





Composition, polyphenol bioavailability, and health benefits of aronia berry: a review

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Abstract

Aronia berries (*Aronia melanocarpa* and *Aronia mitschurinii*) are underutilized functional food, rich in bioactives. Aronia berries have abundant levels of anthocyanins, proanthocyanidins, flavonols, and phenolic acids that may reduce the risk of non-communicable diseases such as diabetes, metabolic syndrome, and neurological disease. Aronia polyphenols are bioavailable, and the majority are transformed into low molecular-weight phenolics. The impact of biotransformation on aronia polyphenols health effects is not fully understood. The objective of this review is to analyze aronia berry composition, including polyphenols nutrients. Additionally, this review summarizes recent preclinical and clinical studies on the polyphenol bioavailability and health benefits upon aronia berry consumption to better understand its potential as a functional food.

Keywords: Aronia berry; Anthocyanin; Chronic disease; Composition; Polyphenol.

1. Introduction

Aronia Medik. (chokeberries), are deciduous shrubs and a member of the Rosaceae family (Sidor and Gramza-Michałowska, 2019; Mahoney et al., 2019). Aronia berry color ranges from red, purple, to black, depending on the species (Sidor and Gramza-Michałowska, 2019). The four main Aronia species include A. arbutifolia (L.), A. melanocarpa (Michx.), A. prunifolia (Marshall), and A. mitschurinii (Mahoney et al., 2019; Kulling and Rawel, 2008). A. mitschurinii is the primary species used for commercial aronia berry production in North America (Brand et al., 2017; Mahoney et al., 2019). The 'Nero' and 'Viking' cultivars of A. mitschurinii are crosses between A. melanocarpa and Sorbus aucuparia L. and grow from 3 to 6 feet tall (Mahoney et al., 2019; Kulling and Rawel, 2008). From May to June, white flowers grow on the shrubs, and by late August and September, the berries are fully matured, with a diameter between 6.1 to 17.8 mm (Kulling and Rawel, 2008).

Aronia berry is consumed as whole berries, but most of the crop is processed to juice, juice concentrates, extracts, fruit powders, jams, or fermented products. Aronia berry is rich in polyphenols, which may contribute to its health benefits. Aronia berries contain a mixture of polyphenolic components and have abundant anthocyanins and proanthocyanidins (Taheri et al., 2013). Although the *in vivo* bioactivity and bioavailability of polyphenols are not fully understood, the antioxidant and antiinflammatory actions of polyphenols may decrease the risk of cardiovascular disease, metabolic syndrome, inflammation, and neurodegenerative disease (Jakobek and Seruga, 2012; Bhaswant et al., 2017). The mechanisms by which anthocyanins, proanthocyanidins, and other polyphenols found in aronia berries are an active reseach area.

Aronia berry remains an underutilized functional food. Prior reviews have addressed aronia berry composition (Sidor and Gramza-Michałowska, 2019), polyphenol bioavailability (Denev, Kratchanov, Ciz, Lojek, and Kratchanova, 2012), and health benefits (Sidor and Gramza-Michałowska, 2019). Additional reports on the composition, polyphenol bioavailability, and bioactive mechanisms have been published since these publications. Therefore, this paper aims to provide an updated and expanded review of the Composition, polyphenol bioavailability, and health benefits of aronia berry: a review

Sugar	Mass in fruit (g/100 g fwb)	Mass in juice (g/100mL)
total sugars	6.21–42.10 ^{a,g}	8.9–19.6 ^{e,f}
fructose	2.2–15.8 ^b	1.5–4.1 ^{e,f}
glucose	1.09–5.7 ^{c,d}	1.5–4.2 ^{e,f}
sorbitol	4.36–12.99 ^{c,b}	3.5–7.7 ^e
sucrose	0.07–1.53 ^{b,d}	0.03–6.8 ^{e,f}
inositol	0.0684 ^g	_*
maltose	0.0496 ^g	-
glycerol	0.00983 ^g	-

Table 1. Aronia berry and juice sugar content and profile

Abbreviations: fwb, fresh weight basis. [†]Data not available in literature. ^aOchmian et al., 2012. ^bDenev et al., 2018. ^cMayer-Miebach et al., 2012. ^dŠnebergrová et al., 2014. ^eSosnowska et al., 2015. ^fHandeland et al. 2014. ^gVázquez-Espinosa et al. 2019.

current scientific literature on aronia berry composition, the pharmacokinetics of the polyphenols, and its potential for improving health. We expect that characterizing the progress and knowledge gaps in these areas will accelerate research, development, and aronia berry utilization.

2. Methods

Publications were identified through Medline, Elsevier, Google Scholar, and Pubmed databases using keywords such as aronia, antioxidants, anthocyanins, bioavailability, chokeberry, proanthocyanidins, cancer, cardiovascular disease, diabetes, functional foods, and polyphenols. Studies were limited from 2010 until 2020 and from 2015 for preclinical and human intervention studies. Compositional data were compiled for black aronia berry (*A. melanocarpa* and *A. mitschurinii*).

3. Aronia composition

The functional components in aronia berry include nutrients, polyphenols, fiber, and sorbitol. Other components such as organic acids, protein, and lipids contribute to fruit quality and stability. The abundance and distribution of these components vary significantly among the studies reviewed in this paper. Variability may arise from aronia genetic variability, environment (location, humidity, temperature, rain, fertilizers, and infections), harvest

Table 2. Aronia berry pomace fiber content

time, and other factors (Veberic et al., 2015). Furthermore, analysis may further introduce variability from extraction methods (e.g. selection of solvent, berry particle sizes, solid-solvent ratio, time, and temperature) or analytical approach (e.g. HPLC vs. gas chromatography, specific or non-specific methods) (Denev et al., 2018).

3.1. Carbohydrates

Aronia berry carbohydrates are primarily sugars and fiber (Tables 1 and 2). Fresh aronia berry contains 15 to 20.9 g/100 g fwb of carbohydrates (Sidor and Gramza-Michałowska, 2019). The sugars in aronia berry and juice are mainly fructose and sorbitol with lower amounts of glucose and sucrose, ranging from 6.2 to 20.9 g/100 g fwb or 8.9 to 19.6 g/100 mL. Sorbitol is abundant in aronia berry with 4.36–12.99 g/100 g fwb of the whole berry. As a sugar alcohol, sorbitol contains about 2.6 calories per gram and has diuretic, laxative, and cathartic properties (US Food and Drug Administration, 2020; Featherstone, 2015). A majority of the berry sugars are extracted into juice, whereas its fiber is mainly distributed in the pomace. The pomace contains 57.8 to 71.5 g/100 g dwb of total dietary fiber and insoluble fiber at 43.8 to 61.7 g/100 g dwb (Schmid et al., 2020). The berry pomace fibers include cellulose (34 g/100 g dwb), hemicellulose (32 g/100 g dwb), lignin (22.7 g/100 g dwb), and pectin (7.52 g/100 g dwb). Although these fibers' structures and solubility vary, increased fiber reduces cardiovascular disease risk, aids in glycemic control, and helps maintain a healthy weight (Schmid et al., 2020;

Fiber		Mass in pomace (g/100 g dwb)		
total fiber		57.8–71.6 ^a		
insolul	ole fiber	43.8–61.7ª		
soluble	e fiber (high molecular weight)	7.3–15.1 ^a		
soluble	e fiber (low molecular weight)	0.8–2.6ª		
lignin		22.68 ^b		
cellulo	se	34.56 ^b		
total pectin		7.52 ^b		
hemice	ellulose	32.08 ^b		

Abbreviations: dwb, dry weight basis. ^aSchmid et al., 2020. ^bNawirska and Uklańska, 2008.

Protein or amino acid	Mass in fruit (mg/100 g fwb)	Pomace (mg/100 g dwb)
total protein	700 ^a	4,900–24,000 ^b
aspartic acid (mg)	0.01–0.035 ^c	0.891 ^d
glutamic acid (mg)	0.04–0.029 ^c	1.979 ^d
serine (mg)	0.023–0.039 ^c	0.368 ^d
histidine (mg)	0.007–0.008 ^c	0.247 ^d
glycine (mg)	0.009–0.018 ^c	0.638 ^d
threonine (mg)	0.033–0.039 ^c	0.298 ^d
arginine (mg)	0.01–0.013 ^c	0.791 ^d
alanine (mg)	0.015–0.022 ^c	0.41 ^d
tyrosine (mg)	0.003–0.006 ^c	0.247 ^d
valine (mg)	0.021–0.025 ^c	0.421 ^d
methionine (mg)	ND-0.001 ^c	0.177 ^d
phenylalanine (mg)	0.004–0.006 ^c	0.43 ^d
isoleucine (mg)	0.008–0.012 ^c	0.378 ^d
leucine (mg)	0.007–0.01 ^c	0.686 ^d

Table 3.	Protein content and	amino acid	profile of aro	nia berry and pomace
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Abbreviations: fwb, fresh weight basis; dwb, dry weight basis; ND, not detected. aLancrajan, 2012. bSójka et al., 2013. cHwang and Thi, 2016. dPieszka et al., 2015.

Slavin, 2013).

3.2. Protein and amino acids

Similar to other berries, the protein and amino acid content of aronia berry is relatively low (Table 3), 700 mg/100 g fwb and 4,900 to 24,000 mg/100 dwb. Most of the protein and amino acids are distributed in the pomace (Sidor and Gramza-Michałowska, 2019). Aronia berry contains both essential and nonessential amino acids. Threonine (0.033 to 0.39 mg/100 g fwb berry) is the most abundant among the amino acids, followed by serine (0.023 to 0.39 mg/100 g fwb berry).

3.3. Lipids

Aronia berry has a low lipid content, with a total fat of 0.14 % in a fresh berry (Table 4). A significant amount of the lipids are in the seed oil, represented mostly by sterols and phospholipids (Sidor and Gramza-Michałowska, 2019). The remaining lipids are mainly in the pomace, primarily polyunsaturated fatty acids, which is 90.49% of total fatty acids, whereas saturated fatty acid is 9.51%. Among the different types of fatty acids, the linoleic (C18:2) and oleic acids (C18:1) are abundant, at 43.43% and 16.38% of fatty acids, respectively (Sidor and Gramza-Michałowska, 2019). While the lipid content of aronia berry is low, processing strategies to recover lipids from aronia seeds and pomace could be used to develop new sources of seed oils with high proportions of unsaturated fatty acids.

3.4. Vitamins and minerals

Aronia berry contains vitamin A, vitamin E, and vitamin C (Table 5). Total carotenoids (including α -, β -, and ζ -carotenes) can be up to 97.8 μ g/L of aronia berry juice (Oprea et al., 2014; Sidor and Gramza-Michałowska, 2019). However, vitamin C is the most abundant micronutrient, with 7.25 to 98.75 mg/100 g fwb (Catană et al., 2017). Aronia berries also contain a variety of minerals. The ash content of aronia berry is 0.37 to 0.49 g/100 g fwb, providing major minerals (calcium, magnesium, phosphorus, potassium) and trace minerals (iron, copper, iodine, zinc, and selenium) (MedlinePlus, 2019). Lead and other heavy metals have been reported in aronia berry, but these levels are below thresholds where toxicity is a concern (Juranović Cindrić et al., 2017).

3.5. Organic acids

The content of organic acids in aronia leads to its sour flavor (Famiani et al., 2015). The titratable acidity is in range with other berries at 0.85 to 1.22% (Bolling et al., 2015). At least eight organic acids are present in aronia berry, including quinic, malic, ascorbic, shikimic, citric, oxalic, succinic, and isocitric acids (Table 6). Fumaric and tartaric acids were also reported in juice. Quinic acid is the most abundant organic acid in aronia berry (293 to 591 mg/100 g fwb), followed by malic acid (308 to 350 mg/100 g fwb), but others have reported that malic acid is the primary organic acid (Denev et al., 2018; Kulling and Rawel, 2008).

3.6. Polyphenols

Aronia berry has a significantly higher polyphenol and antioxidant content than most fruits and vegetables (Nour et al., 2015; Jakobek and Seruga, 2012; Pérez-Jiménez, 2010). Aronia berry total polyphenols range from 1.0 to 3.6 g/100 g fwb (Denev et al., 2018; Nour et al., 2015). The primary polyphenols in aronia berries include anthocyanins, proanthocyanidins, flavonols, and phenolic acids. These polyphenols contribute to the health ben-

Lipid component	Fruit (%) [†]	Pomace (%) [†]	Seed oil (g/kg) [†]
total fat	0.14 ^a	2.9–13 ^b	-
phospholids	-	-	2.8 ^d
sterols	-	-	1.2 ^d
tocopherols	-	-	0.0555 ^d
fatty acid			
C8:0	-	0.02 ^c	-
C12:0	-	0.07 ^c	ND ^d
C14:0	-	0.12 ^c	tr ^d
C16:0	-	5.48 ^c	5.1 ^d
C16:1	-	0.19 ^c	tr ^d
C18:0	-	1.71 ^c	1.1 ^d
C18:1	-	16.38 ^c	21.4 ^d
C18:2	-	43.43 ^c	71.1 ^d
Gamma 18:3	-	0.03 ^c	-
C18:3	-	29.78 ^c	0.5 ^d
CLA c9-t11	-	0.01 ^c	-
C18:4	-	0.02 ^c	-
C20:0	-	1.52 ^c	ND ^d
C20:1	-	0.33 ^c	-
C20:2	-	0.14 ^c	-
C22:0	-	0.59 ^c	0.8 ^d
C22:5	-	0.17 ^c	-
SFA	-	9.51 ^c	0.7 ^d
UFA	-	90.49 ^c	-
PUFA n-6	-	43.6 ^c	-
PUFA n-3	-	29.97 ^c	-
PUFA n-6/n-3	-	1.45 ^c	-

Table 4. Lipid content and profile of aronia berry fruit and pomace.

Abbreviations: CLA, conjugated linoleic acid; SFA, saturated fatty acid; UFA, unsaturated fatty acid; ND, not detected; tr, less than 0.1 g/kg; PUFA, polyunsaturated fatty acid. [†]Total fat on fresh weight basis. [‡]Data not available in literature. ^aLancrajan, 2012. ^bSójka et al., 2013. ^cPieszka et al., 2015. ^dZlatanov, 1999.

efits as well as the astringent and bitter flavor associated with aronia berry.

3.6.1. Anthocyanins

Within black aronia berry, anthocyanins are the most abundant polyphenol and pigment (Wathon et al., 2019; Sidor and Gramza-Michałowska, 2019). The structure of anthocyanins determines the pigmentation of the fruit by the type of aglycon base or flavylium ring, sugar, and acylation (Sidor and Gramza-Michałowska, 2019). Anthocyanins also reduce photooxidation and limit CO_2 assimilation in plant tissue. Thus, most of the anthocyanins are located on the skin's external layer in the pomace (Veberic et al., 2015). There are six major anthocyanin aglycons in berries: cyanidin, delphinidin, petunidin, peonidin, pelargonidin, and malvidin, which typically contain a sugar moiety (Bueno et al., 2012). In aronia berry, cyanidin glycosides account for 90 to 98.7% of the total anthocyanin content (Veberic et al., 2015; Denev et al., 2018). The major cyanidin glycosides are include its 3-galactoside (126 to 990 mg/100 g fwb), 3-glucoside (1.7 to 21.5 mg/100 g fwb), 3-arabinoside (52 to 399 mg/100 g), 3-xyloside (2.7 to 81.2 mg/100 g fwb), 3,5-hexoside (epi)catechin (14.3 mg/100 g dwb), 3-pentoxide-(epi)catechin (7.26 mg/100 g dwb), 3-hexoside-(epi)cat-(epi) cat (13.6 mg/100 g dwb) (Table 7). On a fresh weight basis, aronia berries are among the richest dietary sources of anthocyanins (Denev et al., 2018; Pérez-Jiménez et al., 2010).

3.6.2. Proanthocyanidins

Aronia contains proanthocyanidins with predominately (–)-epicatechin units (32.2 to 99.6 mg/100 g) with a trace amount of (+)-catechin (Table 8, Jurikova et al., 2017). Oligomeric and polymeric

Component		Mass in fruit (mg/100 g fwb)
vitamins	vitamin C	7.25–98.75ª
	vitamin A	0.77 ^b
	vitamin E	0.008-0.031 ^c
	vitamin B2	0.873 ^h
	vitamin B5	2.845 ^h
	vitamin B6	1.132 ^h
	vitamin B7	0.615 ^h
nitrate/nitrite	nitrate	4.520–9.850 ^d
	nitrite	0.062–0.187 ^d
minerals	Na	0.427–1.18 ^{e,f}
	К	135–679 ^{g, f}
	Са	11.9–116.7 ^{g,e}
	Mg	8.3–66.9 ^{g,f}
	Р	23.9–95.6 ^e
	Zn	0.055–0.84 ^{f,e}
	Fe	0.132–1.42 ^{f,e}
	Se	0.0021–0.0028 ^f
	Cu	0.82–.211 ^f
	Мо	0.0016-0.0021 ^f
	Mn	0.132–1.789 ^f
	Ni	0.0143–0.0740 ^f
	V	0.0040–0.0158 ^f
	Si	0.237–0.637 ^f
	Cr	0.035–0.211 ^f
	Li	0.0016-0.0021 ^f
	Sr	0.132–1.789 ^f
	Al	0.288–0.440 ^f
	Sn	0.062–0.072 ^f
	As	0.020–0.036 ^f
	Cd	0.0016-0.0041 ^f
	Ва	0.148–0.666 ^f
	Pb	0.0048-0.0091 ^f
	Sb	ND-0.029 ^f
	Со	0.0019–0.0043 ^f
	В	0.288–1.422 ^f

Table 5. Micronutrient, nitrate/nitrite, and mineral content of aronia berry fruit

Abbreviations: fwb, fresh weight basis. ^aCatană et al., 2017. ^bLancrajan, 2012. ^cBorowska and Brzóska, 2016. ^dOchmain et al., 2012. ^ePavlović et al., 2015. ^fJuranović Cindrić et al., 2017. ^gSnebergrová et al., 2014. ^hAsănică et al., 2019.

(-)-epicatechins make up monomers, dimers, tetramers, hexamers, octamers, and decamers, but most proanthocyanidins in aronia berries have polymerization greater than 10 (Taheri et al., 2013). Aronia berry proanthocyanidins are distributed 70% in the flesh, 25% in the skin, and 5% in the seeds (Mayer-Miebach et al., 2012).

Proanthocyanidin consumption may have a wide range of human health benefits, including reducing oxidative stress, improving blood circulation, and reducing cancer symptoms (Rauf et al., 2019). Additionally, when used as food ingredients, proanthocyanidins create foamability, oxidative stability, and heat stability and

Acid	Mass in fruit (mg/100 g fwb)	Mass in juice (mg/100 mL)
quinic acid	293–474 ^a	280 ^b
malic acid	308–350ª	708 ^b
ascorbic acid	56. 9–72.2ª	4 ^b
shikimic acid	8.76–8.99ª	6 ^b
citric acid	31.1–33.5 ^a	7 ^b
oxalic acid	3.21–3.39ª	_*
succinic acid	7.08–7.48 ^a	-
fumaric acid	-	5.1-10.7 ^d
tartaric acid	-	32.1–207 ^d
isocitric acid	2.25–3.73 ^c	-

 Table 6. Organic acid content of aronia berry and juice

Abbreviations: fwb, fresh weight basis. [†]Data not available in literature. ^aDenev et al., 2018. ^bMarkkinen et al., 2019. ^cŠnebergrová et al., 2014. ^dDjuric et al., 2015.

An	hocyanin	Mass in fruit (mg/100 g fwb)	Mass in juice (mg/100 mL)	Mass in pomace (mg/100 g dwb)
tot	al anthocyanins	284–631ª		
	cyanidin-3-galactoside	126–990 ^b	87.4–94.0 ^a	4520–9760 ^f
	cyanidin-3-glucoside	trace-21.5 ^{b,c}	10.2–13.5 ^a	21–225.8 ^f
	cyanidin-3-arabinoside	52–392 ^{c,d}	24.7–58.6 ^a	1840-3120 ^f
	cyanidin-3-xyloside	2.7-81.2 ^{c,d}	0.48–1.25 ^a	167–275 ^f
	pelargonidin-3-arabinoside	5.04 ^e		
	cyanidin-3,5-hexoside-(epi)catechin			14.3 ^f
	cyanidin-3-pentoside-(epi)catechin			7.26 ^f
	cyanidin-3-hexoside-(epi)cat-(epi)cat			13.6 ^f

Abbreviations: fwb, fresh weight basis; dwb, dry weight basis. ^aDenev et al., 2018. ^bBorowska and Brzóska, 2016. ^cWangensteen et al., 2014. ^dHwang and Thi, 2016. ^eVeberic et al., 2015. ^fOszmiański and Lachowicz, 2016.

Table 8. Tannin content and profile of aronia berry and pomace

Proanthocyanidin	Mass in fruit (mg/100 g fwb) [†]	Mass in pomace (mg/100 g dwb) [‡]
total tannins	522–1000 ^a	6200–9720 ^c
monomers (total)	5.17 ^b	-
(–)-epicatechin	62.9–124 ^a	-
dimers	12.5ª	-
procyanidin B2	-	21.9–28.2 ^c
trimers	10.3ª	-
tetramers	0.7 ^a	-
pentamers	0.75ª	-
hexamers	1.04 ^a	-
heptamers	0.56ª	-
octamers	0.51ª	-
decamers	0.16 ^a	-
>10-mers	69ª	

Abbreviations: fwb, fresh weight basis; dwb, dry weight basis; -, data not available in literature. [†]Data for monomers through > 10-mers as (-)-epicatechin equivalents. [‡]Data for monomers through > 10-mers as (+)-catechin equivalents. ^aDenev et al., 2018. ^bDudonné et al., 2015. ^cOszmiański and Lachowicz, 2016.

Flavonols	Mass in fruit (mg/100 g fwb)	Mass in juice (mg/100 mL)	Mass in pomace (mg/100 g dwb)
quercetin-3-rutinoside	3.9–61.7ª	15.5–62.8ª	22.7–43.7 ^d
quercetin-3-glucoside	4.4–29.2 ^{a,b}	11.5–36.7ª	32.7–67.1 ^d
quercetin	6.5–30.2 ^{a,c}	6.8–8.2ª	_
quercetin-3-galactoside	6.6–30.2 ^{a,c}	-	49.0–102 ^d
quercetin-3-robinobioside	1.03–11.3 ^a	-	14.9 ^e
quercetin-3-vicianoside	2.6–4.3 ^a	-	19.6 ^e
isorhamnetin pentoside hexoside	1.12 ^f	-	_
isorhamnetin 3-O-neohesperidoside	1.16 ^f	-	_
isorhamnetin 3-O-rutinoside	0.83 ^f	-	-

Table 9.	Flavonol	content	reported in	aronia be	rry fruit,	juice, and	pomace
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Abbreviations: fwb, fresh weight basis; dwb, dry weight basis; –, data not available in literature. ^aDenev et al., 2018. ^bOchmian et al., 2012. ^cBorowska and Brzóska, 2016. ^dOszmiański and Lachowicz, 2016. ^eRodríguez-Werner et al., 2019. ^fTian et al., 2017.

increase astringency (Shi et al., 2003).

3.6.3. Flavonols

Flavonols are less abundant in aronia berry relative to proanthocyanidins and anthocyanins. The major flavonol in aronia is quercetin (Table 9), and it can increase by 5% during juice pasteurization (Jurikova et al., 2017). Quercetin distributed as its 3-rutinoside (3.9 to 61.7 mg/100 g fwb), 3-glucoside (4.03 to 29.2 mg/100 g fwb), 3-galactoside (6.6 to 30.2 mg/100 g fwb), 3-robinobioside (1.03 to 11.3 mg/100 g fwb), 3-vicianoside (2.36–5.38 mg/100 g fwb) (Table 9). Other aronia berry flavonols include myricetin, isorhamnetin, and kaempferol.

3.6.4. Phenolic acids

The phenolic acid profile of aronia berries is mainly neochlorogenic and chlorogenic acids with lower levels of other phenolic acids (Table 10). These other phenolic acids include vanillic, ferulic, syringic, and gallic acids.

3.6.5. Non-extractable polyphenols

In the studies reviewed, most of the experiments have used analytical methods to assess extractable polyphenols (EPs). The solvents used to obtain EPs are typically aqueous-organic, commonly containing water, methanol, or acetone (Han et al., 2019). In contrast, non-extractable polyphenols (NEPs), also known as insolublebound phenolics, require enzymatic, acidic, or alkaline hydrolysis to be liberated to the extraction medium (de Camargo et al., 2016). These methods dissociate non-extractable polyphenols from cellulose, hemicellulose, polysaccharides, and polypeptides. Advanced techniques, such as ultrasound-assisted extraction and microwaveassisted extraction, are not able to successfully extract NEPs. The most effective method to release the NEPs is hydrolysis (Dzah et al., 2020). However, hydrolysis conditions may degrade polyphenols. After juicing, aronia pomace is a good source of NEPs.

Table 10. Phenolic acid content reporte	d in aronia berry	fruit, juice and	l pomace
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Phenolic acid	Mass in fruit (mg/100 g fwb)	Mass in juice (mg/100 mL)	Mass in pomace (mg/100 g dwb)
neochlorogenic acid	59–186 ^{a,b}	41.6–172 ^{a,f}	169 ^j
chlorogenic acid	17–188 ^{a,c}	88.6–139ª	204 ^j
caffeic acid	60–75 ^{a,c}	0.12-0.18 ^g	_
protocatechuic acid	0.77 ^d	2.49–5.74 ^f	-
vanillic acid	0.25–0.46 ^c	-	_
p-Coumaric acid	0.02 ^d	-	-
dicaffeoylquinic acid	3.74 ^e	-	_
ferulic acid	0.01 ^d	1.99 ^h	_
syringic acid	-	-	4.16 ^k
ellagic acid	1.57 ^d	-	-
gallic acid	-	0.004–0.69 ^{h,i}	_
salicylic acid	_	_	-

Abbreviations: fwb, fresh weight basis; dwb, dry weight basis; –, data not available in literature. ^aDenev et al., 2018. ^bOchmian et al., 2012. ^cHwang and Thi, 2016. ^dDudonné et al., 2015. ^eTian et al., 2017. ^fSosnowska et al., 2016. ^gPozderović et al., 2016. ^hValcheva-Kuzmanoca et al., 2013. ⁱWilkoska et al., 2017. ^jRodríguez-Werner et al., 2019. ^kSop et al., 2013.

Enzyme-assisted extraction and high-pressure extraction methods have also been applied to recover NEPs from aronia berry pomace (Grunovaitė et al., 2016; Kitrytė et al., 2017); however, alkaline hydrolysis is expected to be the most efficient method to recover NEPs (de Camargo et al., 2016). The profile of aronia berry NEPs recovered by enzyme- assisted extraction and high-pressure extraction is similar to its free polyphenol profile (Grunovaitė et al., 2016; Kitrytė et al., 2017).

3.7. Astringent compounds

Although aronia berries show potential for numerous health benefits, consumption of whole aronia berry is limited by its astringency and bitterness. Adding sugar or ethyl butyrate to aronia juice may reduce its bitterness, but does not change astringency and consumers' preference (Duffy et al., 2016). Amygdalin is an aromatic cyanogenic glucoside compound that is responsible for a bitter-almond smell and contributes to aronia berry astringency. There are no recent studies on amygdalin content in aronia berries, but prior studies have reported 52.3 mg/100 g fwb in pomace (Kulling and Rawel, 2008). Proanthocyanidins also contribute to astringency, due to its interactions with salivary proteins (Soares et al., 2018). Some hydroxycinnamic acids, including vanillic and syringic acid, may also contribute to astringency (Sáenz-Navajas et al., 2010). Additional research to characterize the relative astringency of these components in aronia berries can help inform product development and masking strategies in food production.

4. Polyphenol bioavailability

Aronia berries contain higher antioxidant capacity than most foods (Pérez-Jiménez et al., 2010), but these tests neglect polyphenol metabolism and bioavailability. Thus, to understand the antioxidant and human health-promoting mechanisms of aronia polyphenol consumption, it is necessary to consider the metabolism and bioavailability of its polyphenols. While reports on polyphenol bioavailability have increased, there is still limited knowledge about the dynamics of polyphenol metabolism (Neilson et al., 2017; Shahidi et al., 2019). Bioavailability depends on their physicochemical stability, complex formation, food interaction, gastrointestinal absorption, and hepatic and gut metabolism (Luca et al., 2019). Recent work has described how the gut microbiome contributes to aronia polyphenol metabolism and bioavailability (Istas et al, 2019). Significant inter-individual variability of polyphenol metabolism and bioavailability may explain the variable outcomes in human intervention studies and are summarized in Table 11. The following subsections focus on describing the metabolism and bioavailability of aronia berry polyphenols.

4.1. Anthocyanins

Anthocyanin structure and stability is pH-dependent. Upon consumption, the oral cavity pH is 5.6–7.9, which enhances anthocyanin hydrolysis. Given the short time period at this pH, only a fraction are hydrolyzed in the oral cavity (Braga et al., 2018). In the stomach, the acidic pH of 1.5–3.5 increases the proportion of anthocyanins as flavylium cations (Braga et al., 2018). At the small intestine, the pH of 6.7 to 7.4 favors the anthocyanin chalcone and quinoidal base formation, promoting hydrolysis to low molecular weight phenolics (Braga et al., 2018). A fraction of anthocyanins are directly absorbed in the stomach and small intestine. However, anthocyanins have low absorption in the small intestine, and the majority of them will be catabolized by gut microbiota (Denev et al., 2012). Upon absorption, anthocyanins and their metabolites undergo phase I and phase II metabolism in enterocytes and the liver (Denev et al., 2012). Phase I metabolites include phloroglucinaldehyde, 3,4-dihydroxybenzaldehyde, and hydroxybenzoic acid. Phase II metabolites are glucuronidated and methylated cyanidin. In the colon, gut microbiota hydrolyzes the anthocyanins into phenolic acids such as hippuric acid, phenylpropanoid acid, and ferulic acid (Wiczkowski et al., 2010; Denev et al., 2012). A human supplementation study utilizing ¹³C-cyanidin-3-glucoside reported 5.4% urinary excretion of anthocyanins and its metabolites (Czank et al., 2013). It would be expected that cyanidin-3-galactoside metabolism would be similar (Figure 1).

4.2. Proanthocyanidins

Proanthocyanidins have limited bioavailability because of their polymeric structures. Larger polymers are unable to translocate across the phospholipid bilayer of intestinal cells membrane, be transported by carrier proteins, and entirely dissolve into the aqueous phase of the small intestine (Luca et al., 2019). Moreover, human enzymes are not able to hydrolyze proanthocyanidins. Therefore, only monomers or dimers can be absorbed in the small intestine (Williamson and Clifford, 2017). The majority of proanthocyanidins reach the colon unchanged. A proportion of B-type proanthocyanidins are hydrolyzed by gut microbiota to valerolactones and phenolic acids (Appeldoorn et al., 2009). Thus, phenolic catabolites' rate and profile are highly dependent on the gut microbial composition (Luca et al., 2019). The remaining proanthocyanidins are excreted and in the feces (Neilson et al., 2016).

4.3. Quercetin

The metabolism and bioavailability of quercetin depends on its glycosylation (Kaşıkcı and Bağdatlıoğlu, 2016). Quercetin glucosides and galactosides are absorbed in the small intestine, whereas rutinosides are not. Rutin (quercetin-3-rutinoside) is deglycosylated by microbiota and then absorbed through passive diffusion (Luca et al., 2019). Quercetin glucosides and galactosides are hydrolyzed in the gut and liver, and aglycones mainly undergo Phase II metabolism. If the flavonols are not absorbed in the small intestine, they enter the gut, where the microbiota hydrolyzes the quercetin (Luca et al., 2019). The bacteria hydrolyze quercetin to low molecular polar metabolites (Santhakumar et al., 2018). Some of these compounds are 4-dihydroxyphenylacetic acid (DOPAC), 3-hydroxyphenyl acetic acid (3-OPAC), 3,4-dihydroxybenzoic acid (PCA), and vanillic acid (Ameida et al., 2018). The rate of formation of microbial metabolites depends in-part on nutrients presence (Rodriguez-Castaño et al., 2019). These low molecular compounds are absorbed in the large intestine and then metabolized in the liver (Atala et al., 2017). Quercetin metabolites are excreted through feces and urine, mainly as benzoic and hippuric acid (Luca et al., 2019).

4.4. Phenolic acids

The predominant phenolic acids in aronia, chlorogenic and neochlorogenic acids, require de-esterification of quinic acid prior to absorption (Denev et al., 2012). The bioavailability of these com-

Table 11. Summary of aronia berry juice and extract polypheno	ol bioavailability to plasma				
Source (dose)	Polyphenols	C _{max} (nM)	T _{max} (hours)	Participants	Reference
aronia extract (500 mg)	protocatechuic acid	32.4 ± 6.5	1.00 ± 0.00	6 adults	Xie et al., 2016
	hippuric acid	12100 ± 2700	5.33 ± 0.67		
	3-(4-hydroxyphenyl)propionic acid	1270 ± 810	6.33 ± 1.45		
	cyanidin-3- <i>O</i> -glucoside	122 ± 50	1.60 ± 0.24		
	peonidin-3-0-galactoside	150 ± 98	2 67 ± 0.67		
aronia extract (7.1 g)	total anthocyanins	96.1	2.8	3 male adults	Kay et al., 2005
	cyanidin 3-galactoside	23.4 ± 2.3	2.5 (2–3)		
	cyanidin glucuronide	14.5 ± 4.0	2.0 (2)		
	cyanidin 3-arabinoside	8.85 ± 0.50	3.5 (3–4)		
	peonidin 3-galactoside	3.76 ± 0.78	4.0 (4)		
	methylated cyanidin glucuronide	12.8 ± 0.4	2.5 (2–3)		
	methylated cyanidin glucuronide	32.8 ± 10.9	2.5 (2–3)		
aronia juice (0.8 mg of anthocyanins/g of body weight)	total anthocyanins	32.7	1.3 ± 0.1	13 adults	Wiczkowski et al., 2010
	cyanidin 3-galactoside	9.1	1		
	cyanidin-3- <i>O</i> -glucoside	1.0	1		
	cyanidin 3-arabinoside	2.2	1		
	cyanidin glucuronide	4.4	2		
	peonidin 3-galactoside	1.1	1		
	peonidin monoglucuronide	16	2		
	peonidin 3-arbinoside	1	0.5		
	total metabolites	21.3	2		
Data are means ± standard deviation, when available from primary data so	ource.				

pounds are low because esterase is limited in the small intestine. In the colon, bacteria release caffeic acid from chlorogenic acid (Heleno et al., 2015). Subsequently, absorbed phenolic acids undergo phase II metabolism in tissues prior to excretion.

4.5. Aronia polyphenol bioavailability

The maximum plasma concentration of dietary polyphenols in humans is in the nanomolar or low micromolar range (Istas et al., 2019). The highest level of polyphenols reported in plasma after aronia consumption has been 1.4 and 592 nmol/L, and appearing in the plasma between 0.5 and 4 h after consumption (Pojer et al., 2013). This wide range of absorption is partly because of differences in fruit composition, glycosylation, and anthocyanidin type. For example, a review on anthocyanins bioavailability concluded that galactosides were more bioavailable than arabinosides (Pojer et al., 2013). Additionally, the report compared aronia berry and elderberry extract bioavailability, containing 721 mg and 720 mg anthocyanidins, respectively (Pojer et al., 2013; Cao et al., 2001; Kay et al., 2005). After participants consumed both extracts, plasma anthocyanin concentrations were similar at 96.08 nM and 97.20 nM, respectively (Cao et al., 2001; Kay et al., 2005). In a double-blind, placebo-controlled study compared polyphenol metabolites in plasma after the consumption of whole aronia fruit or aronia extract (Istas et al., 2019). The total plasma polyphenol concentration was $30 \pm 156 \,\mu\text{M}$ and 14 \pm 106 μ M after 12 weeks of consuming extract and whole berries, respectively (Table 11). In addition, 20 metabolites were found in the volunteers consuming aronia extract, while only five metabolites were found after whole fruit consumption.

Another study evaluated the pharmacokinetics of anthocyanins and selected metabolites in adults' plasma (n = 6) that consumed 500 mg of aronia extract (Xie et al., 2016). Hippuric acid had the highest plasma concentration (0.87 to 3.5 µg/mL), followed by 3-(4-hydroxyphenyl)propionic acid (0.033 to 0.48 µg/mL), peonidin-3-O-galactoside (0.029 to 0.266 µg/mL), cyanidin-3-O-glucoside (0.014 to 0.180 µg/mL), and lastly protocatechuic acid (0.004 to 0.007 µg/mL) (Xie et al., 2016). A separate study found eight metabolites in the blood and urine consuming of 0.8mg of anthocyanins/ kg of body weight from aronia juice (Wiczkowski et al., 2010). The concentration of anthocyanins in the plasma was maximal at 1.3 hours after consumption, reaching 20.4 to 51.8 nmol/L. However, all of these studies had significant inter-individual variability in polyphenol pharmacokinetics. Larger studies are needed to characterize the importance of these variations to human health.

5. Health benefits of aronia berry consumption

Non-communicable diseases (cancer, diabetes, cardiovascular diseases, and depression) have an enormous social and economic toll worldwide (Centers for Disease Control and Prevention, 2020). Thus, multiple studies have focused on effective treatments to lower the risks of non-communicable diseases. Given the high polyphenol content of aronia berries, preliminary evidence indicates its preventive and therapeutic effects on non-communicable diseases. Here, we summarize recent preclinical (Table 12) and human intervention studies (Table 13) with aronia berry, juice, or extracts.

5.1. Cancer prevention

Recent studies have demonstrated anti-carcinogenic mechanisms of aronia polyphenols in vitro. Aronia extract prevents the growth, migration, and invasion of SK-Hep1 human liver cancer cells (Thi and Hwang, 2018). Liver cancer cell growth, adhesion, and migration were reduced by the aronia extract. Furthermore, aronia extract inhibited the expression of proteases involved in metastasis (MMP-2/9, MT-1 MMP). Another study isolated catechol from fermented aronia juice and characterized its inhibition of cancer stem cells (Choi et al., 2018). Catechol inhibited the formation of cancer stem cells and reduced the production of IL-6, which enhances cancer cells' survival. Additionally, catechol inhibited Stat3, a key transcription factor necessary for cancer stem cell formation. Thus, aronia components can inhibit multiple cancer mechanisms in vitro. Further studies that account for aronia polyphenol and bioavailability are needed to determine these studies' relevance to human health.

5.2. Diabetes

Preclinical and human studies have reported that aronia consumption may reduce insulin resistance. In rodents, aronia juice concentrate consumption increased plasma levels of adiponectin, the most abundant peptide secreted by key monomers that have an interrelationship between insulin resistance and inflammation (Baum et al., 2016). Aronia juice consumption also decreased intestinal glucosidase activity and increased DPP IV activity in diabetic mice (Yamane et al., 2016). An open-label trial of aronia juice for adults with type 2 diabetes reported improvement of glycemic control, with decreased fasting blood glucose and glycated hemoglobin (Milutinović et al., 2019). Thus, aronia consumption appears to be a promising treatment for diabetes. Further well-controlled human intervention studies on aronia berry are needed to increase the evidence base for its anti-diabetic activity.

5.3. Cardiovascular disease

Cardiovascular disease is a leading cause of death in the US (Centers for Disease Control and Prevention, 2020). Poor diets, low physical activity, excessive drinking, and smoking may increase cardiovascular disease risk. Increased blood pressure, adiposity, total and LDL cholesterol, and elevated inflammation and oxidative stress increase cardiovascular disease risk. Preclinical experiments have demonstrated aronia berry extract increases vasodilatory nitric oxide in cultured endothelial cells, and L-NAME induced hypertensive rats (Varela et al., 2015; Cebova et al., 2017). In rats, this increase is associated with increased nitric oxide synthase activity, reduced inflammation, and hypertension (Cebova et al., 2017). Consumption of aronia berry powder, extract, and juice also inhibits weight gain, lipid dysmetabolism, and inflammation in diet-induced obesity in mice (Bhaswant et al., 2017; Jeong and Kim, 2019; Yamane, Kozuka, Yamamoto, et al., 2016).

Aronia berry consumption improves biomarkers associated with cardiovascular disease risk in human intervention studies, but these changes depend on the participant populations. Aronia extract (116 mg) and powder (12 mg) consumption improved flow-mediated dilation, a marker of vascular function, in healthy men (Istas et al., 2019). Consumption of 300 mL/day of aronia juice and 3 g/day of aronia powder reduced systolic/diastolic blood pressure in adults with mildly elevated hypertension, but did not modulate serum lipids (Loo et al., 2016). Aronia extract reduced total and LDL cholesterol in healthy former smokers, and these changes were associated with increased urinary excretion of peonidin-3-galactoside, cyanidin-3-galactoside, and 3-(4-hydroxyphenyl)propionic acid (Xie et al., 2017). Open-label trials of aronia juice for adults with type 2



Figure 1. Overview of cyanidin-3-galactoside metabolism, adapted from de Ferrars et al. (2014). Cyanidin-3-galactoside undergoes methylation by the host to form peonidin-3-galactoside, or is hydrolyzed and glucoronidated. Anthyocyanins are subsequently hydrolyzed in host tissue or by the gut microbiota into phenolic acids. Phenolic catabolites can undergo enterohepatic circulation and are subjected to further host metabolism prior to excretion.

Table 12. Recen	nt preclinical studies on aronia berry hea	alth benefits		
Effect	Experimental model	Intervention	Outcomes	Reference
vascular health	spontaneously hypertensive rats	freeze-dried aronia berry in diet (10%), 25 days	blood pressure \downarrow , WG \downarrow , ACE activity in kidney \downarrow	Yamane et al., 2017
	L-NAME-induced hypertensive rats	aronia extract (57.90 mg/ kg/day), 21 days	BW \downarrow , heart weight \downarrow , TNF- $lpha$ \downarrow , IL-6 \downarrow , conjugated dienes \downarrow , nitric oxide synthase \uparrow ,	Cebova et al., 2017
	coronary artery endothelial cells	aronia extract (0.0001 – 100 mg/mL)	nitric oxide synthesis $ au$, endothelial nitric oxide synthase activation $ au$	Varela et al., 2015
brain health	adult rats	aronia juice (ad libitum), 30 days	anxiety like behaviors \downarrow	Tomić et al., 2016
	rat skeletal muscle cells, primary neuronal cells from prenatal rats, mice for <i>in vivo</i> MRI study	aonia berry extract in diet (50 mg/kg), applied to cells	skeletal muscle cells (hydrogen peroxide \downarrow) neuronal cells (inflammatory markers \downarrow) in vivo (BNDF \uparrow)	Kim et al., 2019
	BV2 cells	ethanolic aronia extract (0.03 – 2 mg/mL)	nitric oxide \downarrow , COX-2 \downarrow , IL-6 \downarrow , TNF- α \downarrow	Lee et al., 2018
	LPS-induced neural inflammation in mice	ethanolic aronia extract (50 mg/kg/day), 7 days	hippocampal damage \downarrow , neuroinflammaton \downarrow	
	HT22 mouse hippocampal cells	aronia extract (10, 100 µg/ mL); cyanidin-3-O-galactoside (1 or 10 µg/mL)	ROS	H. Y. Lee et al., 2017
	aged rats (24 mo.)	aronia juice (10 mL/kg, diluted 1:1 with water), 105 days	dentate gyrus \leftrightarrow , density of nerve fibers \uparrow , acetylcholinesterase activity \uparrow	Daskalova et al., 2019
	scopolamine-induced memory impaired mice	aronia extract (200, 400 mg/kg); cyanidin-3-O-galactoside (50 mg/kg)	memory \uparrow , acetylcholinesterase \downarrow , brain-derived neurotropic factor \uparrow , cAMP-response element binding protein \uparrow	H. Y. Lee et al., 2016
metabolic function	diabetic KK-Ay mice	aronia juice (ad libitum), 28 days	BW \downarrow , blood glucose \downarrow , white adipose tissue \downarrow , DPP IV activity \uparrow , glucosidase activity in upper small intestine \downarrow ,	Yamane et al., 2016
	high-sucrose, high-fat fed mice	aronia juice concentrate (1.44 g/kg diet), 12 weeks	High-sucrose diets: BW \downarrow , glucose \uparrow , insulin \downarrow , triglycerides \downarrow , adiponectin \uparrow , HOMA-IR \uparrow , HOMA-BCF \uparrow , FAS \downarrow , PPAR _{γ} \uparrow ; High-fat diets: BW \downarrow , glucose \downarrow , insulin \downarrow , triglycerides \downarrow , adiponectin \uparrow , HOMA-IR \uparrow , HOMA-BCF \uparrow , FAS \downarrow , PPAR _{γ} \uparrow	Baum et al., 2016
	high-carbohydrate, high-fat fed rats	aronia juice (50 mg/kg diet), 8 weeks	adiposity index \downarrow , total body mass \downarrow , systolic blood pressure \downarrow , glucose tolerance \uparrow , liver function \uparrow , cardiovascular function \uparrow , inflammatory in heart/ liver \downarrow	Bhaswant et al., 2017
	high-fructose, high-fat diet- induced dyslipidemic mice	aronia powder (1% in diet), 10 weeks	WG \downarrow , central obesity \downarrow , liver weight \downarrow , epididymal fat \downarrow , triglycerides \downarrow , cholesterol \downarrow , LDL \downarrow , insulin resistance \uparrow	Jeong and Kim, 2019
	high-fat fed mice	lyophilized aronia powder (10% in diet), 28 days	liver weight ↔, total lipid level ↓, cholesterol ↓, triglycerides ↓, LDL ↓, mild fibrosis ↓, FABP1 ↓, FABP4 ↓	Yamane, Kozuka, Yamamoto, et al., 2016

Effect	Experimental model	Intervention	Outcomes	Reference
cancer prevention	human liver cancer SK-Hep1 cells	aronia extract (0–200 µg/mL)	growth of cancer cells \downarrow , cell adhesion \downarrow , wound healing migration \uparrow , MMP-2/9 expression \downarrow	Thi and Hwang, 2018
	human breast cancer cell lines MCF-7 and MDA-MB-231	catechol isolated from fermented aronia juice	proliferation formation	Choi et al., 2018
anti- bacterial	foodborne pathogens and spoilage organisms	ethanol extracts of aronia berry	Gram-positive: B. cereus \downarrow , B. pumilus \downarrow , K. rhizophila \downarrow , L. monocytogenes \downarrow ; Gram- negative: C. jejuni \downarrow , S. Enteritidis \downarrow , E. coli \downarrow	Raudsepp et al., 2019
	foodborne pathogens	ethanol extract of aronia berry	Gram-positive: B. cereus ↓, S. aureus ↔ Gram- negative: S. entritidis ↔, C. sakazakii ↔	DH. Kim et al., 2018
colitis prevention	mice injected with syngeneic CD4+CD62L+ naïve T cells	lyophilized aronia berry (4.5% in diet), 7 weeks	colonic Treg 个,IL-17A+IL-10 ⁺ 个 and IL- 17A+IL-22 ⁺ 个,CD4 ⁺ cells 人,Verrucomicrobia <i></i> ↓; Bacteroidetes 个,Firmicutes 个,Proteobacteria ,colonic weight/length ratio	Pei et al., 2018
	male Wistar rats with TNBS-induced colitis	aronia fruit juice (2.5, 5, and 10 mL/kg, 14 days	TNBS, colon shortening, colone weight \uparrow , inhibited colonic weight/length ratio , lesion extension \downarrow , adhesion score \downarrow , wall thickening \downarrow	Valcheva-Kuzmanova et al., 2018
	Human umbilical vein endothelial cells stimulated with (TNF-α)	aronia berry extract	TNF-œ-induced moncyte/endothelia adhesion ↓, VCAM-1 ↓, ICAM-1 ↔, STAT3 ↓ IRF1 ↓, NF-ĸB ↔	lwashima et al., 2019
	mice injected with CD4+CD62L+ naïve T cells to induced colitis	lyophilized aronia berry (4.5% in diet), 5 weeks	colon weight/length ratio \downarrow , FDG uptake in spleen, liver and lungs \downarrow , TNF- α \downarrow , IFN- γ \downarrow , prevention of rGSH from \downarrow , GPx,activity \leftrightarrow , splenic mitochondrial H_2O_2 \downarrow	Pei et al., 2019
	Caco-2 cells with IC	whole Aronia berry powder (0.5–10 mg/mL)	Inhibited loss of TEER, prevented IC- induced barrier permeability, inhibited CLDN1, ZO-1 \downarrow	Valdez et al., 2020
Abbreviations: AC nase-2; DPP IV, dip Assessment of Ins oattractant proteir	E, angiotensin converting enzyme; ALDH, Aldel beptidyl peptidase 4; FAS, fatty acid synthase; Fl uin Resistance; IC, inflammatory cocktail; ICAN n; MMP 2/9, Matrix Metalloproteinase-2/9; NF hvaror of transcrintion. TEFR transcontendia	hyde dehydrogenases; BDNF, brain-derived neurot DG, 2-deoxy-2-[¹⁴ F]fluoro-d-glucose; GPx, glutathio M, intercellular adhesion molecule; IFN-Y, interferc Afsh nuclear lactor-kappa B; PPARy, perovisome pl	rophic factor; BW, body weight; cAMP, Cyclic adenosine monophosphate; ne peroxidase; HOMA-BCF, Homeostatic Model Assessment beta cell funct n gamma; IL-6, Interleukin 6; LDL, low-density lipoproteins; MDA, malon coliferator-activated receptor gamma; rGSH, reduced glutathione; ROS, re Activations and services and service wid, unsider activations; ACM - activa-	CLDN, claudin, <i>COX2, Cyclooxyge-</i> ion; HOMA-IR, Homeostatic Model- dialdehyde; MCP, monocyte chem- active oxygen species; STAT, signal

Composition, polyphenol bioavailability, and health benefits of aronia berry: a review

Participants	Design	Treatment	Outcomes	References
healthy men (n = 66)	double-blinded, placebo- controlled RCT	(poly)phenol-rich extract (116 mg, equivalent to 75 g berries), whole fruit powder (12 mg, equivalent to 10 g berries), 12 weeks	<i>Extract:</i> flow-mediated dilation \uparrow , plasma phenolic metabolites \uparrow , gut microbiota diversity \leftrightarrow , <i>Anaerostipes:</i> genus \uparrow , <i>Bacteroides</i> \uparrow ; <i>Powder:</i> flow-mediated dilation \leftrightarrow , plasma phenolic metabolites \uparrow , <i>Anaerostipes</i> genus \uparrow , <i>Bacteroides</i> \uparrow	lstas et al., 2019
adults with mildly elevated hypertension (n = 66)	single-blinded, placebo- controlled RCT	aronia juice (300 mL/d), oven-dried aronia powder (3 g/d)	blood pressure \downarrow , low-grade inflammation (TNF- α , IL-10) \downarrow , serum lipids \leftrightarrow , serum glucose \leftrightarrow	Loo et al., 2016
adolescents with metabolic syntdrome (n = 77)	open-label	aronia extract (3 × 100 mg/day), 8 weeks	total cholesterol \downarrow , LDL \downarrow , triacylglycerol \downarrow , HDL \uparrow , lipid peroxidation \downarrow , acetylcholinesterase \downarrow , oxidative stress \downarrow	Duchnowicz et al., 2018
former smokers (n = 49)	single-blinded, placebo- controlled RCT	aronia extract (500 mg/day), 12 weeks	Total cholesterol \downarrow , LDL \downarrow , LDL receptor proteins \downarrow , blood pressure \leftrightarrow , inflammation and oxidative stress biomarkers \leftrightarrow	Xie et al., 2017
healthy, adult handball players (n = 32)	double-blinded, placebo- controlled RCT	aronia juice (100 mL/d),	$\begin{array}{l} \textit{Men:} \mbox{lipid peroxidation } \downarrow, \mbox{oleic acid } \downarrow, \\ \alpha\mbox{-linolenic acid} \downarrow, \mbox{TAG } \downarrow, \mbox{TBARS } \downarrow; \mbox{Women:} \\ \mbox{lipid peroxidation} \leftrightarrow, \mbox{TAG } \uparrow \mbox{TBARS} \leftrightarrow \end{array}$	Petrovic et al., 2016
adults with type 2 diabetes (n = 35)	open-label	aronia juice (50 mL × 3/d), 3 months	glucose \downarrow , glycated hemoglobin \downarrow , total cholesterol \downarrow , LDL \downarrow , HDL \leftrightarrow , triglycerides \downarrow , high sensitive C-reactive protein \uparrow , blood urea \downarrow , creatinine \leftrightarrow , BMI \leftrightarrow , systolic blood pressure \leftrightarrow , diastolic blood pressure \leftrightarrow	Milutinović et al., 2019
healthy, adult runners (n = 10)	double-blinded, placebo- controlled RCT	aronia juice (200 mL with breakfast prior to running a half-marathon)	platelet-monocyte aggregates \downarrow , platelet-neutrophil aggregates \downarrow	Stevanović et al., 2019

Table 13. Recent human intervention studies with aronia berry interventions

Abbreviations: BMI, body mass index; HDL, high-density lipoproteins; IL-10, Interleukin-10; LDL, low-density lipoproteins; TAG, triglycerides; TBARS, thiobarbituric acid reactive substances; TNF- α , Tumor necrosis factor alpha.

diabetes and aronia extract for adolescents with metabolic syndrome have also been promising for modulating serum lipids and reducing oxidative stress and inflammation biomarkers. A systematic review and meta-analysis of literature concluded that among human interventions, aronia consumption leads to increases in HDL and diastolic blood pressure (Rahmani et al., 2019). Thus, further trials are needed to strengthen the evidence base for the specific populations that benefit from aronia berry consumption.

5.4. Modulation of gut microbiota

The microbiota contributes to modulation of the immune system and is now recognized to contribute to the progression of chronic disease (Festi et al., 2014). Furthermore, the gut microbiota contributes to the development of the metabolic syndrome and affects energy, lipid, and insulin metabolism. Furthermore, diets high in prebiotics that increase commensal microbes may positively change the microbiota composition and reduce inflammation related to metabolic syndrome. In aronia, sorbitol, fiber, and polyphenols may modulate microbial populations. Aronia polyphenols have direct anti-microbial activity against some food pathogens; however, the in vivo modulation of these microbes are not clear (Table 12). In mice, gut microbiota modulation precedes its antiinflammatory and immunomodulatory effects (Pei et al., 2019). In healthy adults, polyphenols' consumption equivalent to 75 g of aronia berry or berry powder equivalent to 10 g of aronia berry for 12 weeks did not affect microbial diversity (Istas et al., 2019). However, both groups had significant increases in Anaerostipes and Bacteroides, which are thought to be beneficial commensals. In a simulated human microbiota experiment, aronia juice increased firmicutes, proteobacteria and Akkermansia (Wu et al., 2018). Further studies are needed to characterize the specific effects of whole aronia berry and juice on gut microbiota and its relationship with inflammation outcomes and other health effects.

5.5. Neuroprotection

Experiments in rodents support the neuroprotective effects of aronia consumption (Table 12). When applied to cultured hippocampal and microglial cells, aronia reduces oxidative stress and inflammation (Lee et al., 2018; H. Y. Lee et al., 2017). In aged rats, aronia juice consumption increased hippocampal nerve fibers (Daskalova et al., 2019). Aronia supplementation may improve memory impairment and motor skills in rats (Daskalova et al., 2018).

Furthermore, aronia juice consumption reduced anxiety-like behaviors in adult rats (Tomić et al., 2016). While more research is necessary, these findings suggest aronia juice or extracts could benefit cognitive function and improve neural health.

5.6. Colitis

Colitis leads to dysmetabolism, loss of body weight, and micro-

biota changes (Ungaro et al., 2019; Pei et al., 2018). Anthocyanins inhibit intestinal inflammation, promote intestinal barrier function, and maybe protective against colitis (Valdez and Bolling, 2019). Aronia berry powder is protective against adoptive transfer and chemical-induced colitis in rodents. In adoptive transfer colitis, aronia consumption reduces colonic CD4+ cells and increases colonic Tregs and anti-inflammatory Th17 (Pei et al., 2018). These changes are linked to reduced oxidative stress and modulation of colonic antioxidant function (Pei et al., 2019). In the trinitrobenzene sulfonic acid (TNBS) colitis model, aronia juice consumption inhibits inflammation (Valcheva-Kuzmanova et al., 2018). The aronia berry juice decreased the lesion extension, adhesion score, and the colon's wall thickening score in colitic Wistar rats. Although these studies are promising, human intervention studies are necessary to determine the efficacy of aronia berry or its coproducts to inhibit inflammatory bowel diseases.

6. Conclusions

Aronia berries are one of the richest plant sources of anthocyanins and other bioactive polyphenols. However, these are presently underutilized in the diet because of its sourness and astringency. Aronia extract, juices, and pomace may be useful as functional ingredients, given their polyphenol content. Developing a better understanding of the contribution of bioactives to astringency could help create aronia products with improved palatability. Developing a more complete nutrient profile of aronia berry and juice will help improve nutrient databanks. Establishing the relationship of aronia plant genotype to berry quality will improve the production of berries with increased bioactive content. Emerging evidence describes the beneficial effects of aronia berry for prevention of diabetes, hypertension, cardiovascular disease, cancer, and colitis. However, future studies on the health benefits of aronia berry should utilize well-characterized aronia material, including description of the genotype, polyphenols, sorbitol, fiber, and micronutrient content.

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