validation of absorption, metabolism, and clearance profiles.

Metal-based carriers have been explored for both protection and targeted delivery. [Li et al. \(2023\)](#) constructed selenium nanoclusters modified with a brain-targeting peptide (TGN-CGA@SeNCs), which improved CGA stability under alkaline gastrointestinal conditions and increased oral bioavailability by 2.3-fold *in vivo*. The system also enhanced neuroprotective effects and positively modulated gut microbiota composition. Likewise, [Roy et](#)

Table 1. Nanoparticles for CGA encapsulation and application

Carrier System	Preparation Method	Surface Modification	Size	Effects	Advantages	Ref.
PVA/ γ -PGA Nanofiber Mats	Electrospinning	/	119 nm	Reduced burst & extended release	82% vs 36% CGA release after 72 h (10% γ -PGA)	(Sandoval-Herrera et al., 2021)
PPCNPs	Nanoprecipitation	/	211.62–429.79 nm	~7.75% higher <i>in vitro</i> bioaccessibility	Improved digestion stability and antioxidant capacity	(Liang et al., 2024)
CGA-SMEDDS	Self-microemulsification	/	16.37 nm	2.5 \times oral bioavailability	Oral bioavailability increased to 249.4% vs suspension	(Chen et al., 2017)
Q[7]@2CGA NPs	Host–guest inclusion	/	100nm	Improved release stability	~70% CGA released steadily over 5 days	(Jiang et al., 2024)
BBR/CGA NPs	Self-assembly	/	360.6 nm	Improved bioavailability	<i>S. aureus</i> inhibition 87.94% vs 59.05% (free); MRSA: 88.10% vs 46.28%	(Fu et al., 2024)
SA-Apo-CGA NPs	Co-encapsulation	/	156.6 nm	Improved bioavailability and GI release control	CGA UV retention: 96.50% vs 77.79%; slower release in GI environment	(Zhang et al., 2023)
CS/RL/CGA NPs	Ionic crosslinking	/	190 nm	CGA fixation in RL and CS	Lower CGA migration and slower release vs F/CGA indicate good fixation	(Ma et al., 2024)
CAGO	Intermolecular interactions	/	/	Prolonged CGA release	CGA release duration >103 h; beyond CGA's pharmacokinetic half-life	(Barahuie et al., 2017)
TGN-CGA@SeNCs	Selenium nanoparticle base	TGN peptide surface modification	58.06 nm	Improved GI stability	CGA degradation under pH 8: 47.83% vs 74.22% for free CGA	(Li et al., 2023)
AgNPs-CGA-BSA	Silver nanoparticle synthesis	BSA surface modification	96 nm	Antibacterial, antioxidant, and anticancer effects	DPPH IC50: 91.72 μ g/mL vs 94.03 μ g/mL (free CGA); enhanced bioactivity	(Roy et al., 2022)
CGA@MIL-125-NH ₂	MOF synthesis + adsorption	/	226–653 nm	Improved aqueous stability	At pH 8, CGA@MOF retained >85% after 7 days vs <10% for free CGA; sustained release	(Rincón et al., 2024)

al. (2022) synthesized CGA-loaded silver nanoparticles (AgNPs-CGA), further stabilized with bovine serum albumin (BSA). These hybrid nanoparticles showed enhanced cellular uptake and cytotoxicity against cancer cell lines, demonstrating potent antioxidant and antitumor effects compared to free CGA.

Metal–organic frameworks (MOFs) are crystalline, porous materials formed through the self-assembly of metal ions or clusters with organic ligands. These structures exhibit high surface areas, tunable pore sizes, and excellent stability, making them ideal candidates for controlled drug delivery (Li et al., 2017). Rincón et al. (2024) encapsulated CGA into MIL-125-NH₂, a titanium-based MOF, using an impregnation method. While free CGA rapidly degraded in pH 8 buffer, the encapsulated form retained over 85% of its content after seven days, with sustained release, indicating superior long-term stability.

Collectively, these inorganic nanocarriers including carbon-based, metal-based, and MOF systems are next generation platform which offer enhanced physicochemical protection, bioavailability, and controlled release of CGA, supporting their application in advanced therapeutic delivery. However, challenges in toxicity, large-scale production, and regulatory acceptance must be addressed before widespread adoption.

4.2. Micelles

Micelles are nanoscale colloidal systems with a distinct core–shell architecture. They typically consist of a hydrophobic core enclosed by a hydrophilic outer shell, which enables them to encapsulate lipophilic active compounds and protect them from degradation in aqueous environments (Li et al., 2022b). This unique structural configuration enhances the solubility, stability, and bioavailability of encapsulated agents (Bui et al., 2012; Jo et al., 2020). In recent years, micelles have become a widely explored platform in drug delivery systems, particularly for combination therapies aimed at achieving synergistic pharmacological effects while minimizing toxicity. Micelle-based delivery systems (Table 2), particularly those formed from biodegradable amphiphilic copolymers, offer a promising strategy to overcome these challenges by stabilizing CGA, improving its solubility, and enabling targeted, sustained release. Poly(lactic-co-glycolic acid) (PLGA)-based polymeric micelles have become the benchmark carriers for CGA, combining FDA-approved biocompatibility, predictable hydrolytic degradation, and versatile surface chemistries that enable active-targeting ligand conjugation (Ge et al., 2022).

Vyawahare et al. (2024) developed PLGA-based micelles with a

Table 2. Micelles for CGA encapsulation and application

Carrier System	Composition/Structure	Size	Release Behavior	Enhancement on CGA	<i>In Vivo</i> /Therapeutic Effects	Ref.
PLGA-b-CGA/ PLGA-b- CGA-MTX	Electrospinning	90–110 nm	Stable at pH 7.4; fast dual release at pH 5.8 with esterase	Prevents premature CGA leakage; pH/enzyme-triggered release; improves inflammation targeting	Suppressed joint swelling, delayed cartilage erosion in arthritis model	(Vyawahare et al., 2024)
CGA-PLGA@PVP	CGA covalently linked to PLGA; surface PVP-modified	154.4 ± 19.3 nm	Sustained CGA release: ~56% at 72 h; ~65% at 132 h	Enhanced solubility, colloidal stability, and prolonged CGA retention in tissue	Reduced gingival inflammation and bone loss in periodontitis model	(Li et al., 2022a)
CGA-loaded PVA/PLGA NPs (F2)	Electrosprayed PVA/PLGA loaded with CGA	454.36 ± 36.74 nm	Initial burst: 29.46%; Cumulative: 97.42%	Enhanced antiviral potency (↓IC ₅₀), prolonged release, higher tissue accumulation	Effective against HCoV-229E, MERS-CoV, and <i>P. aeruginosa</i> in lung infection model	(Saleh et al., 2023)

particle size ranging from 90 to 110 nm, which were designed to co-deliver CGA and methotrexate (MTX). These micelles maintained structural stability in neutral physiological conditions, effectively preventing premature CGA leakage or degradation. Under acidic and enzyme-rich inflammatory environments, however, the micelles underwent triggered destabilization, resulting in synchronized and accelerated release of CGA. PLGA micelles markedly amplified CGA's anti-arthritis efficacy, curbing joint swelling, cartilage loss, and inflammation, while simultaneously prolonging its targeted action. Based on these findings, Li et al. (2022a) developed a refined micellar system-CGA-PLGA@PVP-where CGA was covalently bound to PLGA and further modified with polyvinylpyrrolidone (PVP) to improve hydrophilicity and colloidal stability. The resulting micelles (~154.4 ± 19.3 nm, zeta potential: −19.6 mV) demonstrated excellent aqueous dispersion and controlled CGA release, reaching 56% in 72 h and ~65% by 132 h. This formulation ensured CGA's prolonged retention in target tissues after local administration. In a mouse model of periodontitis, the micelles achieved localized, long-acting delivery of CGA, which led to significant reductions in inflammation and alveolar bone resorption, underscoring their role in preserving CGA's integrity and enabling therapeutic retention.

In addition to anti-inflammatory applications, micelle and nanoparticle-based delivery systems also significantly enhance CGA's antimicrobial and antiviral activities. In a recent study, CGA was encapsulated within PVA/PLGA nanoparticles using electrospraying, achieving an optimal formulation (F2) with high loading efficiency (80.23%), sustained release (97.42% cumulative release), and a particle size of ~454 nm (Saleh et al., 2023). Notably, the nanoparticle-loaded CGA exhibited significantly lower IC₅₀ values against human coronavirus 229E (HCoV-229E) and Middle East respiratory syndrome coronavirus (MERS-CoV) (170 µg/mL and 223 µg/mL, respectively) compared to free CGA ($p < 0.05$). Moreover, in a murine model of *Pseudomonas aeruginosa* lung infection, the CGA nanoparticles showed superior antibacterial effects. These findings demonstrate that nanocarriers not only protect CGA from premature degradation but also amplify its pharmacological action across multiple infection models, further expanding its therapeutic potential beyond inflammatory diseases.

4.3. Hydrogels

Hydrogels are cross-linked polymeric systems characterized by

a three-dimensional network structure composed of hydrophilic polymer chains and a substantial proportion of water molecules. These systems typically exist in a solid or semi-solid state at the macroscopic level and exhibit excellent water retention capacity. Due to their highly hydrated nature, biocompatibility, and tunable physicochemical properties, hydrogels have gained considerable attention as advanced wound treatment matrices (Toyoshima et al., 2022). Hydrogel-based delivery systems have shown considerable promise in addressing the key limitations of CGA, including its chemical instability, low oral bioavailability, and insufficient accumulation at target sites. A subset of advanced hydrogel formulations demonstrates clear advantages over free CGA, as evidenced by improved stability, controlled release, and enhanced bioactivity across various administration routes (Table 3).

Wang et al. (2024) developed a dual pH- and reactive oxygen species (ROS)-responsive hydrogel incorporating CGA encapsulated within zeolitic imidazolate framework-8 (ZIF-8). This system significantly enhanced the thermal stability of CGA, protected it from degradation, and enabled on-demand release under oxidative diabetic wound microenvironments. These features facilitated macrophage polarization and angiogenesis, resulting in accelerated wound repair.

In the context of oral delivery, Harwansh et al. (2025) formulated enteric-coated chitosan–carrageenan hydrogel beads loaded with CGA for the treatment of ulcerative colitis. The beads showed ~95% sustained release of CGA at colonic pH over 24 h, with high encapsulation efficiency (~84%) and targeted delivery to inflamed intestinal sites. This formulation significantly improved CGA's gastrointestinal stability and mucosal retention, which are critical for its therapeutic effect in the colon.

For topical applications, Trivedi and Puranik (2023) designed a nanophytovesicle-loaded hydrogel containing CGA, which achieved a 2.2-fold increase in skin permeation compared to free CGA. The hydrogel also prolonged CGA's antioxidant activity and accelerated wound closure *in vivo*, confirming its role in enhancing transdermal bioavailability and therapeutic effectiveness.

Huang et al. (2023) developed a novel CGA-based hydrogel dressing that self-assembles into a nearly uniform linear microstructure. *In vitro* drug release experiments showed a cumulative release of CGA reaching 69% within 24 h, demonstrating sustained release behavior. Further *in vitro* assessments confirmed the hydrogel's pronounced anti-inflammatory activity. Its therapeutic effects in wound healing were attributed to enhanced wound closure,

Table 3. Hydrogels for CGA encapsulation and application

Hydrogel System	Key Mechanism	Enhancement vs. Free CGA	Target Application	Ref.
Dual pH/ROS-responsive hydrogel with ZIF-8@CGA	ZIF-8 stabilization; oxidative and acidic stimulus-triggered release	↑ Thermal stability, ↑ on-demand release, ↑ wound healing via M1 polarization	Diabetic wound healing	(Wang et al., 2024)
Enteric-coated chitosan–carrageenan CGA hydrogel beads	Colon-specific pH-triggered release; mucoadhesion	↑ GI stability, ↑ mucosal retention, ~95% sustained release at colonic pH	Ulcerative colitis	(Harwansh et al., 2025)
Nanophytovesicle-loaded hydrogel	Nanoencapsulation improves skin permeation and antioxidant retention	↑ Skin permeation (2.2×), ↑ antioxidant persistence, ↑ <i>in vivo</i> wound healing	Topical wound therapy	(Trivedi and Puranik, 2023)
Self-assembling CGA hydrogel dressing	Self-assembly enables structural integrity and controlled release	↑ Anti-inflammatory effect, ↑ collagen deposition, 69% sustained release at 24h	Skin wound healing	(Huang et al., 2023)

increased collagen deposition, and re-epithelialization, accompanied by suppression of pro-inflammatory cytokine expression and upregulation of growth factor expression. CGA-loaded hydrogels excel in oral, topical, and wound-care uses, offering biocompatibility, on-demand release, and sustained antioxidant/antimicrobial action; yet their industrial adoption hinges on stronger mechanics and scalable, reproducible manufacturing.

4.4. Liposomes

Liposomes are spherical vesicles primarily composed of phospholipids and fatty acids, closely mimicking the structure of biological membranes. This bilayer structure allows liposomes to encapsulate both hydrophilic and hydrophobic drugs, offering enhanced protection and stability for therapeutic agents. Compared with other delivery systems such as polymeric nanoparticles, micelles, and hydrogels, liposomes exhibit a more intricate architecture that enables superior drug loading and retention. Furthermore, the ease of surface functionalization and biocompatibility of liposomes have contributed to their widespread application in drug delivery through various administration routes (Waghule et al., 2022). In particular, liposomes have shown great promise in delivering low molecular weight active pharmaceutical ingredients (APIs) (Wu et al., 2020a).

CGA repolarizes M2 tumor-associated macrophages (TAMs) to the antitumor M1 phenotype, positioning it as a potent immunomodulator in cancer therapy. This modulation of the tumor microenvironment contributes to the inhibition of glioblastoma (GBM) growth. However, the clinical application of CGA remains limited owing to its rapid systemic clearance and insufficient accumulation at tumor sites (Xue et al., 2017). To address these challenges, Ye et al. (2020) developed mannoseylated PE-Gylated liposomes (Man-PEG-Lipo) for the encapsulation and delivery of CGA. These liposomes effectively enhance CGA delivery to TAMs, reduce rapid clearance, and strengthen the anti-tumor immune response induced by CGA, while demonstrating minimal toxicity. As a result, the bioavailability and therapeutic potential of CGA in anti-tumor applications were significantly improved (Table 4).

In clinical cancer immunotherapy, the inherent instability of CGA *in vivo* necessitates daily intramuscular injections, which presents considerable challenges for patient compliance. To overcome this limitation, Zhang et al. (2021a) designed a PE-Gylated liposomal system (CPPL) based on a CGA-phospholipid

complex as the intermediate. In animal studies, CPPL administration resulted in plasma CGA concentrations exceeding 200 ng/mL at 24 h, markedly higher than those achieved by CGA solution. Furthermore, CPPL improved the stability of CGA in systemic circulation and sustained its antitumor efficacy, even when the dosing interval was extended to four days. This formulation offers a more convenient and effective strategy for the clinical use of CGA.

In the treatment of aggressive skin tumors such as melanoma, Zhu et al. (2024) proposed an innovative liposomal formulation to enhance therapeutic efficacy. They developed a liposome co-loaded with CGA and doxorubicin (DOX) and functionalized it with a sialic acid–octadecylamine conjugate (SA-ODA), referred to as CGA-DOX-SAL. *In vitro* release studies demonstrated that CGA-DOX-SAL liposomes provided sustained CGA release, achieving a cumulative release of 60.56% over 24 h, which was substantially lower than the rapid 95.97% release observed for free CGA within four hours. *In vivo* studies confirmed that CGA-DOX-SAL liposomes effectively accumulated in tumor tissues and enhanced anti-tumor activity. These findings highlight the potential of this liposomal system for melanoma therapy and suggest its applicability in the treatment of other tumor types.

In addition to their established role in cancer therapy and in enhancing drug potency and targeting, liposomes have shown significant promise in improving the oral absorption and bioavailability of CGA. Feng et al. (2016) reported that chlorogenic acid-loaded liposomes (CAL), formulated with cholesterol and phosphatidylcholine, markedly increased both the oral bioavailability and antioxidant activity of CGA. Compared with free CGA, CAL extended the circulation duration of CGA and achieved a 1.29-fold relative increase in bioavailability.

Based on these findings, Alemán et al. (2022) further explored the protective effects of liposomes on CGA stability within the gastrointestinal tract. They encapsulated an aqueous extract of sea fennel (*Crithmum maritimum*), which is rich in CGA, into soy phosphatidylcholine liposomes. *In vitro* gastrointestinal digestion simulations demonstrated that CGA encapsulated in liposomes remained stable throughout the digestion process, whereas the CGA content in the free extract decreased significantly by 40%. The liposomal encapsulation effectively protected CGA from degradation and morphological alteration, thereby substantially enhancing its stability during gastrointestinal digestion.

The application of liposome technology in CGA delivery not only broadens the scope of CGA in cancer immunotherapy but also provides innovative strategies for the development of potent and

Table 4. Liposomes for CGA encapsulation and application

Formulation Name	Composition & Features	Target/Model	CGA Enhancement	Therapeutic Outcome	Ref.
Man-PEG-Lipo	PEGylated liposomes functionalized with mannose for TAM targeting	Tumor-associated macrophages (TAMs), glioblastoma	Increased TAM targeting; reduced clearance; immune activation	Inhibited GBM growth via M2→M1 polarization	(Ye et al., 2020)
CPPL	PEGylated liposomes based on CGA–phospholipid complex	Systemic tumor model	Sustained plasma CGA levels (>200 ng/mL at 24h); 4-day dosing interval; improved stability	Reduced injection frequency; maintained anti-tumor efficacy	(Zhang et al., 2021a)
CGA-DOX-SAL	CGA + Doxorubicin co-loaded liposomes with SA-ODA surface modification	Melanoma	Sustained CGA release (60.56% at 24h); controlled dual drug release	Enhanced tumor accumulation and anti-melanoma effect	(Zhu et al., 2024)
CAL	CGA-loaded liposomes using cholesterol and phosphatidylcholine	Oral delivery (pharmacokinetics)	Extended circulation; 1.29× increase in oral bioavailability	Improved antioxidant activity and systemic exposure	(Feng et al., 2016)
CGA–SPL (Soy-liposomes)	Liposomes loaded with sea fennel extract (CGA-rich) using soy phosphatidylcholine	Simulated GI digestion	Protected CGA from pH/enzymatic degradation; retained structural integrity during digestion	Enhanced GI stability → potential for nutraceutical or oral formulations	(Alemán et al., 2022)

safe anti-tumor therapies by improving its stability and bioavailability during oral administration.

5. Conclusion and future perspectives

CGA is an exceptionally versatile polyphenol, yet its susceptibility to oxidation, hydrolysis, and photodegradation severely limits its translational potential. Encapsulation technologies ranging from lipid-based nanocarriers and polymeric micelles to metal–organic frameworks and supramolecular hydrogels have emerged as powerful tools to enhance CGA’s physicochemical stability, control its release kinetics, and broaden its formulation space. Comparative analyses indicate that no single platform is universally optimal; instead, carrier selection must be tailored to the intended route of administration, target tissue, and desired release profile. Our review emphasizes the key gaps: (i) poorly characterized CGA–carrier interactions with limited understanding of binding mechanisms and their effects on release performance; (ii) low drug loading efficiency (<15% w/w of CGA relative to total carrier weight) combined with rapid burst release (>40% of encapsulated CGA released within 2–4 hours in PBS at pH 7.4, 37 °C, 100 rpm); (iii) unproven stability under industrial processing and storage conditions that could compromise CGA integrity and delivery performance. Future work should integrate multi-scale modelling (atomistic to population balance) with continuous-flow microfluidics for scalable, size-controlled carriers; extend CGA from nutraceuticals to other production processes.

Acknowledgments

This study was supported by the National Key R&D Program of China (2022YFF1100300), Fundamental Research Funds for the

Central Universities (JZ2024HGTG0287), National Natural Science Foundation of China (32272312), Natural Science Foundation of Ningxia province (2023AAC05043).

Conflict of interest

The authors affirm that they have no conflicts of interest.

References

- Abbrankó, L., and Clifford, M.N. (2017). An Unambiguous Nomenclature for the Acyl-quinic Acids Commonly Known as Chlorogenic Acids. *J. Agric. Food Chem.* 65: 3602–3608.
- Alemán, A., Marín-Peñalver, D., de Palencia, P.F., Gómez-Guillén, M. Del C., and Montero, P. (2022). Anti-Inflammatory Properties, Bioaccessibility and Intestinal Absorption of Sea Fennel (*Crithmum maritimum*) Extract Encapsulated in Soy Phosphatidylcholine Liposomes. *Nutrients* 14: 210.
- Awad, R., Avital, A., and Sosnik, A. (2023). Polymeric nanocarriers for nose-to-brain drug delivery in neurodegenerative diseases and neurodevelopmental disorders. *Acta Pharm. Sin. B* 13: 1866–1886.
- Bae, Y.H., and Park, K. (2020). Advanced drug delivery 2020 and beyond: Perspectives on the future. *Adv. Drug Delivery Rev.* 158: 4–16.
- Barahue, F., Saifullah, B., Dorniani, D., Fakurazi, S., Karthivashan, G., Hussein, M.Z., and Elfghi, F.M. (2017). Graphene oxide as a nanocarrier for controlled release and targeted delivery of an anticancer active agent, chlorogenic acid. *Mater. Sci. Eng. C* 74: 177–185.
- Bondam, A.F., Diolinda da Silveira, D., Pozzada dos Santos, J., and Hoffmann, J.F. (2022). Phenolic compounds from coffee by-products: Extraction and application in the food and pharmaceutical industries. *Trends Food Sci. Technol.* 123: 172–186.
- Bui, L., Abbou, S., Ibarboure, E., Guidolin, N., Staedel, C., Toulmé, J.-J., Le-commandoux, S., and Schatz, C. (2012). Encapsulation of RNA–Polyelectrolyte Complexes with Amphiphilic Block Copolymers: Toward a New Self-Assembly Route. *J. Am. Chem. Soc.* 134: 20189–20196.

- Changizi, Z., Moslehi, A., Rohani, A.H., and Eidi, A. (2021). Chlorogenic acid induces 4T1 breast cancer tumor's apoptosis via p53, Bax, Bcl-2, and caspase-3 signaling pathways in BALB/c mice. *J. Biochem. Mol. Toxicol.* 35: e22642.
- Chen, K., Peng, C., Chi, F., Yu, C., Yang, Q., and Li, Z. (2022). Antibacterial and Antibiofilm Activities of Chlorogenic Acid Against *Yersinia enterocolitica*. *Front. Microbiol.* 13: 885092.
- Chen, L., Liu, C., Chen, Q., Wang, S., Xiong, Y., Jing, J., and Lv, J. (2017). Characterization, pharmacokinetics and tissue distribution of chlorogenic acid-loaded self-microemulsifying drug delivery system. *Eur. J. Pharm. Sci.* 100: 102–108.
- Clifford, M.N., Kerimi, A., and Williamson, G. (2020). Bioavailability and metabolism of chlorogenic acids (acyl-quinic acids) in humans. *Compr. Rev. Food. Sci. Food Saf.* 19: 1299–1352.
- Feng, S., Zhang, Y., Fu, S., Li, Z., Zhang, J., Xu, Y., Han, X., and Miao, J. (2023). Application of Chlorogenic acid as a substitute for antibiotics in Multidrug-resistant *Escherichia coli*-induced mastitis. *Int. Immunopharmacol.* 114: 109536.
- Feng, Y., Sun, C., Yuan, Y., Zhu, Y., Wan, J., Firempong, C.K., Omari-Siaw, E., Xu, Y., Pu, Z., Yu, J., and Xu, X. (2016). Enhanced oral bioavailability and *in vivo* antioxidant activity of chlorogenic acid *via* liposomal formulation. *Int. J. Pharm.* 501: 342–349.
- Flanagan, J., Bily, A., Rolland, Y., and Roller, M. (2014). Lipolytic Activity of Svetol®, a Decaffeinated Green Coffee Bean Extract. *Phytother. Res.* 28: 946–948.
- Fu, S., Yi, X., Li, Y., Li, Y., Qu, X., Miao, P., and Xu, Y. (2024). Berberine and chlorogenic acid-assembled nanoparticles for highly efficient inhibition of multidrug-resistant *Staphylococcus aureus*. *J. Hazard. Mater.* 473: 134680.
- Ge, K., Paus, R., Penner, V., Sadowski, G., and Ji, Y. (2022). Theoretical modeling and prediction of biorelevant solubility of poorly soluble pharmaceuticals. *Chem. Eng. J.* 444: 136678.
- Gupta, A., Atanasov, A.G., Li, Y., Kumar, N., and Bishayee, A. (2022). Chlorogenic acid for cancer prevention and therapy: Current status on efficacy and mechanisms of action. *Pharmacol. Res.* 186: 106505.
- Harwansh, R.K., Bhati, H., Deshmukh, R., and Rahman, M.A. (2025). Preparation, Characterization, and In Vitro Evaluation of Chlorogenic Acid Loaded Hydrogel Beads for the Management of Ulcerative Colitis. *Assay Drug Dev. Technol.* 23: 29–43.
- Huang, H., Chen, L., Hou, Y., He, W., Wang, X., Zhang, D., and Hu, J. (2023). Self-assembly of chlorogenic acid into hydrogel for accelerating wound healing. *Colloids Surf., B* 228: 113440.
- Jiang, Y., Qi, H., Wang, M., Chen, K., Chen, C., and Xie, H. (2024). Chlorogenic Acid-Cucurbit[n]uril Nanocomplex Delivery System: Synthesis and Evaluations for Potential Applications in Osteoporosis Medication. *Int. J. Nanomed.* 19: 11577–11592.
- Jo, M.J., Jin, I.S., Park, C.-W., Hwang, B.Y., Chung, Y.B., Kim, J.-S., and Shin, D.H. (2020). Revolutionizing technologies of nanomicelles for combinatorial anticancer drug delivery. *Arch. Pharm. Res.* 43: 100–109.
- Kai, K., Wang, R., Bi, W., Ma, Z., Shi, W., Ye, Y., and Zhang, D. (2021). Chlorogenic acid induces ROS-dependent apoptosis in *Fusarium fujikuroi* and decreases the postharvest rot of cherry tomato. *World J. Microbiol. Biotechnol.* 37: 93.
- Kiokias, S., Proestos, C., and Oreopoulou, V. (2020). Phenolic Acids of Plant Origin—A Review on Their Antioxidant Activity In Vitro (O/W Emulsion Systems) Along with Their *In Vivo* Health Biochemical Properties. *Foods* 9: 534.
- Kumar, N., and Goel, N. (2019). Phenolic acids: Natural versatile molecules with promising therapeutic applications. *Biotechnol. Rep.* 24: e00370.
- Laffleur, F., and Bauer, B. (2021). Progress in nasal drug delivery systems. *Int. J. Pharm.* 607: 120994.
- Laffleur, F., and Keckeis, V. (2020). Advances in drug delivery systems: Work in progress still needed? *Int. J. Pharm.* 590: 119912.
- Li, H., Xu, J., Hu, J.-F., Hu, Q.-Y., Fang, X., Sun, Z.-J., Xu, Z., and Zhang, L. (2022a). Sustained release of chlorogenic acid-loaded nanomicelles alleviates bone loss in mouse periodontitis. *Biomater. Sci.* 10: 5583–5595.
- Li, L., Zeng, Y., Chen, M., and Liu, G. (2022b). Application of Nanomicelles in Enhancing Bioavailability and Biological Efficacy of Bioactive Nutrients. *Polymers* 14: 3278.
- Li, X., Zhu, S., Yin, P., Zhang, S., Xu, J., Zhang, Q., Shi, S., and Zhang, T. (2021a). Combination immunotherapy of chlorogenic acid liposomes modified with sialic acid and PD-1 blockers effectively enhances the anti-tumor immune response and therapeutic effects. *Drug Deliv.* 28: 1849–1860.
- Li, Y., Pu, R., Zhou, L., Wang, D., and Li, X. (2021b). Effects of a Chlorogenic Acid-Containing Herbal Medicine (LASNB) on Colon Cancer. *Evid. Based Complement. Alternat. Med.* 2021: 9923467.
- Li, Y., Zou, B., Xiao, A., and Zhang, H. (2017). Advances of Metal-Organic Frameworks in Energy and Environmental Applications. *Chin. J. Chem.* 35: 1501–1511.
- Li, Z., Zheng, G., Wang, N., Liang, H., Li, C., Wang, Y., Cui, Y., and Yang, L. (2023). A Flower-like Brain Targeted Selenium Nanocluster Lowers the Chlorogenic Acid Dose for Ameliorating Cognitive Impairment in APP/PS1 Mice. *J. Agric. Food Chem.* 71: 2883–2897.
- Liang, L., Cao, W., Li, L., Liu, W., Wei, X., Chen, J., Ren, G., Zhao, Y., and Duan, X. (2024). Fabrication and characterisation of pea protein isolate-chlorogenic acid nanoparticles. *Int. J. Food Sci. Technol.* 59: 1615–1623.
- Liu, W., Li, W., Zhang, P., Gong, X., Tu, P., Tang, L., Li, J., and Song, Y. (2022). Quality structural annotation for the metabolites of chlorogenic acid in rat. *Food Chem.* 379: 132134.
- Lu, H., Tian, Z., Cui, Y., Liu, Z., and Ma, X. (2020). Chlorogenic acid: A comprehensive review of the dietary sources, processing effects, bioavailability, beneficial properties, mechanisms of action, and future directions. *Compr. Rev. Food Sci. Food Saf.* 19: 3130–3158.
- Lv, G., Li, D., Bao, Y., Qin, Y., Zhang, X., Xu, F., Zhu, K., Zhang, Y., and Zhang, Y. (2025). Effects of methoxy groups and carbon-carbon double bonds in phenolic acids on the physicochemical characteristics and digestibility of cassava starch. *Carbohydr. Polym.* 362: 123687.
- Ma, K., Li, F., Zhe, T., Sun, X., Zhang, X., Wan, P., Na, H., Zhao, J., and Wang, L. (2024). Biopolymer films incorporated with chlorogenic acid nanoparticles for active food packaging application. *Food Chem.* 435: 137552.
- Ma, Z., and Moulton, B. (2011). Recent advances of discrete coordination complexes and coordination polymers in drug delivery. *Coord. Chem. Rev.* 255: 1623–1641.
- Miao, Y.-H., Wang, X., Zhao, X.-M., Hu, Y.-W., Liu, X., and Deng, D.-W. (2025). Co-assembly strategies of natural plant compounds for improving their bioavailability. *Food Med. Homol.* 2: 9420022.
- Rebai, O., Belkhir, M., Sanchez-Gomez, M.V., Matute, C., Fattouch, S., and Amri, M. (2017). Differential Molecular Targets for Neuroprotective Effect of Chlorogenic Acid and its Related Compounds Against Glutamate Induced Excitotoxicity and Oxidative Stress in Rat Cortical Neurons. *Neurochem. Res.* 42: 3559–3572.
- Rincón, I., Contreras, M.c., Sierra-Serrano, B., Salles, F., Rodríguez-Diéguez, A., Rojas, S., and Horcajada, P. (2024). Long-lasting insecticidal activity in plants driven by chlorogenic acid-loaded metal-organic frameworks. *J. Mater. Chem. B* 12: 4717–4723.
- Roy, T., Dey, S.K., Pradhan, A., Chaudhuri, A.D., Dolai, M., Mandal, S.M., and Choudhury, S.M. (2022). Facile and Green Fabrication of Highly Competent Surface-Modified Chlorogenic Acid Silver Nanoparticles: Characterization and Antioxidant and Cancer Chemopreventive Potential. *ACS Omega* 7: 48018–48033.
- Saleh, A., Abdelkader, D.H., El-Masry, T.A., Eliwa, D., Alotaibi, B., Negm, W.A., and Elekhawey, E. (2023). Antiviral and antibacterial potential of electrosprayed PVA/PLGA nanoparticles loaded with chlorogenic acid for the management of coronavirus and *Pseudomonas aeruginosa* lung infection. *Artif. Cells Nanomed. Biotechnol.* 51: 255–267.
- Sandoval-Herrera, I., Romero-García, J., Ledezma-Pérez, A., Alvarado-Canché, C., Torres-Lubian, R., and De-León, A. (2021). Controlled Release of Chlorogenic Acid from Polyvinyl Alcohol/Poly (γ -Glutamic Acid) Blended Electrospun Nanofiber Mats with Potential Applications in Diabetic Foot Treatment. *Polymers* 13: 2943.
- Shin, H.S., Satsu, H., Bae, M.-J., Zhao, Z., Ogiwara, H., Totsuka, M., and Shimizu, M. (2015). Anti-inflammatory effect of chlorogenic acid on the IL-8 production in Caco-2 cells and the dextran sulphate sodium-induced colitis symptoms in C57BL/6 mice. *Food Chem.* 168: 167–175.
- Sun, J., Wang, D., Sun, Z., Liu, F., Du, L., and Wang, D. (2021). The combination of ultrasound and chlorogenic acid to inactivate *Staphylococcus*

- aureus* under planktonic, biofilm, and food systems. *Ultrason. Sonochem.* 80: 105801.
- Sung, W.S., and Lee, D.G. (2010). Antifungal action of chlorogenic acid against pathogenic fungi, mediated by membrane disruption. *Pure Appl. Chem.* 82: 219–226.
- Tom, E.N.L., Girard-Thernier, C., and Demougeot, C. (2016). The Janus face of chlorogenic acid on vascular reactivity: A study on rat isolated vessels. *Phytomedicine* 23: 1037–1042.
- Toyoshima, Y., Kawamura, A., Takashima, Y., and Miyata, T. (2022). Design of molecularly imprinted hydrogels with thermoresponsive drug binding sites. *J. Mater. Chem. B* 10: 6644–6654.
- Trivedi, H.R., and Puranik, P.K. (2023). Chlorogenic acid-optimized nanophytosomes: a novel approach for enhanced permeability and oral bioavailability. *Future J. Pharm. Sci.* 9: 116.
- Vyawahare, A., Jori, C., Kumar, J., Kanika, Fareed, M., Ali, N., Parida, K., and Khan, R. (2024). A chlorogenic acid-conjugated nanomicelle attenuates disease severity in experimental arthritis. *Biomater. Sci.* 18: 3241–3472.
- Waghule, T., Saha, R.N., Alexander, A., and Singhvi, G. (2022). Tailoring the multi-functional properties of phospholipids for simple to complex self-assemblies. *J. Control. Release* 349: 460–474.
- Wan, C.-W., Wong, C.N.-Y., Pin, W.-K., Wong, M.H.-Y., Kwok, C.-Y., Chan, R.Y.-K., Yu, P.H.-F., and Chan, S.-W. (2013). Chlorogenic Acid Exhibits Cholesterol Lowering and Fatty Liver Attenuating Properties by Up-regulating the Gene Expression of PPAR- α in Hypercholesterolemic Rats Induced with a High-Cholesterol Diet. *Phytother. Res.* 27: 545–551.
- Wang, L., Shi, Y., Jiang, J., Li, C., Zhang, H., Zhang, X., Jiang, T., Wang, L., Wang, Y., and Feng, L. (2022). Micro-Nanocarriers Based Drug Delivery Technology for Blood-Brain Barrier Crossing and Brain Tumor Targeting Therapy. *Small* 18: 2203678.
- Wang, S., Liu, Y., Wang, X., Chen, L., Huang, W., Xiong, T., Wang, N., Guo, J., Gao, Z., and Jin, M. (2024). Modulating macrophage phenotype for accelerated wound healing with chlorogenic acid-loaded nanocomposite hydrogel. *J. Control. Release.* 369: 420–443.
- Wu, H., Luo, Y., Xu, D., Ke, X., and Ci, T. (2020a). Low molecular weight heparin modified bone targeting liposomes for orthotopic osteosarcoma and breast cancer bone metastatic tumors. *Int. J. Biol. Macromol.* 164: 2583–2597.
- Wu, Y., Liang, S., Zhang, M., Wang, Z., Wang, Z., and Ren, X. (2020b). The Effect of Chlorogenic Acid on *Bacillus subtilis* Based on Metabolomics. *Molecules* 25: 4038.
- Xue, N., Zhou, Q., Ji, M., Jin, J., Lai, F., Chen, J., Zhang, M., Jia, J., Yang, H., Zhang, J., Li, W., Jiang, J., and Chen, X. (2017). Chlorogenic acid inhibits glioblastoma growth through repolarizing macrophage from M2 to M1 phenotype. *Sci. Rep.* 7: 39011.
- Yan, Y., Liu, N., Hou, N., Dong, L., and Li, J. (2017). Chlorogenic acid inhibits hepatocellular carcinoma *in vitro* and *in vivo*. *J. Nutr. Biochem.* 46: 68–73.
- Yang, H., Said, A.M., Huang, H., Papa, A.P.D., Jin, G., Wu, S., Ma, N., Lan, L., Shangguan, F., and Zhang, Q. (2021). Chlorogenic acid depresses cellular bioenergetics to suppress pancreatic carcinoma through modulating c-Myc-TFR1 axis. *Phytother. Res.* 35: 2200–2210.
- Ye, J., Yang, Y., Jin, J., Ji, M., Gao, Y., Feng, Y., Wang, H., Chen, X., and Liu, Y. (2020). Targeted delivery of chlorogenic acid by mannosylated liposomes to effectively promote the polarization of TAMs for the treatment of glioblastoma. *Bioact. Mater.* 5: 694–708.
- Zhang, S., Deng, X., Guo, X., and Zhang, J. (2023). Sustained release of chlorogenic acid by co-encapsulation of sodium alginate binding to the Northern pike (*Esox Lucius*) liver ferritin. *Food Chem.* 429: 136924.
- Zhang, Y., Yang, Y., Ye, J., Gao, Y., Liao, H., Zhou, J., Feng, Y., Liu, D., Meng, Y., Chen, X., Gao, L., and Liu, Y. (2021a). Construction of chlorogenic acid-containing liposomes with prolonged antitumor immunity based on T cell regulation. *Sci. China Life Sci.* 64: 1097–1115.
- Zhang, Z., Qiu, C., Li, X., McClements, D.J., Jiao, A., Wang, J., and Jin, Z. (2021b). Advances in research on interactions between polyphenols and biology-based nano-delivery systems and their applications in improving the bioavailability of polyphenols. *Trends Food Sci. Technol.* 116: 492–500.
- Zhu, S., Li, X., Luo, Z., Ding, M., Shi, S., and Zhang, T. (2024). Combined immunochemotherapy achieving targeted co-delivery of chlorogenic acid and doxorubicin by sialic acid-modified liposomes enhances anti-cancer efficacy. *Drug Deliv. Transl. Res.* 14: 718–729.