



Selection of polyphenol oxidase affects biotransformation efficacy of targeted theaflavins

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DOI: 10.31665/JFB.2022.17297

Received: March 28, 2022; Revised received & accepted: March 30, 2022

Citation: Wang, W., Ho, C.-T., and Li, S. (2022). Selection of polyphenol oxidase affects biotransformation efficacy of targeted theaflavins. J. Food Bioact. 17: 2–5.

Abstract

Theaflavins in black tea and other fermented tea have attracted many studies because of their stronger anti-oxidant and anti-inflammatory effects among bioactives other than catechins. However, within the four major theaflavins, namely theaflavin, theaflavin-3-*O*-gallate, theaflavin-3'-*O*-gallate and theaflavin-3,3'-*O,O*-digallate, their biological properties are different. A method to efficiently and selectively synthesize targeted theaflavins with desired property is a key condition for further evaluation. Herein, we have summarized the sources of polyphenol oxidase (PPO) and the yields of total and individual theaflavins based on some available publications. This overview lays the foundation for a comprehensive review in this area of research in the near future.

Keywords: Theaflavins; Polyphenol oxidase; Catechins; Theaflavin monogallate; Theaflavin digallate; Anti-inflammation.

1. Introduction

Tea is the most consumed functional beverage in the world, among which black tea is the most popular tea and accounts for 78% of the overall tea industry. The major polyphenols in black tea consists of theaflavins and catechins. The chemical components of black tea have high amount of green tea catechins (4–15%) and theaflavins are usually between 0.5 and 2% by dry weight. Major theaflavins are theaflavin (TF1), theaflavin-3-*O*-gallate (TF2a), theaflavin-3'-*O*-gallate (TF2b) and theaflavin-3,3'-*O,O*-digallate (TF3). Although to a lesser degree as compared to green tea, black tea also contain major catechins, including (–)-epicatechin (EC), (–)-epicatechin gallate (ECG), (–)-epigallocatechin (EGC) and (–)-epigallocatechin gallate (EGCG) (Li et al., 2013; 2021). Catechins and theaflavins are reported to be the chief contributors to the biological activity of black tea by additive or synergistic effects.

The health-promotional properties of black tea and its major theaflavin components include antioxidant (de Majia, Ramirez-Mares and Puangpraphant, 2009; Xu, et al., 2021), anti-inflamma-

tory (Arent et al., 2010; Gosslau et al., 2011), anti-cancer (Yang et al., 2009; Pan et al., 2013), cardioprotective (Stangl et al., 2007; Santesso and Manheimer, 2014) effects as well as controlling of obesity and metabolic syndrome, among others (Lin et al., 2007; Tang et al., 2013; Vermeer et al., 2008).

2. Formation of theaflavins

Despite the fact that manufacturing process of black tea is traditionally termed as fermentation of green tea, it is actually a first order oxidation reaction under the catalysis of polyphenol oxidase and peroxidase followed by a polymerization of the oxidized quinones. This is a completely different process from microbial fermentation that produces ethanol and vinegar from crops and soy sauce from soybean. As mentioned above, theaflavin formation undergoes oxidation and polymerization, two simplified steps (Li et al., 2013). The first step is the oxidation reaction (Figure 1), i.e. green tea catechins, particularly EGC and EGCG are partially oxidized to quinones under the catalysis of polyphenol oxidase

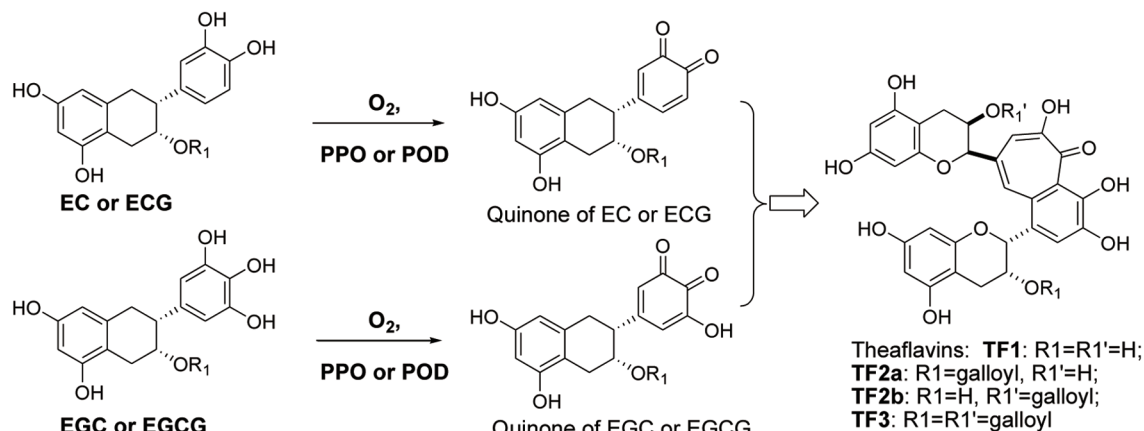


Figure 1. Formation of black tea theaflavins.

(PPO, EC1.14.18.1) and/or peroxidase (POD, EC1.11.1.7). PPO and POD exist naturally in fresh tea leaves, fruits and vegetables.

3. Examples of PPO sources from plants and microorganisms

The initial enzyme-catalyzed oxidation is a key step and, therefore the existence of PPO/POD in plants or microorganisms crucial for this process. Many studies have found that the sources of PPO and POD play very important roles in multiple aspects, such as yields, conversion rate and composition/ratio of individual theaflavins such as TF1, TF2a, TF2b and TF3. Traditionally, the source from fresh green tea leaves is used in the ‘fermentation’ process to produce black tea. However, there are limitations to rely solely on tea PPO/POD, such as the off season time of tea harvesting and low conversion yields from green tea catechins to black tea theaflavins. However, it is reported that PPO has greater efficacy than POD in catalyzing the oxidation of catechins to their corresponding quinones (Jiang et al., 2018). Therefore, other sources of PPO have been explored from vegetables, fruits, mushrooms and microbes (Lin et al., 2017; Sang et al., 2011). Examples of PPO derived from different sources are listed in Table 1. As seen in Table 1, the optimal conditions to obtain maximum yields of theaflavins, including temperature, pH, time and concentration, are different from one source of enzyme to another. For instance, between the two different tea leaves, the synthesized content of theaflavins catalyzed by the PPO of *C. sinensis* is 38.8% higher than that of *C. sinensis* var. *assamica* (Huang et al., 2017; Lin et al., 2017). Pear, *Malus pumila* and *Ipomoea batatas*, especially pear (*Echeveria ‘Sulli’*) affords the highest catalytic capacity (Wang et al., 2007).

4. Yields and ratios of major theaflavins influenced by different sources of PPOs

Theaflavins in black tea or related products are important for their biological activities as demonstrated by a proprietary 40% theaflavins-containing black tea extract to induce a strong inhibition of pro-inflammatory markers. The markers considered were COX-2, TNF- α , ICAM-1, IL-1 β , IL-6, IL-8, NF- κ B, C-JUN and p53 and up-regulation of anti-inflammatory IL-10 in a cDNA and oligomicroarray analysis by the use of a human cell-based monocyte-macrophage differentiation model (Arent et al., 2010; Li et

al., 2019). Therefore, sufficient content of theaflavins is a key for black tea or its products to demonstrate efficacy against inflammation, among others. Furthermore, it is also evidenced that TF2 and TF3 are more effective in their bioactivities than TF1 (Gossiau et al. 2011; Zhou et al., 2022). Hence, the content of total theaflavins and proportions of TF2 and/or TF3 are of great importance in order to reach the desired functional bioactivity. The exploration has generally focused on the conversion yield of total theaflavins starting with green tea or catechins. Table 2 shows several examples of the impact on the conversion efficiency to theaflavins with different sources of PPO. It is evident that PPOs from pear genus (*Pyrus* family) has high catalytic efficiency in producing total theaflavins regardless of enzyme activity. However, more than 65% of the theaflavins was TF1, which does not show any outstanding biofunctionality, indicating that enzymes from pear are not good catalysts for theaflavin products with high percentage of TF2 and TF3 (Wang et al., 2007). The combined contents of TF2 and TF3 are over 50% from tea PPOs (*Camellia* family), representing natural ‘fermentation’ of green tea, but overall transformation yield of theaflavins is much lower than that of pear PPOs (Wang et al., 2007; Li, 2006; Lin et al., 2017). Thus, more exploration and optimization are needed to obtain high yield of total theaflavins from tea PPOs. It is worth to pay special attention that two sources of PPO can catalytically transfer catechins to very high percentages of TF2 and TF3. In particular, 54% of TF2a, 28% of TF2b were obtained with the PPO from *Malus pumila* Mill (Lin et al., 2017) and nearly 60% of TF2 and 37% of TF3, a total of 97% of TF2 and TF3 combined, were generated with the catalysis of PPO from *Dioscoreae Rhizoma* (Lin et al. 2017). Therefore, we have learned from the limited reports that there are options to selectively produce high percentages of TF1, TF2a, TF2b or TF3 with a careful selection of PPOs from different sources, but further optimization of conversion yield and reaction conditions are warranted to reach the desired goals.

5. Conclusion

Theaflavins as a whole have a multitude of biological activities, but different theaflavins, i.e. TF1, TF2 and TF3, have demonstrated different potency in various biological assays. TF2 and TF3 have possess better efficacy than TF1. In preparation of theaflavin-rich black tea or its related products, aside from the ratios of natural catechins, PPO plays a key role in the synthesis of theaflavins.

Table 1. Examples of different sources of PPO used in theaflavin synthesis with mixed catechins as substrates

Plants	Plant parts	Temp-erature (°C)	pH	Time (min)	Catechin concen-tration (mg/mL)	Content of theaflavins/Conversion rate	Reference
Tea (<i>Camellia sinensis</i>)	Leaves	37	4.3	49	5.59	0.754 mg/mL	Lin et al., 2017
Tea (<i>Camellia sinensis</i> var. <i>assamica</i> cv)	Leaves	37	4.0	–	4.5	0.461 mg/mL 15.31 %	Huang et al., 2017
Pear (<i>Pyrus sorotina</i>)	fruit	20	5.5	40	–	45.7%	Lin et al., 2017
Pear (<i>Echeveria 'Sullii'</i>)	fruit	30	5.5	40	10	673.57 mg/g	Wang et al., 2007
Pear (<i>Pyrus</i> spp)	fruit	30	4.4	15	1	0.3253g/L	Xu, 2013
<i>Solanum melongena</i>	Fruit	30	4.8	15	1	0.2872g/L	Xu, 2013
<i>Solanum melongena</i>	Fruit	25	4.5	40	2.5	7.45 mg	Fang et al., 2011
<i>Solanum tuberosum</i>	Fresh potato	30	–	90	5	22.86%	Li et al., 2021
<i>Solanum tuberosum</i>	Fresh potato	25	4.8	45	1	0.068g/L	Xu, 2013
<i>Malus pumila</i>	Leaves	30	4.8	40	18	86.8%	Lin et al., 2017
<i>Musa nana Lour.</i>	Fruit	30	6.0	30	1	0.09g/L	Xu, 2013
<i>Dioscoreae rhizoma</i>	Fresh root	35	5.5	40	22	28.1%	Lin et al., 2017
<i>Ipomoea batatas</i>	Fresh root	25	6.0	10	5	86.9%	Lin et al., 2017
<i>Trametes trogii</i>	Fungi extract	36	4.5	720	5	0.2%	Lin et al., 2017
<i>Trametes trogii</i> (CGMCC5.629)	Fungi extract	28	–	60	10	10.193%	Wang et al., 2007
Mushroom	Fresh mushroom	30	5.5	40	10	44.96 mg/g	Wang et al., 2007

Note: Substrate of catechins was mixtures extracted from green tea.

Table 2. Examples for contents of four major theaflavins produced by different sources of PPO with catechins as substrates

PPO Source	PPO purity	Activ-ity (U/ mL/min)	TF1	TF2a	TF2b	TF3	Total TFs	Unit	Reference
<i>Pyrus sorotina</i>	crude	655.50	451.31 (67.0%)	132.06 (19.6%)	82.89 (12.3%)	7.32 (1.1%)	673.57 (100%)	mg/g	Wang et al., 2007
<i>Pyrus pyrifolia</i>	crude	195.53	396.32 (73.0%)	94.64 (17.4%)	40.85 (7.5%)	10.78 (2.0%)	542.59 (100%)	mg/g	Wang et al., 2007
<i>Pyrus nivalis</i>	crude	83.88	334.70 (66.6%)	120.75 (24.0%)	35.25 (7.0%)	11.93 (2.4%)	502.63 (100%)	mg/g	Wang et al., 2007
<i>Pyrus sinkiangensis</i>	crude	57.28	257.52 (65.3%)	98.74 (25.1%)	26.4 (6.7%)	11.51 (2.9%)	394.16 (100%)	mg/g	Wang et al., 2007
<i>Pyrus pyrifolia 'Shineiki'</i>	crude	373.83	232.81 (70.6%)	44.03 (13.4%)	45.18 (13.7%)	7.55 (2.3%)	329.56 (100%)	mg/g	Wang et al., 2007
<i>Camellia sinensis</i>	crude	128.97	94.50 (49.3%)	50.30 (26.3%)	38.60 (20.1%)	8.20 (4.3%)	191.60 (100%)	mg/g	Wang et al., 2007
<i>Camellia sinensis</i>	Slat + column	1306	9.25% (46.2%)	2.55% (12.7%)	3.71% (18.5%)	4.50% (22.5%)	20.01% (100%)	%	Li, 2006
<i>Camellia sinensis</i> 'Longjing'	crude	–	10.27% (22.5%)	17.16% (37.5%)	11.68% (25.5%)	6.63 % (14.5%)	45.74% (100%)	%	Lin et al., 2017
<i>Malus pumila Mill</i>	crude	–	1.71% (8.9%)	10.32 % (54.0%)	5.36 % (28.0%)	1.73% (9.0%)	19.12% (100%)	%	Lin et al., 2017
<i>Dioscoreae Rhizoma</i>	Slat + column	–	0.85% (3.3%)	12.4% (48.6%)	2.78% (10.9%)	9.46% (37.1%)	25.49% (100%)	%	Lin et al., 2017

From the knowledge gained from only limited publications available, we recommend a careful source selection of PPOs as a priority in producing theaflavin-rich products with targeted individual theaflavins prior to further optimization for mass production of tea products.

Acknowledgment

This research is funded by the grant from Hubei Province, China (2019ABA100).

References

- Arent, S.M., Senso, M., Golem, D.L., and McKeever, K.H. (2010). The effects of theaflavin-enriched black tea extract on muscle soreness, oxidative stress, inflammation, and endocrine responses to acute anaerobic interval training: a randomized, double-blind, crossover study. *J. Int. Soc. Sports Nutr.* 7(1): 11.
- de Majia, E.G., Ramirez-Mares, M.V., and Puangpraphant, S. (2009). Bioactive components of tea: cancer, inflammation and behavior. *Brain Behav. Immun.* 23: 721–731.
- Fang, W.P., Wang, L.P., Yu, J., Yue, P.X., Jiang, X., Feng, W.Y., Chenzhou, Y.Q., and Li, X.H. (2011). Studies on optimum conditions of synthesizing theaflavins by using bio-enzyme method. *Appl. Mech. Mater* 138: 929–932.
- Gosslau, A., Li, S., Ho, C.-T., Chen, K.-Y., and Rawson, N.E. (2011). The importance of natural product characterization in studies of their anti-inflammatory activity. *Mol. Nutr. Food Res.* 55: 74–82.
- Huang, Y., Wu, M., Yao, Y., and Huiang, Y. (2017). Effects of different conditions on theaflavins synthesis by polyphenol oxidase of *Camellia sinensis* var. *assamica* cv. Mengku. *Food Sci.* 38(22): 54–59.
- Jiang, Y., Hua, J., Yuan, H., and Ma, H. (2018). Effect of Different Tea Cultivars on Theaflavin Formation during Suspended Fermentation. *Food Sci.* 39(20): 71–77.
- Li, J., Li, D., Tong, K., Lei, Y., Jiang, B., Tang, X., and Li, Y. (2021). Enzymatic properties of purified polyphenol oxidase from potato and its ability to enzymatic synthesis of theaflavins. *Food Fermentation Ind.* 47(11): 26–31.
- Li, S. (2006). Study on Theaflavins Synthesis by Screening and Using Microorganism Polyphenol Oxidase. Dissertation, Hunan Agricultural University, Changsha, China. (in Chinese with English abstract).
- Li, S., Gosslau, A., Lange, K.W., and Ho, C.-T. (2019). Profiled tea extracts exemplifying the importance of characterizing food bioactives: opinion piece. *J. Food Bioac.* 5: 1–5.
- Li, S., Lo, C.-Y., Pan, M.-H., Lai, C.-S., and Ho, C.-T. (2013). Black tea: chemical analysis and stability. *Food Funct.* 4: 10–18.
- Lin, C.L., Huang, H.C., and Lin, J.K. (2007). Theaflavins attenuate hepatic lipid accumulation through activating AMPK in human HepG2 cells. *J. Lipid Res.* 48: 2334–2343.
- Lin, C.-X., Yang, J.-R., Wang, G.-Y., Ni, H., and Li, H.-H. (2017). Catalytic synthesis of theaflavins with polyphenol oxidase. *Plant Physiol. J.* 53(8): 1359–1364.
- Pan, M.-H., Lai, C.-S., Wang, H., Lo, C.-Y., Ho, C.-T., and Li, S. (2013). Black tea in chemo-prevention of cancer and other human diseases. *Food Sci. Hum. Well.* 2: 12–21.
- Sang, S., Lambert, J.D., Ho, C.-T., and Yang, C.-S. (2011). The chemistry and biotransformation of tea constituents. *Pharm. Res.* 64(2): 87–99.
- Santesso, N., and Manheimer, E. (2014). A summary of a cochrane review: Green and black tea for the primary prevention of cardiovascular disease. *Glob. Adv. Health Med.* 3(2): 66–67.
- Stangl, V., Dreger, H., Stangl, K., and Lorenz, M. (2007). Molecular targets of tea polyphenols in the cardiovascular system. *Cardiovasc. Res.* 73: 348–358.
- Tang, W., Li, S., Liu, Y., Huang, M.-T., and Ho, C.-T. (2013). Anti-diabetic activity of chemically profiled green tea and black tea extracts in a type 2 diabetes mice model via different mechanisms. *J. Funct. Foods.* 5: 1784–1793.
- Vermeer, M.A., Mulder, T.P., and Molhuizen, H.O. (2008). Theaflavins from black tea, especially theaflavin-3-gallate, reduce the incorporation of cholesterol into mixed micelles. *J. Agric. Food Chem.* 56: 12031–12036.
- Wang, K.-B., Liu, Z.-H., Zhao, S.-J., Huang, J.-A., Fu, D.-H., and Liu, F. (2007). Effect of PPO isoenzyme on formation of theaflavin during in vitro oxidation. *Res. Agric. Modernization.* 28(5): 618–621.
- Xu, H. (2013). Study on the pear polyphenol oxidase biocatalytic synthesis of theaflavins and the application in the tea extraction. Dissertation, Zhejiang University of Technology, Hangzhou, China. (in Chinese with English abstract).
- Xu, X.-X., Zheng, G., Tang, S.-K., Liu, H.-X., Hu, Y.-Z., and Shang, P. (2021). Theaflavin protects chondrocytes against apoptosis and senescence via regulating Nrf2 and ameliorates murine osteoarthritis. *Food Funct.* 12: 1590–1602.
- Yang, C.-S., Lambert, J.D., and Sang, S. (2009). Antioxidative and anti-carcinogenic activities of tea polyphenols. *Arch Toxicol.* 83: 11–21.
- Zhan, J., Cao, H., Hu, T., Shen, J., Wang, W., Wu, P., Yang, G., Ho, C.-T., and Li, S. (2021). Efficient preparation of black tea extract (BTE) with high content of theaflavin mono- and di-gallates and the protective effects of BTE on CCl₄-induced rat liver and renal injury. *J. Agric. Food Chem.* 69: 5938–5947.
- Zhou, J., Liu, C., Zhao, S., Liu, Y., Zhang, S., Zhao, Q., Wang, F., Xu, G., Huang, J., and Liu, Z. (2022). Improved yield of theaflavin-3,3'-digallate from *Bacillus megaterium* tyrosinase via directed evolution. *Food Chem.* 375: 131848.