

Review J. Food Bioact. 2021;13:40–51

Phytochemicals and biological activities of Pueraria flower: a review

Juan Wang^{a#}, Fan Yang^{a#}, Yongqing Tao^a, Meiyan Wang^a, Zhibo Han^{b,c} and Hui Zhao^{a*}

^aTianjin Key Laboratory of Food and Biotechnology, State Experimental and Training Centre of Food and Drug, School of Biotechnology and Food Science, Tianjin University of Commerce, No. 409 Guangrong Road, Beichen, Tianjin 300134, China

^bState Key Lab of Experimental Hematology, Institute of Hematology and Blood Diseases Hospital, Chinese Academy of Medical Sciences, Tianjin 300020, China

^cJiangxi Engineering Research Center for Stem Cell, Shangrao, Jiangxi, China

*These two authors contributed equally to this work.

*Corresponding author: Hui Zhao, Tianjin Key Laboratory of Food and Biotechnology, State Experimental and Training Centre of Food and Drug, School of Biotechnology and Food Science, Tianjin University of Commerce, No. 409 Guangrong Road, Beichen, Tianjin 300134, China. E-mail: zhaohui@tjcu.edu.cn

DOI: 10.31665/JFB.2021.13258

Received: December 11, 2020; Revised received & accepted: March 28, 2021

Citation: Wang, J., Yang, F., Tao, Y., Wang, M., Han, Z., and Zhao, H. (2021). Phytochemicals and biological activities of Pueraria flower: a review. J. Food Bioact. 13: 40–51.

Abstract

Pueraria lobata (Kudzu root) has been well documented as a food and also an herbal plant for its ability to alleviate hangover, diarrhea and cardiovascular diseases. However, the flower of Pueraria lobata has been attracted attention only in recent decades. Bioactive phytochemicals such as isoflavones, saponins, essential oils and other components have been isolated and identified in Pueraria flower extracts (PFE). Both *in vivo* and *in vitro* research have indicated the health promoting effects of Pueraria flower including hepatoprotective property, estrogenic effects, antioxidant activity, anti-inflammation activity and other pharmacological activities. In this review, we have summarized the chemical compositions and pharmacological actions of Pueraria flower and updated knowledge with recent progress.

Keywords: Pueraria flower; Isoflavone; Saponin; Phytochemicals; Biological activities.

1. Introduction

Dietary plants are important sources of nutraceuticals and support 70–80% of the population as a primary and non-conventional medicine worldwide (Chan, 2003). Growing evidence from epidemiological and case-control studies indicate that the reduced risk of chronic diseases is tightly associated with the intake of phytochemicals originating from dietary plants (Li et al., 2020; Wen et al., 2020; Wu et al., 2019). Indeed, dietary interventions including raw plant materials and nutraceuticals have been identified to prevent various diseases such as obesity, diabetes, cardiovascular diseases, Alzheimer's disease, and cancers (Li et al., 2019; Sheng et al., 2019; Wang et al., 2014; Wang et al., 2020; Wirngo et al., 2016; Zhang et al., 2018).

Pueraria lobata belongs to *Leguminosae* family, and it is one of the earliest medicinal plants used in traditional Chinese medicine.

The components and pharmacological activities of the root of Pueraria lobata have been extensively studied (Keung and Vallee, 1998; Wong et al., 2011; Zhang et al., 2017; Zhou et al., 2014). There are more than 70 phytochemicals identified in the root of Pueraria lobate (Kudzu root). Among these compounds, isoflavonoids and triterpenoids are the major constituents. Thus, compounds-oriented tactics lead to Kudzu root as an effective medicinal intervention for diabetes, cardiovascular diseases and imbalance in endocrine systems (Wong et al., 2011).

As the flower-based herb from Pueraria lobata, Pueraria flower (Puerariae Flos) has attracted increasing attention due to its bioactivities in hypoglycemia, hypolipidemia, and weight loss. Assessment of phytochemicals indicates that isoflavonoid and essential oils are the chief components in Pueraria flower (Lertpatipanpong et al., 2020; Wang et al., 2013; Yu et al., 2011). On the basis of its promising development potential, we summarized the up to date

Table 1. Major flavonoids in Pueraria flower

| No. | compounds | R ₁ | R ₂ | R ₃ | Reference |
|-----|-----------------------------|-----------------|----------------|----------------|---|
| 1 | Rutin | glc-rha | ОН | ОН | Ding et al., 2013; Lertpatipanpong et al., 2020 |
| 2 | Luteolin | Н | ОН | ОН | Ding et al., 2013 |
| 3 | Apigenin | Н | ОН | Н | Lu et al., 2013Lertpatipanpong et al., 2020 |
| 4 | Nicotiflorin | O-glc-(6→1)-xyl | ОН | Н | Lu et al., 2013 |
| 5 | Apigenin 4'-O-β-D-glucoside | O-glc | Н | Н | Lu et al., 2013 |

knowledge regarding phytochemicals and pharmacological activities of Pueraria flower. In particular, key issues involving the relationship between active ingredients and molecular mechanisms are highlighted in this review.

2. Methods

The current review considered the literature published prior to September 2020 on phytochemistry, pharmacology and toxicity of extracts isolated from Pueraria flower. All the available information on Pueraria flower was collected via electronic search such as PubMed, Google Scholar, and Web of Science. The literature was searched from the databases using the keywords "Pueraria flower" with no exact time limit (all fields) as well as various books that were accessed for information that were directly related to the present contribution. Information of all related books, full-text articles and conference notes written in English and Chinese were also very reliable.

3. Phytochemicals

3.1. Flavonoids

Chemical structures of flavonoids from Pueraria flower are summarized in Table 1. The flowers of Pueraria flower were extracted with MeOH. Identification by high performance liquid chromatography (HPLC) and other methods showed that there were three major flavonoids, apigenin (0.0047 mg/g), nicotiflorin (0.034 mg/g), and apigenin 4'-O- β -D-glucoside (0.016 mg/g) from the dry powder of Pueraria flower (Ding et al., 2013). In addition, rutin and luteolin were analyzed using ultra-performance liquid chromatography coupled with quadrupole/time-of-flight mass spectrometry (UPLC-QTOF/MS), their contents were 0.09 mg/g and 0.04–0.07 mg/g, respectively (Lu et al., 2013).

3.2. Isoflavones

Isoflavones are generally considered as the major bioactive com-

pounds in Pueraria flower. Isoflavones from Pueraria flower are usually thought to be chemoprotective and also serve as an alternative therapy for female hormonal disorders including ovarian cancer and menopausal symptoms (Han et al., 2018; Tousen et al., 2019; Yang et al., 2012). Until now, more than 30 isoflavones have been quantified in Pueraria flower (Table 2) and some of their structures are well elucidated (Tong et al., 2018). A summary of the current findings are presented below. Ultrafiltration with liquid chromatography and mass spectrometry (UF-LC-MS) coupled with high-speed counter-current chromatography (HSCCC) are the fundamental tools for rapidly screening and isolating isoflavones from Pueraria flower. Tectoridin and kakkalide were identified as the main isoflavones in Pueraria flower, followed by puerarin, genistin, and tectorigenin which are valuable as α -glucosidase and lactate dehydrogenase (LDH) inhibitors and are effective in drug design for preventing and treating diabetes mellitus and stroke (Wu et al., 2018).

Zhang et al.(2012) used HPLC combined with 2,2'-diphenyl-1-picrylhydrazyl (DPPH) assays to access the antioxidant activity of isoflavones identified in Pueraria flower. In this research, it was found that the antioxidant activity of extracts from Pueraria flower was strongly dependent on the solvent. Solvents with different polarities were used to further fractionate crude ethanolic extract of Pueraria flower. The ethyl acetate fraction showed more potent capacity to scavenge DPPH radical than petroleum ether or n-BuOH fractions (Zhang et al., 2012).

In addition, Lu et al.(2013) determined a total of 25 isoflavones by using ultra-performance liquid chromatography-quadrupole time-of-flight mass spectrometry and mass spectrometry (UPLC -QTOF/MS). According to these authors, kakkalide and irisolidone were abundant in Pueraria flower (10.3–17.7 mg/g kakkalide, and 2.76–4.95 mg/g irisolidone) (Lu et al., 2013). Interestingly, when the estrogenic activity of kakkalide and its metabolite irisolidone were investigated, the results showed that irisolidone had a better estrogenic effect than kakkalide (Shin et al., 2006). Moreover, kakkalide was isolated as a potent 3-hydroxy-3-methylglutaryl (HMG)-CoA reductase (HCR) inhibitor down-regulating the biosynthesis of triacylglycerols and cholesterol (Min and Kim, 2007).

Yuan et al. (2009) found 11 isoflavones from Pueraria flower with therapeutic potential for alcoholism. Among them, two new isoflavones, 6-hydroxybiochanin A-6,7-di-O-β-D-glucopyranoside

Table 2. Major isoflavones in Pueraria flower

$$R_3O$$
 R_2
 R_1
 R_5
 R_5
 R_6
 R_7

| No. | Compounds | R ₁ | R ₂ | R ₃ | R ₄ | R ₅ | R ₆ | Analysis | Reference |
|-----|--|----------------|-------------------|----------------|----------------|----------------|----------------|-----------------------|-------------------------------|
| 1 | Kakkalide | ОН | ОМе | Glc-Xyl | Н | Н | Me | IR, UV | Kubo et al., 1975 |
| 2 | 3'-Hydroxypuerarin | ОН | Н | Н | Glc | ОН | Н | UFLC-MS | Li et al., 2017 |
| 3 | Puerarin | Н | Н | Н | Glc | Н | Н | UFLC-MS | Li et al., 2017 |
| 4 | Puerarinxyloside | Н | Н | Xyl | Glc | Н | Н | UFLC-MS | Li et al., 2017 |
| 5 | Tectoridin | ОН | OMe | Glc | Н | Н | Н | UFLC-MS | Li et al., 2017 |
| 6 | Tectorigenin | ОН | O-CH ₃ | Н | Н | Н | Н | UFLC-MS | Li et al., 2017 |
| 7 | Ononin | ОН | ОН | Glc | Н | Н | Н | UV, MS, TLC | Kurihara and Kikuchi, 1976 |
| 8 | Puerarin-4'- <i>O</i> -β-D-glucopyranoside | Н | Н | Н | Glc | Н | Glc | HPLC-MS/MS | Zhang et al., 2012 |
| 9 | Glycitin | Н | OMe | Glc | Н | Н | Н | UPLC-QTOF/MS | Lu et al., 2013 |
| 10 | Tectorigenin-7- <i>O</i> -β-D-xylosyl- (1→6)-β-D-glucopyranoside | ОН | OMe | Glc-Xly | Н | Н | Н | HPLC-MS/MS | Wang et al., 2013 |
| 11 | Genistein-8- <i>C</i> -β-D-glucopyranoside | ОН | Н | Н | Glc | Н | Н | HPLC-MS/MS | Zhang et al., 2012 |
| 12 | Irisolidone-7- O -β-D-glucopyranpsyl-(1 \rightarrow 6)-β-D-glucopyranoside | ОН | OMe | Glc-Glc | Н | Н | Me | HPLC-MS/MS | Zhang et al., 2012 |
| 13 | Biochanin A-7- \textit{O} - β -D-glucopyranoside | ОН | Н | Glc | Н | Н | Me | HPLC-MS/MS | Zhang et al., 2012 |
| 14 | Daidzein | Н | Н | Glc | Н | Н | Н | IR, NMR | Kurihara and Kiruchi, 1973 |
| 15 | 3'-methoxydaidzin | Н | Н | Glc | Н | OMe | Н | HPLC-MS/MS | Zhang et al., 2012 |
| 16 | Irisolidone | ОН | OMe | Н | Н | Н | Me | IR, TLC | Kurihara and Kiruchi, 1973 |
| 17 | Formononetin | Н | Н | Н | Н | Н | Me | UV, MS | Kurihara and Kiruchi, 1973 |
| 18 | 6-Hydroxygenistein-6,7-di- <i>O</i> -glucoside | ОН | OGlc | Glc | Н | Н | Н | UPLC-QTOF/MS | Lu et al., 2013 |
| 19 | Tectorigenin-7-O-xylosylglucoside | ОН | OMe | Glc-Xyl | Н | Н | Н | UPLC-QTOF/MS | Lu et al., 2013 |
| 21 | 6-Hydroxybiochanin A-6,7- di- <i>O</i> -glucoside | ОН | OGlc | Glc | Н | Me | Н | UPLC-QTOF/MS | Lu et al., 2013 |
| 22 | Gehuain | Н | OMe | Glc-Xyl | Н | Me | Н | UPLC-QTOF/MS | Lu et al., 2013 |
| 23 | Glycitein | Н | OMe | Н | Н | Н | Н | UPLC-QTOF/MS | Lu et al., 2013 |
| 24 | Genistein | ОН | Н | Н | Н | Н | Н | IR, NMR | Kurihara and Kiruchi, 1973 |
| 25 | Biochanin A | ОН | Н | Н | Н | Me | Н | IR, TLC | Kurihara and Kiruchi, 1973 |
| 26 | Tectorigenin-7- O -[β -D-xylopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside] | ОН | OMe | Glc-Xyl | Н | Н | Н | HPLC-ESI-Q/ TOF-MS | Ma et al., 2019 |
| 27 | Genistein-7-glucoside | ОН | Н | Glc | Н | Н | Н | HPLC-ESI-Q/ TOF-MS | Wang et al., 2013 |

Table 2. Major isoflavones in Pueraria flower - (continued)

| No. | Compounds | R ₁ | R ₂ | R ₃ | R ₄ | R ₅ | R ₆ | Analysis | Reference |
|-----|--|----------------|----------------|----------------------|----------------|----------------|----------------|----------------------|-------------------------------|
| 28 | 3'-Hydroxytectorigenin-7- O -β-D-xylosyl-(1 \rightarrow 6)-β-D-glucopyranoside | ОН | OMe | HO OH HO OH | Н | ОН | Н | HR-ESI-MS and NMR | Wang et al., 2013 |
| 29 | Sissotorin | ОН | Н | Glc | Н | Н | Me | UV, MS, TLC | Kurihara and Kikuchi, 1976 |
| 30 | Quercetin | ОН | Н | Н | Н | ОН | Н | IR, NMR | Kurihara and Kikuchi, 1973 |
| 31 | Genistein | ОН | Н | OH _{IIII} O | | Н | Н | UV, MS, TLC | Kurihara and Kiruchi, 1973 |

and 6-hydroxygenistein-7-O-β-D-glucopyranoside, were isolated from the ethanolic extract of the flowers and their structures elucidated by ultraviolet (UV), infrared spectroscopy (IR), high resolution mass spectrum (HR-MS), and 1D and 2D nuclear magnetic resonance (NMR) spectroscopic methods. The release of lipopolysaccharide (LPS)-induced nitric oxide, using primary cultured rat cortical microglia, was tested for all these 11 isoflavones. Tectorigenin, genistein and irisolidone were stronger in inhibiting nitric oxide release activity than gehuain, tectoridin, tectorigenin-7-O-β-D-xylosyl-(1→6)-β-D-glucopyranoside and 6-hydroxygenistein-6,7-di-O-β-D-glucopyranoside. However, there was little inhibitory activity for 6-hydroxybiochanin A-6,7-di-O-β-D-glucopyranoside, 6-hydroxygenistein-7-O-β-D-glucopyranoside, 6-hydroxygenistein-7-O-β-D-glucopyranoside genistin and 6-hydroxygenistein-7-O-β-D-glucopyranoside kakkalide. As far as the structure-activity relationship is concerned, the glycosylation at the C-7 hydroxyl group reduced the inhibitory activity of microglial activation. The methoxylation of 4'-hydroxyl group of 7-glycosylated isoflavones reduced the inhibitory activity, while the methoxy group at the 6-position enhanced the activity (Yuan et al., 2009).

Additionally, prolonged storage of Pueraria flower led to chemical transformation of some compounds. For example, tectoridin in Pueraria flower could be methylated and transform into kakkalide (Kim et al., 2003). Thus, tectoridin is a prodrug of tectorigenin.

3.3. Saponins

Saponins are a large family of amphiphilic glycosides of steroids and triterpenes found in plants and some marine organisms (Yang et al., 2014). Naturally occurring saponins constitute a structurally diverse class of glycosides that are composed of one or more sugar moieties and aglycones linked via glycosidic bonds (Sahu and Achari, 2001). The amounts of total saponins varied from 0.43 to 2.00% according to analysis of Pueraria flower collected from 30 different areas (Niiho et al., 2010). Based on the chemical structure of saponins, a number of saponins identified from Pueraria flower are listed in Table 3.

Lu et al.(2013) simultaneously quantified 12 saponins by UPLC -QTOF/MS analysis. The total content of the saponins was 23.2–60.6 mg/g (Lu et al., 2013). The major saponin compositions are kaikasaponin III (1.26~15.2 mg/g) and soyasaponin I (2.65~19.1 mg/g). The contents of saponins secondary to kaikasaponin III were kaikasaponin II (1.63~5.74 mg/g) and kakkasaponin I (5.08~12.3 mg/g). The rest of saponins identified included soyasaponin IV and baptisiasaponin I, phaseoside IV, astragaloside VIII, kaikasaponin I, azukisaponin I, kakkasaponin II, and kakkasaponin III, which

were detected at a much lower amounts compared to those mentioned above.

The health promoting activities of saponin compositions found in Pueraria flower are reported in the literature. Kaikasaponin III was found to possess hypoglycemic and hypolipidemic effects in the streptozotocin (STZ)-induced diabetic rat (Choi et al., 2004). Soyasaponin I and kaikasaponin III from Pueraria flower have been reported to inhibit testosterone 5α -reductase and to promote hair growth (Murata et al., 2012).

3.4. Essential oils (EOs)

EOs are characterized by volatile and semi-volatile compounds with lower molecular weight (Song et al., 2019). Generally, EOs are secondary plant metabolites containing complex mixtures of volatile organic compounds (Aziz et al., 2018) such as terpenes and their oxygenated derivatives, and some aromatic and aliphatic compounds (Abad et al., 2012).

Kurihara and Kikuchi (1973) identified 13 essential oil components extracted from Pueraria flower. A total of 2.9 g essential oils were obtained, including 2.4 g neutral oil and 0.5 g acid oil. The neutral oils included 1-octen-3-ol, cis-3-hexene-1-ol, benzyl alcohol, eugenol, isoamyl alcohol, octyl alcohol, phenethyl alcohol, llinalool; and acid oil included methyl benzoate, methyl propionate, methyl isovalerate, and methyl caproate (Table 4).

In addition, at least 12 more compounds from Pueraria flower essential oils were identified by gas chromatography-mass spectrometry (GC-MS). These were nonanal, camphor, terpinyl acetate, trans-car-yophyllene, thujopsene, humulene, 2.6-bis (1-dimethylethyl)-4-methyl-phenol, hexahydrofarnesy lacetone, methyl palmitate, dibutyl terephthalate, hexadecanoic acid, n-docosane. Among them, hexahydrofarnesy lacetone was the richest compound with a content of 15.9% and was considered as a spice (Wang et al., 2002).

3.5. Other bioactive compounds

Apart from isoflavones, saponins and essential oils, some other compounds were identified in Pueraria Flower. β -sitosterol, and β -sitosterol-3O- β -D-glucoside were isolated from the methanolic extract of the Pueraria flower.(Kurihara and Kikuchi, 1976) Additionally, 10 compounds were isolated from Pueraria flower including 3,5-di-tert-butyl-4-hydroxybenzaldehyde, palmitic acid, 1-octadecene, octadecanoic acid, eicosanoic acid, squalene, (E)-23-ethylcholesta-5,22-dien-3 β -ol, 24-methylenecycloartanol, β -amyrin, lupenone. Among them, β -amyrin was present atthe highest percent-

Table 3. Major saponins in Pueraria flower (Lu et al., 2013)

| No. | Compounds | R_1 | R ₂ | R ₃ |
|-----|-----------------------------|---------|--------------------|----------------|
| 1 | AstragalosideVIII | H, OH | CH ₂ OH | Xly-rha |
| 2 | SoyasaponinI | H, OH | CH ₂ OH | Gal-rha |
| 3 | SoyasaponinIII | н, β-ОН | CH ₂ OH | Gal |
| 4 | SoyasaponinIV | H, OH | CH ₂ OH | Ara |
| 5 | KaikasaponinIII | H, OH | Me | Gla-rha |
| 6 | KaikasaponinII | H, OH | Me | Glc-rha |
| 7 | KaikasaponinI | H, OH | Me | Gal |
| 8 | KakkasaponinI | H, OH | Me | Ara-rha |
| 9 | AzukisaponinI | H, OH | Me | Glc |
| 10 | Baptisiasaponinl | H, OH | Me | Xyl-rha |
| 11 | PhaseosideIV | 0 | Me | Gal-rha |
| 12 | KakkasaponinII | 0 | Me | Gal |
| 13 | KakkasaponinIII | 0 | Me | Xyl-rha |
| 14 | Sophoradiol monoglucuronide | 0 | Me | Rha-gal |

age of 19.7%, followed by palmittic acid (6.8%), lupenone (5.6%), and 24-methylenecycloartanol (4.0%) (Kim et al., 2015).

4. Pharmacological activities

Both *in vivo* and *in vitro* research suggest that Pueraria flower has a wide range of biological activities including hepatoprotective and estrogenic effects, as well as antioxidant, and anti-inflammation activities. The current findings regarding the main bioactive components of Pueraria flower and their underlying action mechanisms are summarized in this contribution (Table 5).

4.1. Hepatoprotective effect

Akin to other herbal medicines, extracts or compounds from Pueraria flower possess protective effects and therapeutic properties against liver diseases. Among the bioactives, isoflavones obtained from Pueraria flower have been well documented to be active against liver dysfunction and damage caused by liver diseases (Miltonprabu et al., 2017). Tectoridin, one of the main isoflavones in

Pueraria flower, showed significant protective effect on hepatic steatosis induced by ethanol through the modulation of PPAR α pathway and protection of mitochondrial injury. The hepatoprotective effect may be connected with inhibiting the increased levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST) and triacylglycerol (TG), adjusting the levels of mitochondrial permeability transition (MPT) and transmembrane potential ($\Delta \psi m$) (Xiong et al., 2010). Furthermore, tectoridin is an inhibitor of β -glucuronidase and treatment with tectoridin attenuated the increase of β -glucuronidase in blood caused by liver damage. In addition, the hepatoprotective effect of tectoridin could be linked to the metabolism with intestinal bacteria (Lee et al., 2005). Intraperitoneal tectorigenin injection protected mice from CCl₄-induced liver injury by inhibiting the increase of ALT, AST and lactic acid dehydrogenase levels by 22.4, 44.4 and 58.7%, respectively (Lee et al., 2003).

4.2. Estrogenic effects

Estrogenic effects are associated with a variety of physio- or pathological effects, in addition to regulating female reproduction and secondary sex characteristics. Abnormal estrogen is closely asso-

Table 4. Major essential oils in Pueraria flower

| Compounds | Chemical structure | Analysis | Molecular weight | References |
|---------------------|--|------------------|------------------|----------------------------|
| 1-Octen-3-ol | CH ₂ CH ₃ | IR, GLC, IR, GLC | 128.21200 | Kurihara and Kikuchi, 1973 |
| Leaf Alcohol | OH CH ₃ | IR, GLC | 100.15900 | Kurihara and Kikuchi, 1973 |
| Benzyl Alcohol | OH | IR, GLC | 108.14 | Kurihara and Kikuchi, 1973 |
| Eugenol | CH ₂ OH | IR, GLC | 164.2 | Kurihara and Kikuchi, 1973 |
| Isoamyl Alcohol | CH ₃ OH | IR, GLC | 88.1481 | Kurihara and Kikuchi, 1973 |
| Octyl Alcohol | CH ₃ OH | IR, GLC | 130.2279 | Kurihara and Kikuchi, 1973 |
| l-Linalool | OH CH ₃ CH ₃ CH ₃ | IR, GLC | 154.25 | Kurihara and Kikuchi, 1973 |
| Methyl Benzoate | CH ₃ | IR, GLC | 136.15 | Kurihara and Kikuchi, 1973 |
| Methyl Propionate | OH CH ₃ | IR, GLC | 100.159 | Kurihara and Kikuchi, 1973 |
| Methyl Isovalerate | CH ₃ CH ₃ | IR, GLC | 116.16 | Kurihara and Kikuchi, 1973 |
| Methyl Caproate | CH ₃ | IR, GLC | 130.18 | Kurihara and Kikuchi, 1973 |
| Nonanal | 0 | GC-MS | 142.24 | Kurihara and Kikuchi, 1973 |
| Camphor | H ₃ C CH ₃ CH ₃ | GC-MS | 152.23 | Wang et al., 2002 |
| Terpinyl acetate | CH ₃ CH ₃ CH ₃ O | GC-MS | 196.29 | Wang et al., 2002 |
| Trans-caryophyllene | CH ₃ CH ₂ CH ₂ | GC-MS | 204.35 | Wang et al., 2002 |

Table 4. Major essential oils in Pueraria flower - (continued)

| Compounds | Chemical structure | Analysis | Molecular weight | References |
|--|---|-----------|------------------|-------------------|
| Thujopsene | CH ₃ CH ₃ | GC-MS | 204.35 | Wang et al., 2002 |
| Humulene | H ₃ C CH ₃ | GC-MS | 204.35 | Wang et al., 2002 |
| 2.6-Bis(1-dimethylethyl)- 4-methyl-phenol | H ₃ C CH ₃ CH ₃ C CH ₃ | GC-MS | 220.35 | Wang et al., 2002 |
| Hexahydrofarnesyl acetone | $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | GC-MS | 268.48 | Wang et al., 2002 |
| Methyl palmitate | H,C CH | GC-MS | 270.45 | Wang et al., 2002 |
| Dibutyl terephthalate | H ₃ C O | GC-MS | 278.34 | Wang et al., 2002 |
| Hexadecanoic acid | HO | GC-MS CH3 | 256.42 | Wang et al., 2002 |
| n-Docosane | $CH_3(CH_2)_{20}CH_3$ | GC-MS | 310.60 | Wang et al., 2002 |

ciated with broad spectrum of diseases. Pueraria flower is rich in isoflavones, which is the most well-known subgroup of phytoestrogens and plays protective roles against abnormal estrogenic effects (Ososki and Kennelly, 2003; Wang et al., 2020). Therefore, it is not surprising that Pueraria flowers possess estrogenic effects. The estrogenic effect of Pueraria flower has been confirmed by many researchers. Park et al. (2002) found that Pueraria flower could cause significant reversal of stress-induced deficits in learning and memory on a spatial memory task, and also increased choline acetyltransferase (ChAT) immunoreactivities in ovariectomized (OVX) mice. Besides, tectorigenin has estrogenic effect

through attenuating the levels of RANKL, follicle-stimulating hormone (FSH) and luteinizing hormone (LH) and enhancing the levels of estrogen, estrogen receptor (ER)- β , 5-HT1A, 5-HT2A, and tryptophan hydroxylase (Han et al., 2018). In addition, Shin et al. (2006) found that kakkalide was metabolized to irisolidone and tectoridin, which were further metabolized to tectorigenin by human intestinal microflora. Interestingly, kakkalide and tectoridin showed less potent estrogenic effect than their metabolites (Shin et al., 2006).

In addition, Pueraria flower extracts displayed anti-endometriotic effects. Pueraria flower extracts suppressed the adhesion

Table 5. The main bioactives of Pueraria flower.

| Compounds | Bioactivities | Experimental models | Effects | References |
|----------------------------------|---|--|--|-------------------------------------|
| Tectoridin | Hepatoprotective Effect | Ethanol-induced mice | ↓ALT, AST, TG, MDA MPT, Δψm levels.↑PPARα, MCAD, CYP 4A10/14, DGAT, GPAT | Xiong et al., 2010 |
| Kakkalide, irislidone | Hypolipidemic effect | Trition WR1339- induced mice | ↓HCR activity, TC, TG level | Min and Kim, 2007 |
| Kakkalide, irislidone | Anti-inflammation | TNBS-induced mice | ↓NF-кВ activation, M1 macrophage polarization marker expression. ↑gut Proteobacteria population | Jang et al., 2019 |
| Kaikasaponin, Tectorigenin | Antimutagenic, anti-lipid peroxidative effect | Bromobenzene induced rats | ↓Salmonella typhymurium TA100, AFB ₁ , MDA | Park et al., 2002 |
| Tectorigenin | Hepatoprotective Effect | CCl ₄ -induced mice | \downarrow ALT, AST, LDH, MDA, calcium levels, \uparrow GSH, GST activity | Lee et al., 2003 |
| Tectorigenin | Antioxidant | H ₂ O ₂ -induced HUVECs | ↓LDH, MDA levels. Bax and Cleaved Caspase-3. ↑Bcl-2 expression, SOD and GSH-Px. | Chen et al., 2021 |
| Tectorigenin, Tectoridin | Antioxidant | In vitro | ↓hydroxy radical, Superoxide anion radical, DPPH radical, Lipid peroxidation levels | Han et al., 2012 |
| KaikasaponinIII | Antidiabetic | Streptozotocin- induced rat | ↓lipid peroxide, hydroxy radical levels, ↑SOD, TF, Phasel, Phasell enzymes activities | Choi et al., 2004 |
| Tectorigenin, genistein | Antileukemia | HL-60cells | ↓Bcl-2, EGF-receptor expression | Lee et al., 2001 |
| Irisolidone, kakkalide | Anti-gastric injury | ethanol-induced mice | \sqrt{TNF} α, IL-8, IFNγ, COX-2expression, NF-κB activation | Kang et al., 2016 |
| Tectorigenin | Anti-inflammation | Palmitate-stimulated HUVECs | \downarrow ROS production, $\Delta \psi$ m, IKK β /NF- κ B and JNK activation, TNF- α , IL-6 expression, IRS-1serine/tyrosine phoshorylation, NO production, ET-1, VCAM-1 expression | Qi et al., 2013Zhan et al., 2013 |
| Kakkalide, irisolide | Anti-inflammation | Carrageenan-Induced | $\sqrt{\text{TNF}}$ α, IL- β , PGE $_2$, COX-2 expreesion, NF- κ B activation. | Min et al., 2011 |
| Tectorigenin | Antioxidant | MPP+-induced SH-SY5Y cell | ↓cell cytotoxicity and poptosis, Bax/Bcl-2, ROS, NOX, antioxidant enzyme expression | Min et al., 2011 |
| Irisolidone | Hepatoprotective effect | Tert-Butyl Hyperoxide(t-BHP)- Induced mice | ↓cell cytotoxicity, ALT, AST. | Lee et al., 2005 |
| Irisolidone | Antibacterial activity | Helicobacter pylori | ↓H+/K+ ATPase | Bae et al., 2001 |
| KaikasaponinIII, Tectorigenin | Hypoglycemic, hypolipid effect, antioxidant | Streptozotocin- Lnduced rats | ↓glucose, body weight, LDL, VLDL cholesterol, DPPH, XOD, superoxide anion, lipid peroxidation.↑HDL cholesterol | Lee et al., 2000 |
| Puerarin | Antioxidant, anti- inflammation | DSS-induced mice | ↓myeloperoxidase (MPO) activity, NF-кВ, pro-inflammatory mediators, Nrf2↑tight junctions | Jeon et al., 2020 |
| Tectoridin | Estrogenic effects | Ovariectomy- induced mice | ↓osteoclastogenesis. Trap, Ctsk, ATP60, DC-Stamp, c-Fos, and NFATc1, NF-кВ | Wang et al., 2020 |

of human endometriotic11Z and 12Z cells to human mesothelial Met5A cells through targeting extracellular signal regulated kinase

(ERK)1/2 pathway to inhibit matrix metalloproteinase (MMP)-2 and MMP-9 in endometriotic cells (Kim et al., 2017).

4.3. Antioxidant activity

Oxidative stress is an imbalance between the production of reactive oxygen species (ROS) and the antioxidants to scavenge the ROS (Nocella et al., 2019). ROS consist of radical and non-radical oxygen-based molecules, such as hydroxyl radical (•OH), hydrogen peroxide (H₂O₂), singlet oxygen (¹O₂), and superoxide (O₂•-) (Yang et al., 2019). To investigate the correlation between the phytochemicals in Pueraria flower and their antioxidant capacity, measurement of antioxidant capacity of isoflavones with different structures was performed *in vitro*. The results so obtained for antioxidant activity from high to low was tectorigenin sodium sulfona te>tectorigenin>tectoridin, demonstrating that appropriate chemical modifications could greatly improve the biological activities of the naturally occurring products (Han et al., 2012). Eighteen antioxidants were screened and identified from Pueraria flowers by DPPH spiking HPLC-MS/MS (Zhang et al., 2012).

Pueraria flower extracts were beneficial in improving the antioxidative function in ethanol-treated rats by modulating redox enzymes such as Cu/Zn SOD, CAT and GSH-Px(Lee et al., 2001). In addition, tectorigenin prevented MPP+-induced human neuroblastoma SH-SY5Y cells damage due to its potent antioxidant activity. The addition of tectorigenin blocked MPP+-induced ROS formation and NADPH oxidase (NOX) expression to protect the antioxidant enzyme activities from MPP wreckage (Gong et al., 2017). Notably, by means of targeting oxidative stress, tectorigenin and kaikasaponin III were reported to alleviate the streptozotocin-induced toxicity and to contribute to hypoglycemic and hypolipidemic effects (Lee et al., 2000). Besides, puerarin, one of components of Pueraria flower, showed antioxidant effect by regulating the expression of Nrf2 pathway and antioxidant enzymes dextran sulfate sodium-induced colitis mice model.(Jeon et al., 2020).

4.4. Anti-inflammatory activity

Increasing evidence proved that the bioactive extracts of various natural plants including Pueraria flower display a variety of pharmacological effects on acute and chronic inflammatory diseases (Lowry, 1993; Arulselvan et al., 2016). Research from a rat model suggested that methanol extracts of Pueraria flower prevented osteoarthritis by inhibiting the pro-inflammatory mediators iNOS, MMP-9 and MMP-3 in the knee tissues (Sun et al., 2019).

Tectorigenin attenuated endothelial dysfunction associated with insulin resistance through inhibiting ROS-related inflammation and facilitating insulin IRS-1/PI3K/Akt/eNOS signaling pathway (Qi et al., 2013). Another isoflavone, kakkalide ameliorated insulin resistance in human umbilical vein endothelial cells induced by palmitate via inhibiting ROS-associated inflammatory tumor necrosis factor-α (TNF-α) and interleukin-6 (IL-6) production and facilitating insulin PI3K/Akt/eNOS signal pathway (Zhang et al., 2013). Furthermore, observation from LPS-stimulated peritoneal macrophages suggested that kakkalide and irisolidone down-regulated TNF-α, interleukin-1 beta (IL-1β) and cyclooxygenase-2 (COX-2) via the NF-κB pathway (Min et al., 2011). In addition, kakkalide and irisolidone alleviated the inflamed gut by inhibiting TLR4-NF-κB signaling pathway and reversing the transition of M1 into M2 macrophage polarization (Jang et al., 2019).

4.5. Other biological effects

In addition to above activities, extracts or compounds from Pu-

eraria flower are reported to have anti-cancer, anti-allergic, and antimicrobial activities.

As for the anti-cancer activity, tectorigenin was found to enhance paclitaxel cytotoxicity against ovarian carcinoma cells involved in the activation of apoptotic caspases and regulation of the NF-κB and Akt pathways (Yang et al., 2012). Additionally, tectorigenin also exhibited antiproliferative activity against human leukemia HL-60 cells and this activity may be based upon the induction of differentiation and apoptosis (Lee et al., 2001).

Allergic diseases such as asthma and atopic dermatitis are based on IgE-mediated pharmacologic processes of a variety of cell populations such as mast cell and basophils (Park et al., 2004). Orally administered tectoridin can be transformed, by intestinal bacteria, into the more active agent tectorigenin which potently inhibited the passive cutaneous anaphylaxis reaction. In vitro experiments suggest that tectorigenin inhibited the release of β -hexosaminidase from RBL-2H3 cells induced by IgE (Stevens and Austen, 1989). Tectorigenin is also considered as inhibitors for expression of IgE receptor (FceRI), the key molecule triggering the allergic reactions, on human mast cells (Tamura et al., 2010). These findings indicate that tectorigenin has potential to be an antiallergic agent.

Akin to the Pueraria root, PFE has traditionally been used as an anti-amnesic medicine for treatment of alcoholic intoxication. Data from the observation of passive avoidance behavior in mice supported that aqueous extract of Pueraria flower improved the scopolamine-induced memory impairment (Yamazaki et al., 2005).

5. Toxicity

Although very few reports are available so far, the toxicity of PFE or compounds from PFE is still needed. Takano et al. (Takano et al., 2013) performed oral toxicological studies of PFE and their results provided a fundamental reference for further development and clinical translation of functional food based on Pueraria flower. In their acute toxicity study with 14 days observation, no death or abnormalities were observed and the estimated oral LD50 of PFE was higher than 5 g/kg body weight. Likewise, subchronic toxicity study using Sprague-Dawley rats for 90 days showed no apparent toxicological issues. Thus, the corresponding human equivalent dose of PFE for low toxicity was estimated to be 5.0% in the diet.

6. Conclusion

This review provides a comprehensive review on bioactive compounds derived from Pueraria flower. The main baioctive classes of compounds present included isoflavones, saponins, and flavonoids, among others. Although Pueraria flower as a dietary source is broadly used in traditional medicine, toxicological assessments, pharmacokinetics, and the metabolites of phytochemicals needs to be further investigated.

Conflict of interest

The authors declare no competing financial interests.

Funding

This work was supported by grants from Tianjin Innovative Team

Project (TD13-5087), Tianjin Natural Science Foundation (19JC-QNJC12400), and Shangrao Crucial Research and Development Project (19A005).

References

- Abad, M.J., Bedoya, L.M., Apaza, L., and Bermejo, P. (2012). The artemisia L. Genus: a review of bioactive essential oils. Molecules 17: 2542–2566.
- Arulselvan, P., Fard, M.T., Tan, W.S., Gothai, S., Fakurazi, S., Norhaizan, M.E., and Kumar, S.S. (2016). Role of Antioxidants and Natural Products in Inflammation. Oxid Med Cell Longev 2016: 5276130.
- Aziz, Z.A.A., Ahmad, A., Setapar, S.H.M., Karakucuk, A., Azim, M.M., Lokhat, D., Rafatullah, M., Ganash, M., Kamal, M.A., and Ashraf, G.M. (2018). Essential Oils: Extraction Techniques, Pharmaceutical And Therapeutic Potential A Review. Curr Drug Metab 19: 1100–1110.
- Bae, E.A., Han, M.J., and Kim, D.H. (2001). In vitro anti-Helicobacter pylori activity of irisolidone isolated from the flowers and rhizomes of Pueraria thunbergiana. Planta Med 67: 161–163.
- Chan, K. (2003). Some aspects of toxic contaminants in herbal medicines. Chemosphere 52: 1361–1371.
- Chen, X., Zhang, W., Sun, L., and Lian, Y. (2021). Tectorigenin protect HU-VECs from H(2)O(2)-induced oxidative stress injury by regulating PI3K/Akt pathway. Tissue Cell 68: 101475.
- Choi, J., Shin, M.H., Park, K.Y., Lee, K.T., Jung, H.J., Lee, M.S., and Park, H.J. (2004). Effect of Kaikasaponin III Obtained from *Pueraria thunbergiana* Flowers on Serum and Hepatic Lipid Peroxides and Tissue Factor Activity in the Streptozotocin-Induced Diabetic Rat. J Med Food 7: 31–37.
- Choi, J., Shin, M.H., Park, K.Y., Lee, K.T., Jung, H.J., Lee, M.S., and Park, H.J. (2004). Effect of kaikasaponin III obtained from Pueraria thunbergiana flowers on serum and hepatic lipid peroxides and tissue factor activity in the streptozotocin-induced diabetic rat. J Med Food 7: 31–37.
- Ding, H.Y., Chen, Y.Y., Chang, W.L., and Lin, H.C. (2013). Flavonoids from the Flowers of Pueraria Lobata. J. Chin. Chem. Soc. 51: 1425–1428.
- Gong, P., Deng, F., Zhang, W., Ji, J., Liu, J., Sun, Y., and Hu, J. (2017). Tectorigenin attenuates the MPP(+)-induced SH-SY5Y cell damage, indicating a potential beneficial role in Parkinson's disease by oxidative stress inhibition. Exp Ther Med 14: 4431–4437.
- Han, N.R., Nam, S.Y., Hong, S., Kim, H.Y., Moon, P.D., Kim, H.J., Cho, H., Lee, B., Kim, H.M., and Jeong, H.J. (2018). Improvement effects of a mixed extract of flowers of Pueraria thomsonii Benth. and peels of Citrus unshiu Markovich on postmenopausal symptoms of ovariectomized mice. Biomedicine & Pharmacotherapy 103: 524–530.
- Han, T., Cheng, G., Liu, Y., Yang, H., Hu, Y.T., and Huang, W. (2012). In vitro evaluation of tectoridin, tectorigenin and tectorigenin sodium sulfonate on antioxidant properties. Food Chem. Toxicol. 50: 0–414.
- Jang, HM, Park, KT, Noh, HD, Lee, SH, and Kim, DH (2019). Kakkalide and irisolidone alleviate 2,4,6-trinitrobenzenesulfonic acid-induced colitis in mice by inhibiting lipopolysaccharide binding to toll-like receptor-4 and proteobacteria population. International Immunopharmacology 73: 246–253.
- Hyun-Jung Park, S.-M.H. (2009). The Effects of Puerariae Flos on Stressinduced Deficits of Learning and Memory in Ovariectomized Female Rats. Korean J Physiol Pharmacol 13: 85–89.
- Jeon, Y.D., Lee, J.H., Lee, Y.M., and Kim, D.K. (2020). Puerarin inhibits inflammation and oxidative stress in dextran sulfate sodium-induced colitis mice model. Biomed Pharmacother 124: 109847.
- Kang, G.D., Lee, S.Y., Jang, S.E., Han, M.J., and Kim, D.H. (2016). Irisolidone attenuates ethanol-induced gastric injury in mice by inhibiting the infiltration of neutrophils. Mol. Nutr. Food Res.
- Keung, W.M., and Vallee, B.L. (1998). Kudzu root: an ancient Chinese source of modern antidipsotropic agents. Phytochemistry 47: 499– 506.
- Kim, C., Shin, S., Ha, H., and Kim, J.M. (2003). Study of substance changes in flowers of Pueraria thunbergiana Benth. during storage. Archives of Pharmacal Research 26: 210.
- Kim, D.Y., Won, K.J., Hwang, D.I., Yoon, S.W., Lee, S.J., Park, J.H., Yoon, M.S., Kim, B., and Lee, H.M. (2015). Potential skin regeneration activity and chemical composition of absolute from Pueraria thunbergiana

- flower. Natural Product Communications 10: 2009–2012.
- Kim, J.H., Woo, J.H., Kim, H.M., Oh, M.S., Jang, D.S., and Choi, J.H. (2017). Anti-Endometriotic Effects of Pueraria Flower Extract in Human Endometriotic Cells and Mice. Nutrients 9.
- Kubo, M., Sasaki, M., Namba, K., Naruto, S., and Nishimura, H. (1975). Isolation of a new isoflavone from Chinese Pueraria flowers. Chem. Pharm. Bull. 23: 2449–2451.
- Kubo, M., Sasaki, M., Namba, K., Naruto, S., and Nishimura, H. (1975). Isolation of a new isoflavone from Chinese Pueraria flowers. Chem. Pharm. Bull. 23: 2449–2451.
- Kurihara, T., and Kikuchi, M. (1976). Studies on the Constituents of Flowers. VI.: On the Components of the Flower of Pueraria thubergiana BENTH. (3). Journal of the Pharmaceutical Society of Japan 96: 1486–1488.
- Kurihara, T., and Kiruchi, M. (1973). Studies on the constituents of flowers. I. On the components of flower of Pueraria thunbergiana Benth. (Japanese). Yakugaku Zasshi J. Pharm. Soc. Japan 93: 1201–1205.
- Kurihara, T., and Kikuchi, M. (1976). Studies on the Constituents of Flowers. VI. On the Components of the Flower of Pueraria thubergiana BENTH. (3). J. Pharm. Soc. Japan 96: 1486–1488.
- Kurihara, T., and Kiruchi, M. (1973). Studies on the constituents of flowers. I. On the components of flower of Pueraria thunbergiana Benth. (Japanese). Yakugaku Zasshi J. Pharm. Soc. Japan 93: 1201–1205.
- Lee, H.U., Bae, E.A., and Kim, D.H. (2005). Hepatoprotective effect of tectoridin and tectorigenin on tert-butyl hyperoxide-induced liver injury. Journal of Pharmacological Sciences 97: 541–544.
- Lee, H.U., Bae, E.A., and Kim, D.H. (2005). Hepatoprotective Effects of Irisolidone on tert-Butyl Hyperoxide-Induced Liver Injury. Biological & Pharmaceutical Bulletin 28: 531–533.
- Lee, H.W., Choo, M.K., Bae, E.A., and Kim, D.H. (2003). Beta-glucuronidase inhibitor tectorigenin isolated from the flower of Pueraria thunbergiana protects carbon tetrachloride-induced liver injury. Liver International 23: 221–226.
- Lee, K.T., Sohn, I.C., Dong, H.K., Choi, J.W., Sang, H.K., and Park, H.J. (2000). Hypoglycemic and hypolipidemic effects of tectorigenin and kaikasaponin III in the streptozotocin-induced diabetic rat and their antioxidant activityin vitro. Archives of Pharmacal Research 23: 461–466.
- Lee, K.T., Sohn, I.C., Kim, D.H., Choi, J.W., Kwon, S.H., and Park, H.J. (2000). Hypoglycemic and hypolipidemic effects of tectorigenin and kai-kasaponin III in the streptozotocin-Induced diabetic rat and their antioxidant activity in vitro. Arch Pharm Res 23: 461–466.
- Lee, K.T., Sohn, I.C., Kim, Y.K., Choi, J.H., Choi, J.W., Park, H.J., Itoh, Y., and Miyamoto, K. (2001). Tectorigenin, an isoflavone of Pueraria thunbergiana Benth., induces differentiation and apoptosis in human promyelocytic leukemia HL-60 cells. Biol Pharm Bull 24: 1117–1121.
- Lee, K.T., Sohn, I.C., Kim, Y.K., Choi, J.H., Choi, J.W., Park, H.J., Itoh, Y., and Miyamoto, K.I. (2001). Tectorigenin, an Isoflavone of Pueraria thunbergiana BENTH., Induces Differentiation and Apoptosis in Human Promyelocytic Leukemia HL-60 Cells. Biological & Pharmaceutical Bulletin 24: 1117–1121.
- Lertpatipanpong, P., Janpaijit, S., Park, E.Y., Kim, C.T., and Baek, S.J. (2020). Potential Anti-Diabetic Activity of Pueraria lobata Flower (Flos Puerariae) Extracts. Molecules 25: 3970.
- Lertpatipanpong, P., Janpaijit, S., Park, E.Y., Kim, C.T., and Baek, S.J. (2020). Potential Anti-Diabetic Activity of Pueraria lobata Flower (Flos Puerariae) Extracts. Molecules 5.
- Li, C., Miao, X., Li, F., Adhikari, B.K., Liu, Y., Sun, J., Zhang, R., Cai, L., Liu, Q., and Wang, Y. (2019). Curcuminoids: Implication for inflammation and oxidative stress in cardiovascular diseases. Phytother Res 33: 1302–1317.
- Li, M., Zhao, H., Wu, J., Wang, L., Wang, J., Lv, K., Liu, S., Wang, M., Guan, W., Liu, J., Ho, C.T., and Li, S. (2020). Nobiletin Protects against Acute Liver Injury via Targeting c-Jun N-Terminal Kinase (JNK)-Induced Apoptosis of Hepatocytes. J Agric Food Chem 68: 7112–7120.
- Li, S., Li, S., Liu, C., Liu, C., and Zhang, Y. (2017). Extraction and isolation of potential anti-stroke compounds from flowers of Pueraria lobata guided by in vitro PC12 cell model. Journal of Chromatography B Analytical Technologies in the Biomedical & Life Sciences 1048: 111–120.
- Lowry, S.F. (1993). Cytokine Mediators of Immunity and Inflammation. Archives of Surgery 128: 1235–1241.
- Lu, J., Xie, Y., Tan, Y., Qu, J., Matsuda, H., Yoshikawa, M., and Yuan, D.

- (2013). Simultaneous Determination of Isoflavones, Saponins and Flavones in Flos Puerariae by Ultra Performance Liquid Chromatography Coupled with Quadrupole Time-of-Flight Mass Spectrometry. Chemical & Pharmaceutical Bulletin 61: 941–951.
- Lu, J., Xie, Y., Tan, Y., Qu, J., Matsuda, H., Yoshikawa, M., and Yuan, D. (2013). Simultaneous determination of isoflavones, saponins and flavones in Flos Puerariae by ultra performance liquid chromatography coupled with quadrupole time-of-flight mass spectrometry. Chem Pharm Bull (Tokyo) 61: 941–951.
- Ma, Y., Shang, Y., Zhong, Z., Zhang, Y., Yang, Y., Feng, J., and Wei, Z. (2019). A new isoflavone glycoside from flowers of Pueraria Montana var. lobata (Willd.) Sanjappa & Pradeep. Nat. Prod. Res.
- Mi-Kyung Lee, S.-Y.C. (2001). Effects of Puerariae Flos and Puerariae Radix Extracts on Antioxidant Enzymes in Ethanol-Treated Rats. American Journal of Chinese Medicine 29(2): 343–354.
- Miltonprabu, S., Tomczyk, M., Skalicka-Woźniak, K., Rastrelli, L., Daglia, M., Nabavi, S.F., Alavian, S.M., and Nabavi, S.M. (2017). Hepatoprotective effect of quercetin: From chemistry to medicine. Food Chem Toxicol 108: 365–374.
- Min, S.W., and Kim, D.H. (2007). Kakkalide and irisolidone: HMG-CoA reductase inhibitors isolated from the flower of Pueraria thunbergiana. Biol Pharm Bull 30: 1965–1968.
- Min, S.W., and Kim, D.H. (2007). Kakkalide and Irisolidone: HMG-CoA Reductase Inhibitors Isolated from the Flower of Pueraria thunbergiana. Biological & Pharmaceutical Bulletin 30: 1965–1968.
- Min, S.W., Park, Y.J., and Kim, D.H. (2011). Kakkalide and its metabolite irisolidone ameliorate carrageenan-induced inflammation in mice by inhibiting NF-kappaB pathway. Inflammation 34: 344–351.
- Min, S.W., Park, Y.J., and Kim, D.H. (2011). Kakkalide and Its Metabolite Irisolidone Ameliorate Carrageenan-Induced Inflammation in Mice by Inhibiting NF-κB Pathway. Inflammation 34: p.344–351.
- Murata, K., Noguchi, K., Kondo, M., Onishi, M., Watanabe, N., Okamura, K., and Matsuda, H. (2012). Inhibitory activities of Puerariae Flos against testosterone 5α -reductase and its hair growth promotion activities. J Nat Med 66: 158–165.
- Niiho, Y., Nakajima, Y., Yamazaki, T., Okamoto, M., Tsuchihashi, R., Kodera, M., Kinjo, J., and Nohara, T. (2010). Simultaneous analysis of isoflavones and saponins in Pueraria flowers using HPLC coupled to an evaporative light scattering detector and isolation of a new isoflavone diglucoside. J Nat Med 64: 313–320.
- Nocella, C., Cammisotto, V., Pigozzi, F., Borrione, P., Fossati, C., D'Amico, A., Cangemi, R., Peruzzi, M., Gobbi, G., Ettorre, E., Frati, G., Cavarretta, E., Carnevale, R., and SMiLe Group (2019). Impairment between Oxidant and Antioxidant Systems: Short- and Long-term Implications for Athletes' Health. Nutrients 11.
- Ososki, A.L., and Kennelly, E.J. (2003). Phytoestrogens: a review of the present state of research. Phytother Res 17: 845–869.
- Park, E.K., Shin, Y.W., Lee, H.U., Lee, C.S., and Kim, D.H. (2004). Passive cutaneous anaphylaxis-inhibitory action of tectorigenin, a metabolite of tectoridin by intestinal microflora. Biol Pharm Bull 27: 1099–1102.
- Park, K.Y., Jung, G.O., Choi, J., Lee, K.T., and Park, H.J. (2002). Potent antimutagenic and their anti-lipid peroxidative effect of kaikasaponin III and tectorigenin from the flower of Pueraria thunbergiana. Arch Pharm Res 25: 320–324.
- Qi, W., Cheng, X.L., Zhang, D.Y., Gao, X.J., Ling, Z., Qin, X.Y., Xie, G.Y., Kang, L., Yong, Q., and Liu, B.L. (2013). Tectorigenin Attenuates Palmitate-Induced Endothelial Insulin Resistance via Targeting ROS-Associated Inflammation and IRS-1 Pathway. Plos One 8: e66417.
- Sahu, N.P., and Achari, B. (2001). Advances in Structural Determination of Saponins and Terpenoid Glycosides. ChemInform .
- Sheng, Y., Liu, J., Zheng, S., Liang, F., Luo, Y., Huang, K., Xu, W., and He, X. (2019). Mulberry leaves ameliorate obesity through enhancing brown adipose tissue activity and modulating gut microbiota. Food Funct 10: 4771–4781.
- Shin, J.E., Bae, E.A., Lee, Y.C., Ma, J.Y., and Kim, D.H. (2006). Estrogenic effect of main components kakkalide and tectoridin of Puerariae Flos and their metabolites. Biol Pharm Bull 29: 1202–1206.
- Song, X., Wen, X., He, J., Zhao, H., Li, S., and Wang, M. (2019). Phytochemical components and biological activities of Artemisia argyi. Journal of Functional Foods 52: 648–662.
- Stevens, R.L., and Austen, K.F. (1989). Recent advances in the cellular and

- molecular biology of mast cells. Immunol Today 10: 381-386.
- Sun, S., Yuan, L., Li, W., Wang, X., Man, Z., and Li, Y. (2019). Protective Effect of Pueraria Flower in the Treatment of Osteoarthritis Rat by Attenuating Inflammatory Pathway. International Journal of Pharmacology 15: 766–771.
- Takano, A., Kamiya, T., Tsubata, M., Ikeguchi, M., Takagaki, K., and Kinjo, J. (2013). Oral toxicological studies of pueraria flower extract: acute toxicity study in mice and subchronic toxicity study in rats. J Food Sci 78: T1814–1821.
- Tamura, S., Yoshihira, K., Tokumaru, M., Zisheng, X., and Murakami, N. (2010). Inhibitors for expression of IgE receptor on human mast cell from Puerariae Flos. Bioorg Med Chem Lett 20: 3872–3875.
- Wu, T., Liu, C., Huang, Y., Li, S., and Wang, Y. (2018). Simultaneous screening and isolation of activated constituents from Puerariae Flos by ultrafiltration-liquid chromatography-mass spectrometry combined with high-speed counter-current chromatography. J. Sep. Sci.
- Tousen, Y., Takebayashi, J., Kondo, T., Fuchino, H., Kawano, N., Inui, T., Yoshimatsu, K., Kawahara, N., and Ishimi, Y. (2019). Safety and Efficacy Assessment of Isoflavones from Pueraria (Kudzu) Flower Extract in Ovariectomised Mice: A Comparison with Soy Isoflavones. Int. J. Mol. Sci. 20.
- Wang, J., Duan, Y., Zhi, D., Li, G., Wang, L., Zhang, H., Gu, L., Ruan, H., Zhang, K., Liu, Q., Li, S., Ho, C.T., and Zhao, H. (2014). Pro-apoptotic effects of the novel tangeretin derivate 5-acetyl-6,7,8,4′-tetramethylnortangeretin on MCF-7 breast cancer cells. Cell Biochem Biophys 70: 1255–1263.
- Wang, J., Tang, Y., Lv, X., Zhang, J., Ma, B., Wen, X., Bao, Y., and Wang, G. (2020). Tectoridin inhibits osteoclastogenesis and bone loss in a murine model of ovariectomy-induced osteoporosis. Exp Gerontol 140: 111057.
- Wang, L., Zhao, H., Wang, L., Tao, Y., Du, G., Guan, W., Liu, J., Brennan, C., Ho, C.T., and Li, S. (2020). Effects of Selected Resveratrol Analogues on Activation and Polarization of Lipopolysaccharide-Stimulated BV-2 Microglial Cells. J Agric Food Chem 68: 3750–3757.
- Wang, Q., Cheng, X.-L., Li, H., Qin, X.-Y., Ge, C.-Y., Liu, R., Qi, L.-W., and Qin, M.-J. (2013). Application of an efficient strategy for discovery and purification of bioactive compounds from Chinese herbal medicines, a case study on the Puerariae thomsonii Flos. Journal of Pharmaceutical & Biomedical Analysis 75: 25–32.
- Wang, Q., Cheng, X.L., Li, H., Qin, X.Y., Ge, C.Y., Liu, R., Qi, L.W., and Qin, M.J. (2013). Application of an efficient strategy for discovery and purification of bioactive compounds from Chinese herbal medicines, a case study on the Puerariae thomsonii Flos. J Pharm Biomed Anal 75: 25–32.
- Wang, S., Lei, A., Song, E., and ZHao, K. (2002). Study on the volatile components of radix puerariae. China's pharmaceutical affairs 16(2): 107–109.
- Wen, X., Zhao, H., Wang, L., Wang, L., Du, G., Guan, W., Liu, J., Cao, X., Jiang, X., Tian, J., Wang, M., Ho, C.-T., and Li, S. (2020). Nobiletin Attenuates DSS-Induced Intestinal Barrier Damage through the HNF4 α -Claudin-7 Signaling Pathway. J Agric Food Chem 68: 4641–4649.
- Wirngo, F.E., Lambert, M.N., and Jeppesen, P.B. (2016). The Physiological Effects of Dandelion (Taraxacum Officinale) in Type 2 Diabetes. Rev Diabet Stud 13: 113–131.
- Wong, K.H., Li, G.Q., Li, K.M., Razmovski-Naumovski, V., and Chan, K. (2011). Kudzu root: traditional uses and potential medicinal benefits in diabetes and cardiovascular diseases. J Ethnopharmacol 134: 584–607.
- Wu, J., Li, M., He, J., Lv, K., Wang, M., Guan, W., Liu, J., Tao, Y., Li, S., Ho, C.T., and Zhao, H (2019). Protective effect of pterostilbene on concanavalin A-induced acute liver injury. Food Funct 10: 7308–7314.
- Wu, T., Liu, C., Huang, Y., Li, S., and Wang, Y. (2018). Simultaneous screening and isolation of activated constituents from Puerariae Flos by ultrafiltration with liquid chromatography and mass spectrometry combined with high-speed counter-current chromatography. J Sep Sci 41: 4458–4468.
- Xiong, Y., Yang, Y., Yang, J., Chai, H., Li, Y., Yang, J., Jia, Z., and Wang, Z. (2010). Tectoridin, an isoflavone glycoside from the flower of Pueraria lobata, prevents acute ethanol-induced liver steatosis in mice. Toxicology 276: 64–72.
- Xiong, Y., Yang, Y., Yang, J., Chai, H., and Wang, Z. (2010). Tectoridin, an iso-

- flavone glycoside from the flower of Pueraria lobata, prevents acute ethanol-induced liver steatosis in mice. Toxicology 276: 64–72.
- Yamazaki, T., Yaguchi, M., Nakajima, Y., Hosono, T., Niiho, Y., Hibi, Y., Kinjo, J., and Nohara, T. (2005). Effects of an aqueous extract of Puerariae flos (Thomsonide) on impairment of passive avoidance behavior in mice. J Ethnopharmacol 100: 244–248.
- Yang, B., Chen, Y., and Shi, J. (2019). Reactive Oxygen Species (ROS)-Based Nanomedicine. Chem Rev 119: 4881–4985.
- Yang, Y., Laval, S., and Yu, B. (2014). Chemical synthesis of saponins. Adv Carbohydr Chem Biochem 71: 137–226.
- Yang, Y.I., Kyung-Tae, L., Hee-Juhn, P., Jin, K.T., Seok, C.Y., le-Ming, S., and Jung-Hye, C. (2012). Tectorigenin sensitizes paclitaxel-resistant human ovarian cancer cells through downregulation of the Akt and NFkB pathway. Carcinogenesis 12.
- Yu, Y.L., Liao, Y.T., Li, X., Ye, Y., Ke, C.Q., Li, X.Q., Yang, X.Z., and Yao, M.C. (2011). Isoflavonoid glycosides from the flowers of Pueraria lobata. J Asian Nat Prod Res 13: 284–289.
- Yuan, D., Xie, Y.Y., Bai, X., Wu, X., Yang, J.Y., and Wu, C.F. (2009). Inhibitory activity of isoflavones of Pueraria flowers on nitric oxide production from lipopolysaccharide-activated primary rat microglia. J Asian Nat Prod Res 11: 471–481.
- Zhang, B., Li, W., and Dong, M. (2017). Flavonoids of Kudzu Root Fermented by Eurtotium cristatum Protected Rat Pheochromocytoma Line

- 12 (PC12) Cells against H₂O₂-Induced Apoptosis. Int. J. Mol. Sci. 18.
- Zhang, D., Gao, X., Wang, Q., Qin, M., and Liu, B. (2013). Kakkalide ameliorates endothelial insulin resistance by suppressing reactive oxygen species-associated inflammation. J. Diabetes 5.
- Zhang, D., Gao, X., Wang, Q., Qin, M., Liu, K., Huang, F., and Liu, B. (2013). Kakkalide ameliorates endothelial insulin resistance by suppressing reactive oxygen species-associated inflammation. J Diabetes 5: 13–24.
- Zhang, L., Wen, X., Li, M., Li, S., and Zhao, H. (2018). Targeting cancer stem cells and signaling pathways by resveratrol and pterostilbene. Biofactors 44: 61–68.
- Zhang, Y.P., Shi, S.Y., Xiong, X., Chen, X.Q., and Peng, M.J. (2012). Comparative evaluation of three methods based on high-performance liquid chromatography analysis combined with a 2,2'-diphenyl-1-picrylhydrazyl assay for the rapid screening of antioxidants from Pueraria lobata flowers. Anal Bioanal Chem 402: 2965–2976.
- Zhang, Y.P., Shi, S.Y., Xiong, X., Chen, X.Q., and Peng, M.J. (2012). Comparative evaluation of three methods based on high-performance liquid chromatography analysis combined with a 2,2'-diphenyl-1-picrylhydrazyl assay for the rapid screening of antioxidants from Pueraria lobata flowers. Analytical & Bioanalytical Chemistry 402: 2965–2976.
- Zhou, Y.X., Zhang, H., and Peng, C. (2014). Puerarin: a review of pharmacological effects. Phytother Res 28: 961–975.